# Package 'escape'

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Title Easy single cell analysis platform for enrichment

Version 2.4.0

**Description** A bridging R package to facilitate gene set enrichment analysis (GSEA) in the context of single-cell RNA sequencing. Using raw count information, Seurat objects, or SingleCell-Experiment format, users can perform and visualize ssGSEA, GSVA, AUCell, and UCell-based enrichment calculations across individual cells.

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densityEnrichment

Visualize the mean density ranking of genes across gene set

## **Description**

This function allows to the user to examine the mean ranking within the groups across the gene set. The visualization uses the density function to display the relative position and distribution of rank.

## Usage

```
densityEnrichment(
  input.data,
  gene.set.use = NULL,
  gene.sets = NULL,
  group.by = NULL,
  palette = "inferno"
)
```

## Arguments

```
input.data The single-cell object to use.

gene.set.use Selected individual gene set.

gene.sets The gene set library to use to extract the individual gene set information from.

group.by Categorical parameter to plot along the x.axis. If input is a single-cell object the default will be cluster.

palette Colors to use in visualization - input any hcl.pals.
```

#### Value

ggplot2 object mean rank gene density across groups

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## **Examples**

escape.gene.sets

Built-In Gene Sets for escape

# Description

A list of gene sets derived from Azizi, et al 2018 PMID: 29961579) relating to tumor immunity.

escape.matrix

Calculate gene set enrichment scores

## **Description**

This function allows users to input both the single-cell RNA-sequencing counts and output the enrichment scores as a matrix.

## Usage

```
escape.matrix(
  input.data,
  gene.sets = NULL,
  method = "ssGSEA",
  groups = 1000,
  min.size = 5,
  normalize = FALSE,
  make.positive = FALSE,
  BPPARAM = SerialParam(),
  ...
)
```

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## **Arguments**

input.data	The count matrix, Seurat, or Single-Cell Experiment object.
gene.sets	Gene sets can be a list, output from $\tt getGeneSets$ , or the built-in gene sets in the escape package $\tt escape.gene.sets$ .
method	Select the method to calculate enrichment, AUCell, GSVA, ssGSEA or UCell.
groups	The number of cells to separate the enrichment calculation.
min.size	Minimum number of gene necessary to perform the enrichment calculation
normalize	Whether to divide the enrichment score by the number of genes <b>TRUE</b> or report unnormalized <b>FALSE</b> .
make.positive	During normalization shift enrichment values to a positive range <b>TRUE</b> for downstream analysis or not <b>TRUE</b> (default). Will only be applied if <b>normalize</b> = <b>TRUE</b> .
BPPARAM	A BiocParallel::bpparam() object that for parallelization.
	pass arguments to AUCell GSVA, ssGSEA, or UCell call

## Value

matrix of enrichment scores

#### Author(s)

Nick Borcherding, Jared Andrews

## See Also

getGeneSets to collect gene sets.

## **Examples**

getGeneSets

Get a collection of gene sets from the msigdb

# Description

This function retrieves gene sets from msigdb and caches the downloaded object for future calls. It allows subsetting by main collection (library), subcollection, or specific gene sets, and only supports human ("Homo sapiens") and mouse ("Mus musculus").

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## Usage

```
getGeneSets(
  species = "Homo sapiens",
  library = NULL,
  subcategory = NULL,
  gene.sets = NULL,
  version = "7.4",
  id = "SYM"
)
```

# Arguments

species	The scientific name of the species of interest; only "Homo sapiens" or "Mus musculus" are supported.
library	A character vector of main collections (e.g. "H", "C5"). If provided, only gene sets in these collections are returned.
subcategory	A character vector specifying sub-collection abbreviations (e.g. "CGP", "CP:REACTOME") to further subset the gene sets.
gene.sets	A character vector of specific gene set names to select. This filter is applied after other subsetting.
version	The version of MSigDB to use (default "7.4").
id	The gene identifier type to use (default "SYM" for gene symbols).

#### Value

A named list of gene identifiers for each gene set.

