Statistical Methods and Software for the Analysis of DNA Microarray Experiments

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Outline

- Introduction to the biology and technology of DNA microarrays
- Overview of the Bioconductor project
- Annotation
- Visualization
- Pre-processing: spotted and Affymetrix arrays
- Differential gene expression
- Software demo

Acknowledgments

Bioconductor core team

- **Ben Bolstad**, Biostatistics, UC Berkeley
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- **Robert Gentleman**, Biostatistics, Harvard
- **Jeff Gentry**, Dana-Farber Cancer Institute
- **Yee Hwa (Jean) Yang**, Biostatistics, UCSF
- **Jianhua (John) Zhang**, Dana-Farber Cancer Institute

References

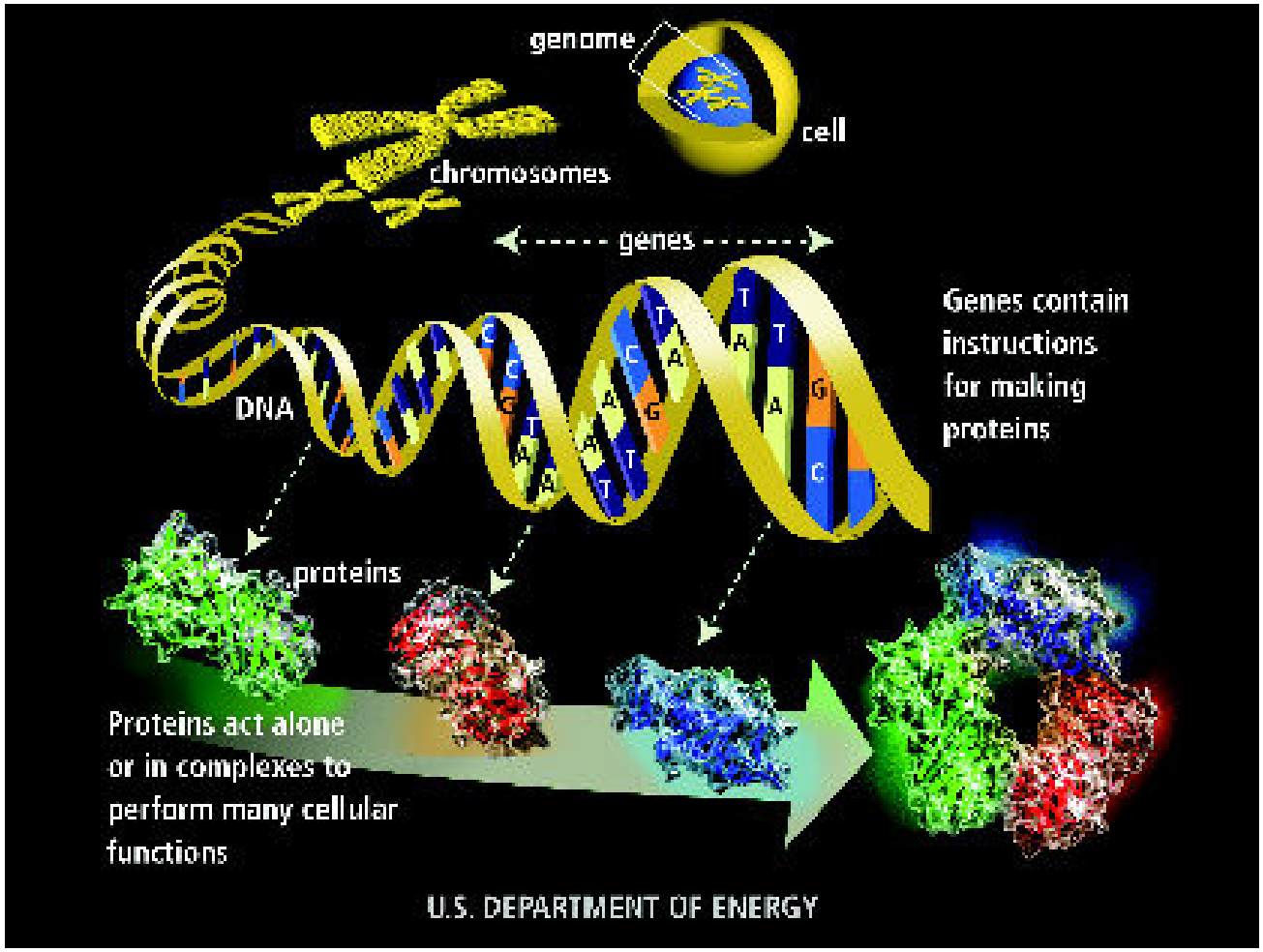
- **Personal webpages**

  www.stat.berkeley.edu/~sandrine

  biosun01.biostat.jhsph.edu/~ririzarr

  articles and talks on: image analysis; normalization; identification of differentially expressed genes; cluster analysis; classification.
- **Bioconductor** www.bioconductor.org
  - software, data, and documentation (vignettes);
  - training materials from short courses;
  - mailing list.
- **R** www.r-project.org
  - software; documentation; RNews.