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geyserEnrichment	Generate a ridge plot to examine enrichment distributions	

# Description

This function allows to the user to examine the distribution of enrichment across groups by generating a ridge plot.

# Usage

```
geyserEnrichment(
  input.data,
  assay = NULL,
  group.by = NULL,
  gene.set = NULL,
  color.by = "group",
  order.by = NULL,
  scale = FALSE,
  facet.by = NULL,
  palette = "inferno"
)
```

# Arguments

input.data	Enrichment output from escape.matrix or runEscape.
assay	Name of the assay to plot if data is a single-cell object.
group.by	Categorical parameter to plot along the x.axis. If input is a single-cell object the default will be cluster.
gene.set	Gene set to plot (on y-axis).
color.by	How the color palette applies to the graph - can be <b>"group"</b> for a categorical color palette based on the <b>group.by</b> parameter or use the <b>gene.set</b> name if wanting to apply a gradient palette.
order.by	Method to organize the x-axis: " <b>mean</b> " will arrange the x-axis by the mean of the gene.set, while " <b>group</b> " will arrange the x-axis by in alphanumerical order. Using <b>NULL</b> will not reorder the x-axis.
scale	Visualize raw values <b>FALSE</b> or Z-transform enrichment values <b>TRUE</b> .
facet.by	Variable to facet the plot into n distinct graphs.
palette	Colors to use in visualization - input any hcl.pals.

## Value

ggplot2 object with geyser-based distributions of selected gene.set

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#### **Examples**

heatmapEnrichment

Generate a heatmap to visualize enrichment values

#### **Description**

This function allows to the user to examine the heatmap with the mean enrichment values by group. The heatmap will have the gene sets as rows and columns will be the grouping variable.

#### Usage

```
heatmapEnrichment(
  input.data,
  assay = NULL,
  group.by = NULL,
  gene.set.use = "all",
  cluster.rows = FALSE,
  cluster.columns = FALSE,
  facet.by = NULL,
  scale = FALSE,
  summary.stat = "mean",
  palette = "inferno"
)
```

#### **Arguments**

input.data Enrichment output from escape.matrix or runEscape.

Name of the assay to plot if data is a single-cell object.

Categorical parameter to plot along the x.axis. If input is a single-cell object the default will be cluster.

Selected gene sets to visualize. If "all", the heatmap will be generated across all gene sets.

Cluster.rows Use Euclidean distance to order the row values.

Cluster.columns

Use Euclidean distance to order the column values.

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facet.by	Variable to facet the plot into n distinct graphs.
scale	Visualize raw values FALSE or Z-transform enrichment values TRUE.
summary.stat	Use 'median' or 'mean' values to display.
palette	Colors to use in visualization - input any hcl.pals.

## Value

ggplot2 object with heatmap of mean enrichment values

## **Examples**

pcaEnrichment

Visualize the PCA of enrichment values

# Description

This function allows to the user to examine the distribution of principal components run on the enrichment values.

#### Usage

```
pcaEnrichment(
  input.data,
  dimRed = NULL,
  x.axis = "PC1",
  y.axis = "PC2",
  facet.by = NULL,
  style = "point",
  add.percent.contribution = TRUE,
  display.factors = FALSE,
  number.of.factors = 10,
  palette = "inferno"
)
```

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## **Arguments**

input.data	PCA from performPCA.	
dimRed	Name of the dimensional reduction to plot if data is a single-cell object.	
x.axis	Component to plot on the x.axis.	
y.axis	Component set to plot on the y.axis.	
facet.by	Variable to facet the plot into n distinct graphs.	
style	Return a "hex" bin plot or a "point"-based plot.	
add.percent.contribution		
	Add the relative percent of contribution of the selected components to the axis labels.	
display.factors		
	Add an arrow overlay to show the direction and magnitude of individual gene sets on the PCA dimensions.	
number.of.factors		
	The number of gene.sets to display on the overlay.	
palette	Colors to use in visualization - input any hcl.pals.	