From chromosomes to proteins



www.ornl.gov/hgmis/graphics/slides/images1.html

Cells

- Cells: the fundamental working units of every living organism.

- Metazoa: multicellular organisms.
  E.g. humans: trillions of cells.
- Protozoa: unicellular organisms.
  E.g. yeast, bacteria.

Cells

- Each cell contains a complete copy of an organism's genome, or blueprint for all cellular structures and activities.

- Cells are of many different types (e.g. blood, skin, nerve cells), but all can be traced back to a single cell, the fertilized egg.

The genome

- The human genome is distributed along 23 pairs of chromosomes
  - 22 autosomal pairs;
  - the sex chromosome pair, **XX** for females and **XY** for males.

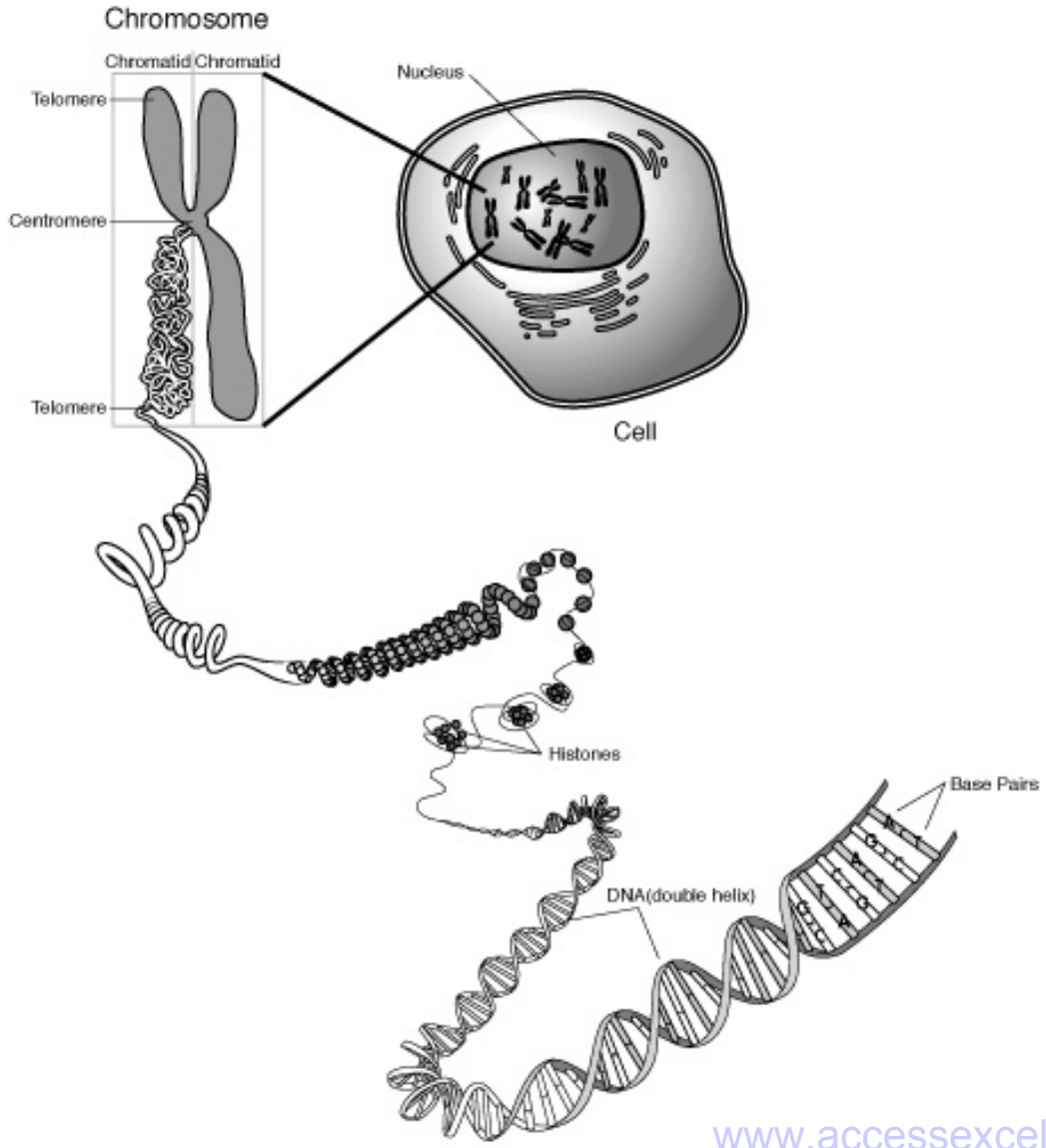
- In each pair, one chromosome is paternally inherited, the other maternally inherited (cf. meiosis).

The genome

- Chromosomes are made of compressed and entwined DNA.

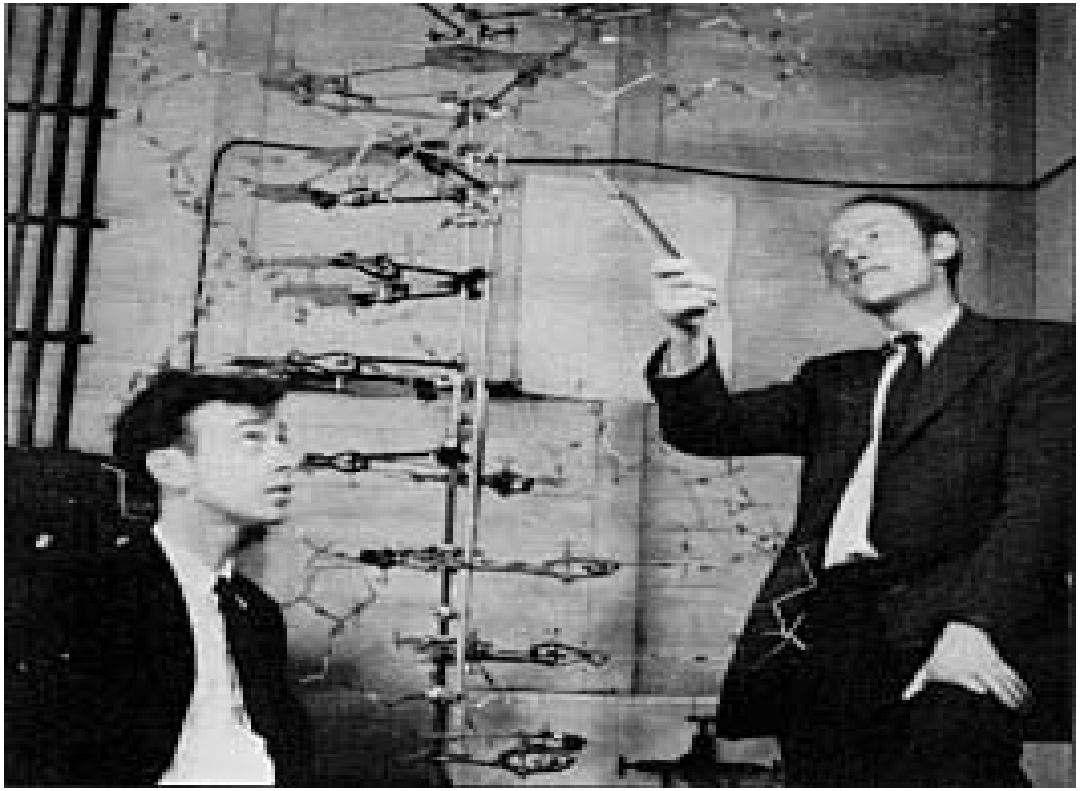
- A (protein-coding) gene is a segment of chromosomal DNA that directs the synthesis of a protein.

Chromosomes and DNA



www.accessexcellence.com/AB/GG/

DNA



"We wish to suggest a structure for the salt of deoxyribose nucleic acid (D.N.A.). This structure has novel features which are of considerable biological interest."

J.D. Watson & F. H. C. Crick. (1953). Molecular structure of Nucleic Acids. *Nature.* **171**: 737-738.