#### Value

ggplot2 object with PCA distribution

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#### **Description**

This function allows users to normalize the enrichment calculations by accounting for single-cell dropout and producing positive values for downstream differential enrichment analyses. Default calculation uses will scale the enrichment values by the number of genes present from the gene set and then use a natural log transformation. A positive range values is useful for several downstream analyses, like differential evaluation for log2-fold change, but will alter the original enrichment values.

## Usage

```
performNormalization(
    sc.data,
    enrichment.data = NULL,
    assay = "escape",
    gene.sets = NULL,
    make.positive = FALSE,
    scale.factor = NULL,
    groups = NULL
)
```

#### **Arguments**

sc.data Single-cell object or matrix used in the gene set enrichment calculation in escape.matrix

 $or \ {\tt runEscape}.$ 

enrichment.data

The enrichment results from escape.matrix or runEscape (optional)

assay Name of the assay to normalize if using a single-cell object

gene.sets The gene set library to use to extract the individual gene set information from

make.positive Shift enrichment values to a positive range TRUE for downstream analysis or

not TRUE (default).

scale.factor A vector to use for normalizing enrichment scores per cell.

groups the number of cells to calculate normalization on at once. chunks matrix into

groups sized chunks. Useful in case of memory issues.

#### Value

Single-cell object or matrix of normalized enrichment scores

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```
assay = "escape",
gene.sets = GS)
```

performPCA

Perform Principal Component Analysis on Enrichment Data

## **Description**

This function allows users to calculate the principal components for the gene set enrichment values. For single-cell data, the PCA will be stored with the dimensional reductions. If a matrix is used as input, the output is a list for further plotting. Alternatively, users can use functions for PCA calculations based on their desired workflow in lieu of using performPCA, but will not be compatible with downstream pcaEnrichment visualization.

## Usage

```
performPCA(
  input.data,
  assay = NULL,
  scale = TRUE,
  n.dim = 1:10,
  reduction.name = "escape.PCA",
  reduction.key = "PCA"
)
```

#### **Arguments**

input.data Enrichment output from escape.matrix or runEscape.

assay Name of the assay to plot if data is a single-cell object.

scale Standardize the enrichment value (TRUE) or not (FALSE)

n.dim The number of components to calculate.

reduction.name Name of the reduced dimensions object to add if data is a single-cell object.

reduction.key Name of the key to use with the components.

## Value

single-cell object or list with PCA components to plot.

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ridgeEnrichment

Visualize enrichment results with a ridge plot

# Description

This function allows to the user to examine the distribution of enrichment across groups by generating a ridge plot.

# Usage

```
ridgeEnrichment(
  input.data,
  assay = NULL,
  group.by = NULL,
  gene.set = NULL,
  color.by = "group",
  order.by = NULL,
  scale = FALSE,
  facet.by = NULL,
  add.rug = FALSE,
  palette = "inferno"
)
```

# Arguments

input.data	Enrichment output from escape.matrix or runEscape.
assay	Name of the assay to plot if data is a single-cell object.
group.by	Categorical parameter to plot along the x.axis. If input is a single-cell object the default will be cluster.
gene.set	Gene set to plot (on y-axis).
color.by	How the color palette applies to the graph - can be "group" for a categorical color palette based on the group.by parameter or use the gene.set name if wanting to apply a gradient palette.
order.by	Method to organize the x-axis: <b>"mean"</b> will arrange the x-axis by the mean of the gene.set, while <b>"group"</b> will arrange the x-axis by in alphanumerical order. Using <b>NULL</b> will not reorder the x-axis.
scale	Visualize raw values FALSE or Z-transform enrichment values TRUE.
facet.by	Variable to facet the plot into n distinct graphs.
add.rug	Add visualization of the discrete cells along the ridge plot (TRUE).
palette	Colors to use in visualization - input any hcl.pals.

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#### Value

ggplot2 object with ridge-based distributions of selected gene.set

## **Examples**

runEscape

Enrichment calculation for single-cell workflows

## **Description**

Run the escape-based gene-set enrichment calculation with Seurat or SingleCellExperiment pipelines

## Usage