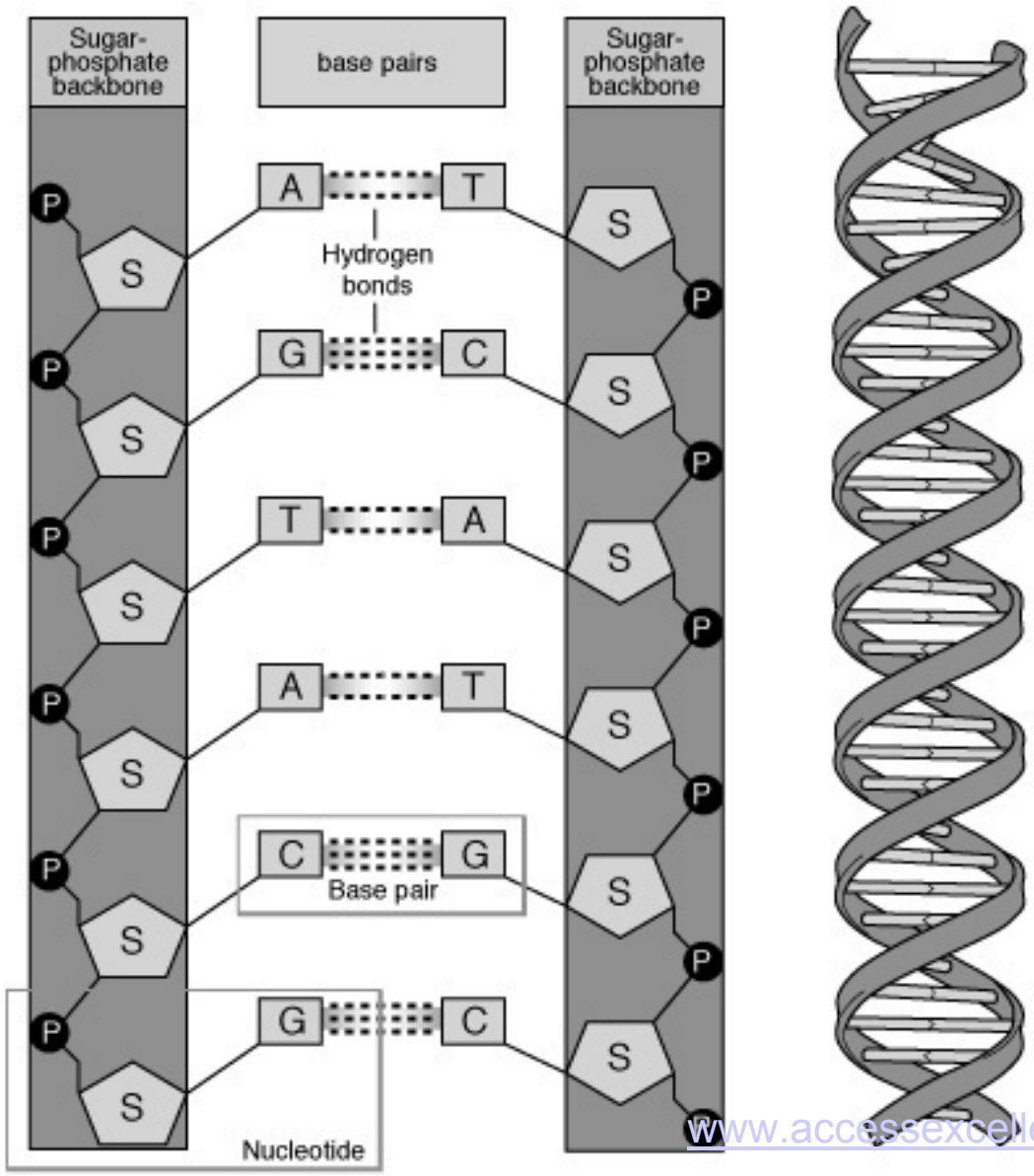
DNA

- A deoxyribonucleic acid or DNA molecule is a double-stranded polymer composed of four basic molecular units called nucleotides.
- Each nucleotide comprises
  - a phosphate group;
  - a deoxyribose sugar;
  - one of four nitrogen bases:
    - purines: adenine (A) and guanine (G),
    - pyrimidines: cytosine (C) and thymine (T).

DNA

- Base-pairing occurs according to the following rule:
  - **C pairs with G**,
  - **A pairs with T**.
- The two chains are held together by hydrogen bonds between nitrogen bases.

DNA



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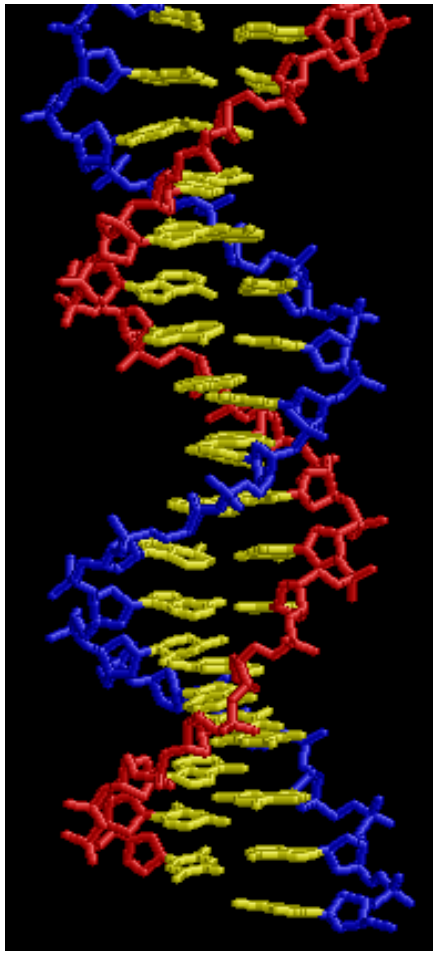
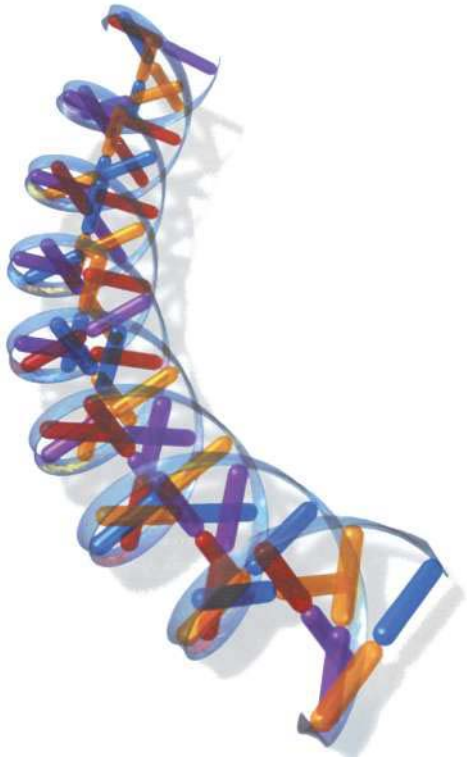
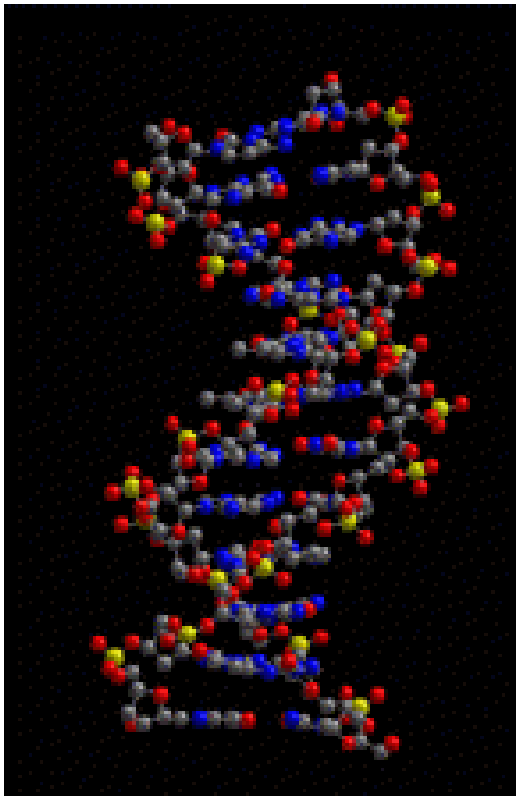
DNA