```
runEscape(
  input.data,
  gene.sets = NULL,
  method = "ssGSEA",
  groups = 1000,
  min.size = 5,
  normalize = FALSE,
  make.positive = FALSE,
  new.assay.name = "escape",
  BPPARAM = SerialParam(),
  ...
)
```

# Arguments

input.data The count matrix, Seurat, or Single-Cell Experiment object.

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gene.sets	Gene sets can be a list, output from getGeneSets, or the built-in gene sets in the escape package escape.gene.sets.
method	Select the method to calculate enrichment, AUCell, GSVA, ssGSEA or UCell.
groups	The number of cells to separate the enrichment calculation.
min.size	Minimum number of gene necessary to perform the enrichment calculation
normalize	Whether to divide the enrichment score by the number of genes $TRUE$ or report unnormalized $FALSE$ .
make.positive	During normalization shift enrichment values to a positive range <b>TRUE</b> for downstream analysis or not <b>TRUE</b> (default). Will only be applied if <b>normalize</b> = <b>TRUE</b> .
new.assay.name	The new name of the assay to append to the single-cell object containing the enrichment scores.
BPPARAM	A BiocParallel::bpparam() object that for parallelization.
	pass arguments to AUCell GSVA, ssGSEA or UCell call

#### Value

Seurat or Single-Cell Experiment object with escape enrichment scores in the assay slot.

#### **Examples**

scatterEnrichment

Generate a density-based scatter plot

#### **Description**

This function allows to the user to examine the distribution of 2 gene sets along the x.axis and y.axis. The color gradient is generated using the a density estimate. See ggpointdensity) for more information.

# Usage

```
scatterEnrichment(
  input.data,
  assay = NULL,
  x.axis = NULL,
  y.axis = NULL,
```

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```
scale = FALSE,
facet.by = NULL,
style = "point",
palette = "inferno"
)
```

## **Arguments**

input.data	Enrichment output from escape.matrix or runEscape.
assay	Name of the assay to plot if data is a single-cell object.
x.axis	Gene set to plot on the x.axis.
y.axis	Gene set to plot on the y.axis. <b>group.by</b> parameter or use the <b>gene.set</b> name if wanting to apply a gradient palette.
scale	Visualize raw values FALSE or Z-transform enrichment values TRUE.
facet.by	Variable to facet the plot into n distinct graphs.
style	Return a "hex" bin plot or a "point"-based plot.
palette	Colors to use in visualization - input any hcl.pals.

#### Value

ggplot2 object with a scatter plot of selected gene.sets

# **Examples**

splitEnrichment

Visualize enrichment results with a split violin plot

## **Description**

This function allows to the user to examine the distribution of enrichment across groups by generating a split violin plot.

splitEnrichment

## Usage

```
splitEnrichment(
  input.data,
  assay = NULL,
  split.by = NULL,
  group.by = NULL,
  gene.set = NULL,
  order.by = NULL,
  facet.by = NULL,
  scale = TRUE,
  palette = "inferno"
)
```

## **Arguments**

input.data	Enrichment output from escape.matrix or runEscape.
assay	Name of the assay to plot if data is a single-cell object.
split.by	Variable to form the split violin, must have 2 levels.
group.by	Categorical parameter to plot along the x.axis. If input is a single-cell object the default will be cluster.
gene.set	Gene set to plot (on y-axis).
order.by	Method to organize the x-axis - "mean" will arrange the x-axis by the mean of the gene.set, while "group" will arrange the x-axis by in alphanumerical order. Using NULL will not reorder the x-axis.
facet.by	Variable to facet the plot into n distinct graphs.
scale	Visualize raw values <b>FALSE</b> or Z-transform enrichment values <b>TRUE</b> .
palette	Colors to use in visualization - input any hcl.pals.

#### Value

ggplot2 object violin-based distributions of selected gene.set

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