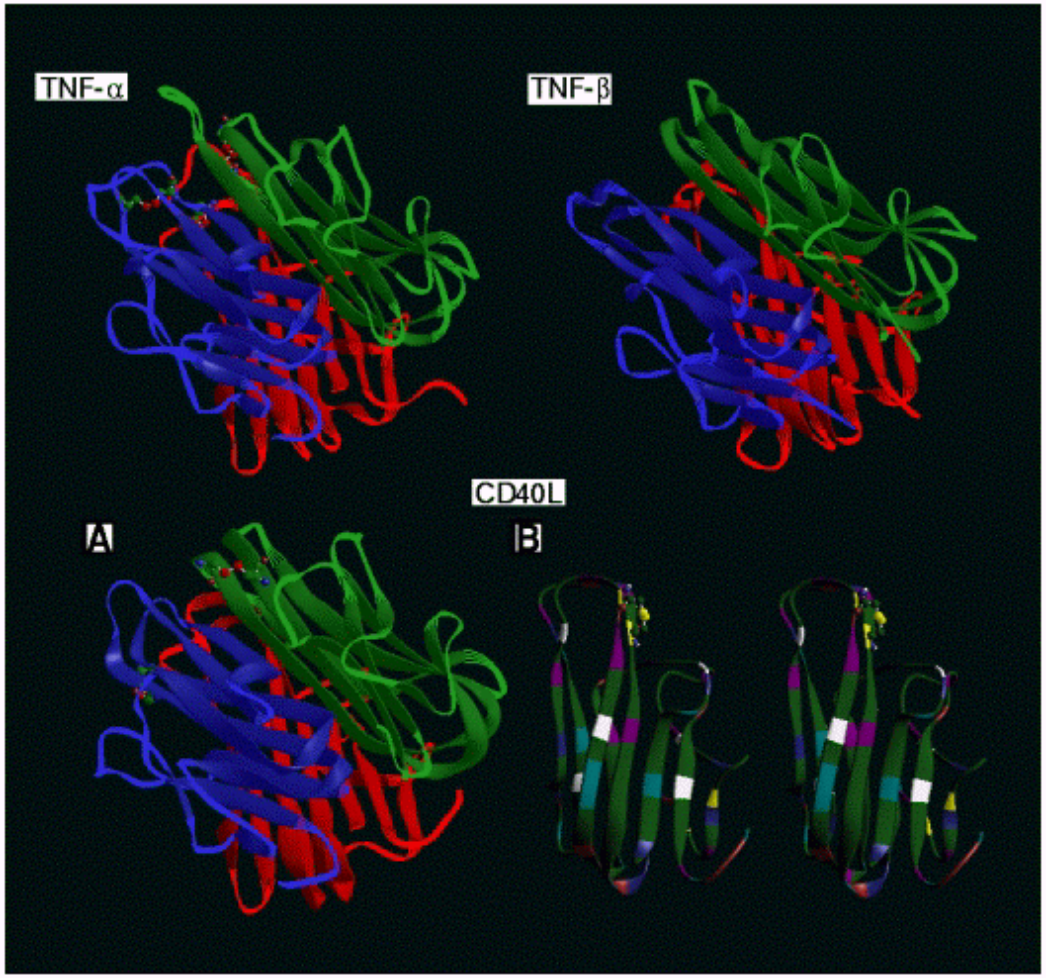






academy.d20.co.edu/kadets/lundberg/dnapic.html

Proteins

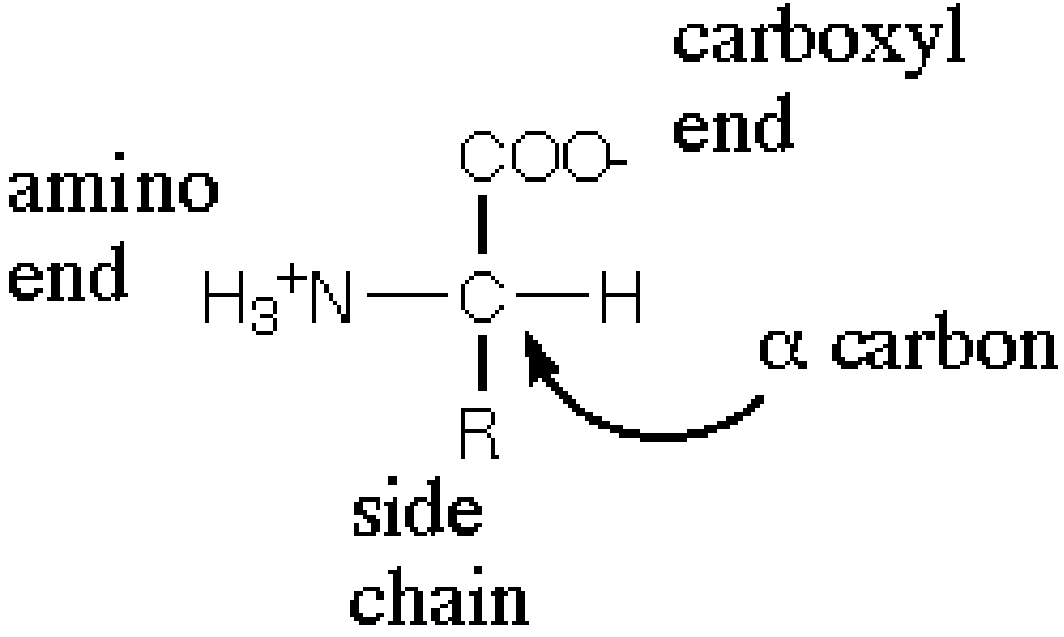


www.biochem.szote.u-szeged.hu/astrojan/protein1.htm

Proteins

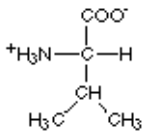
- Proteins: large molecules composed of one or more chains of amino acids, polypeptides.
- Amino acids: class of 20 different organic compounds containing a basic amino group ($-NH_2$) and an acidic carboxyl group (-COOH).
- The order of the amino acids is determined by the base sequence of nucleotides in the gene coding for the protein.
- E.g. hormones, enzymes, antibodies.

Amino acids

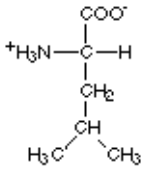


Amino acids

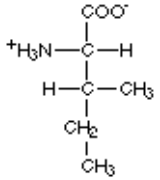
Amino acids with hydrophobic side groups



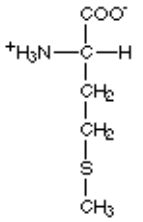
Valine
(val)



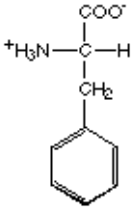
Leucine
(leu)



Isoleucine
(ile)

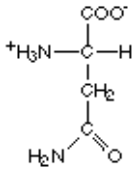


Methionine
(met)

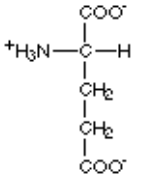


Phenylalanine
(phe)

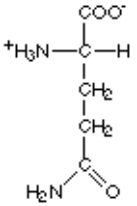
Amino acids with hydrophilic side groups



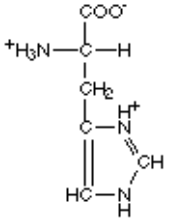
Asparagine
(asn)



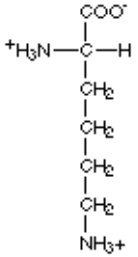
Glutamic acid
(glu)



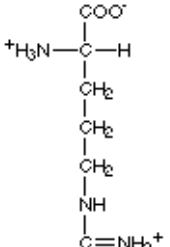
Glutamine
(gln)



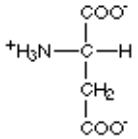
Histidine
(his)



Lysine
(lys)

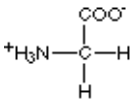


Arginine
(arg)

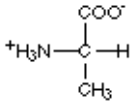


Aspartic acid
(asp)

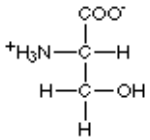
Amino acids that are in between



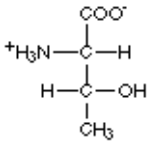
Glycine
(gly)



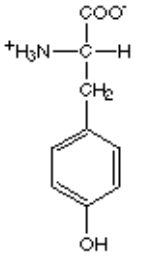
Alanine
(ala)



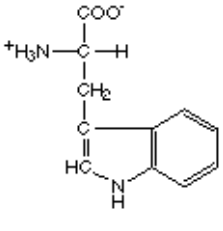
Serine
(ser)



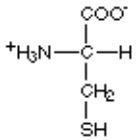
Threonine
(thr)



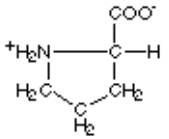
Tyrosine
(tyr)



Tryptophan
(trp)

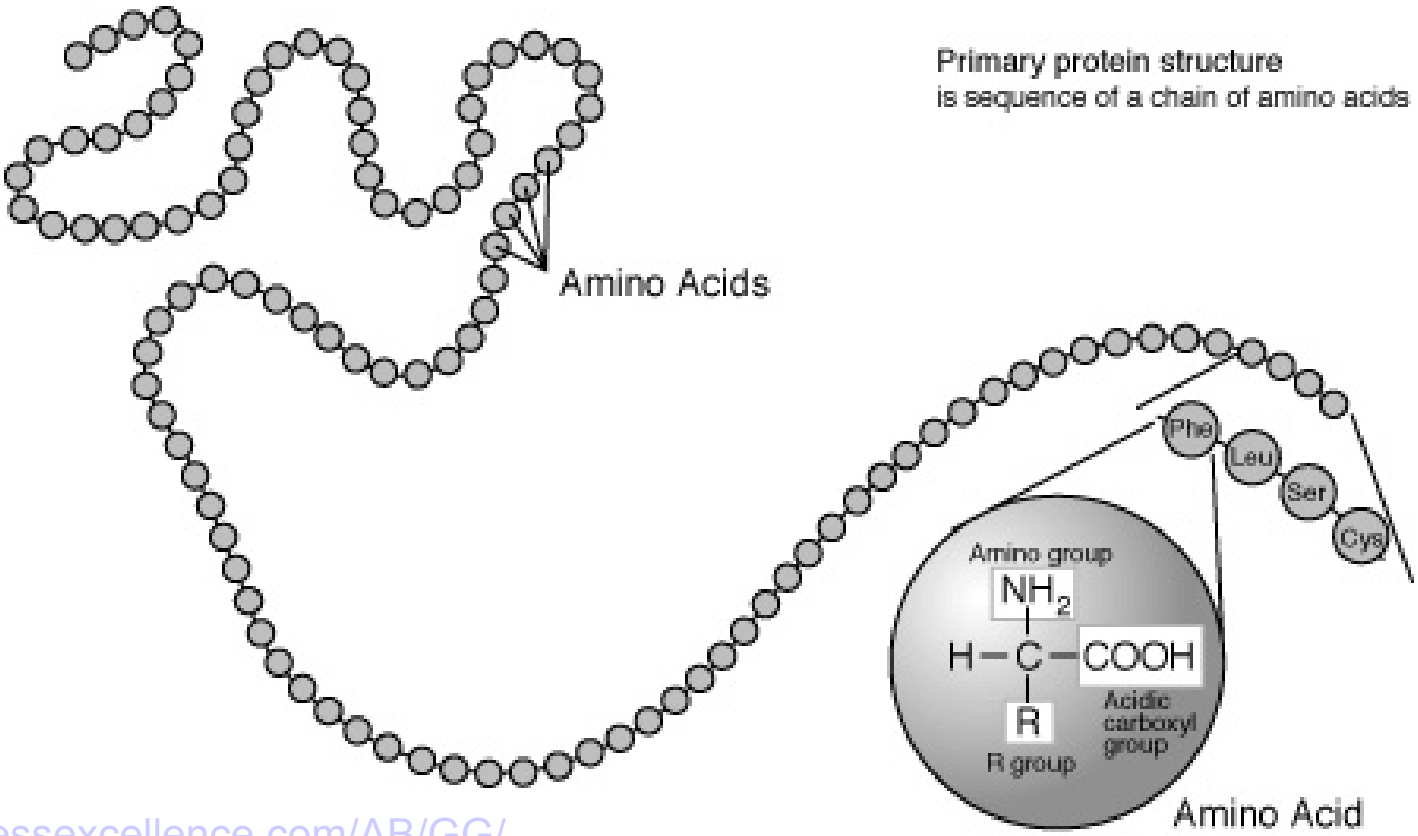


Cysteine
(cys)



Proline
(pro)

Proteins



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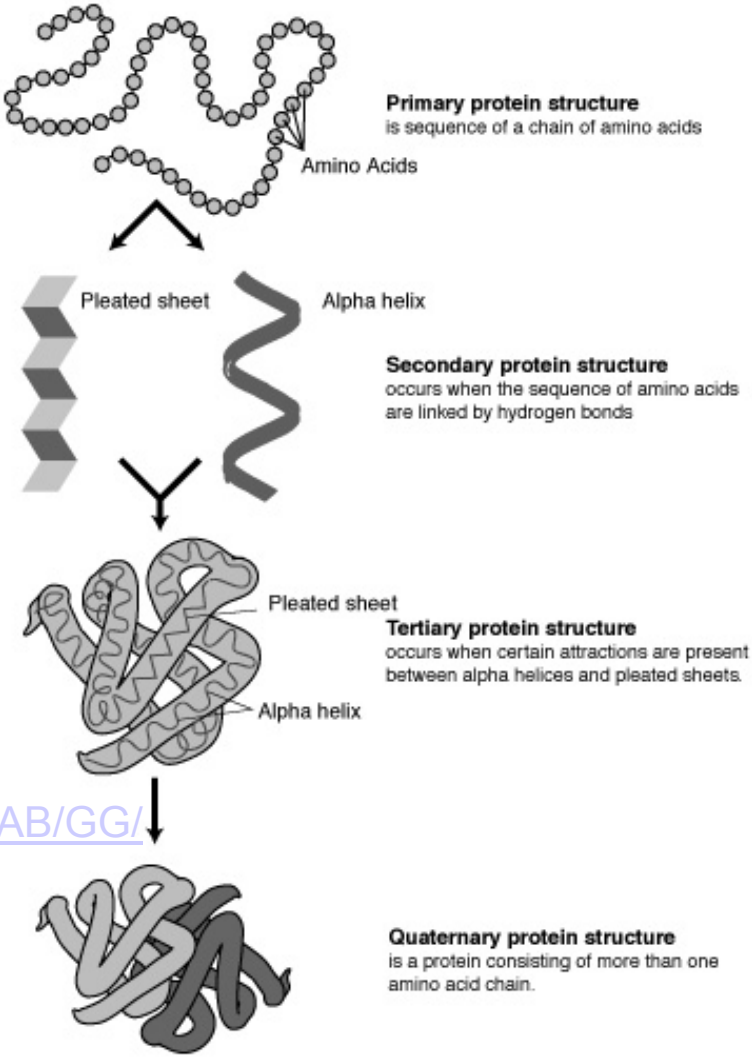
Proteins



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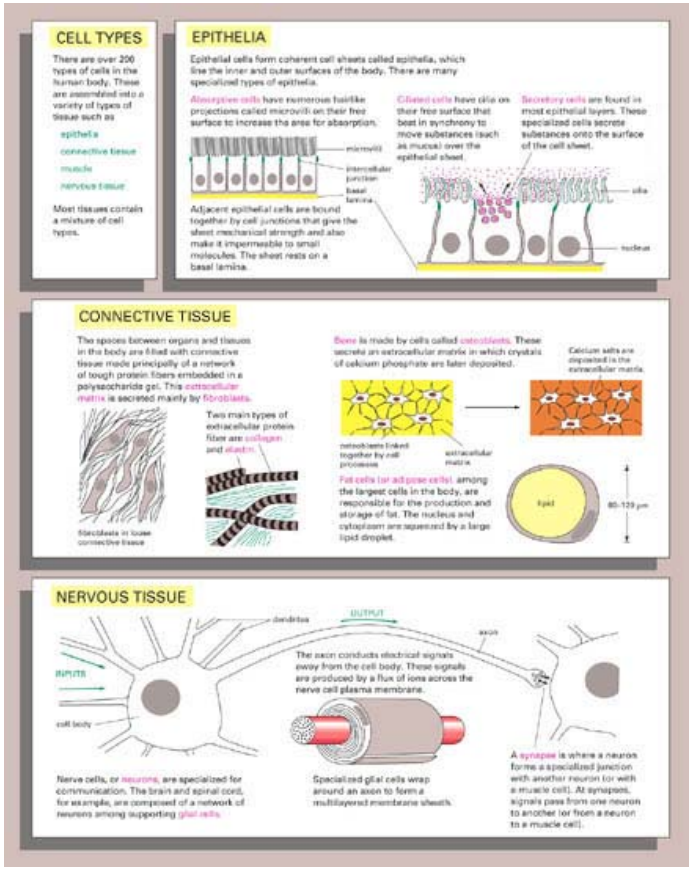
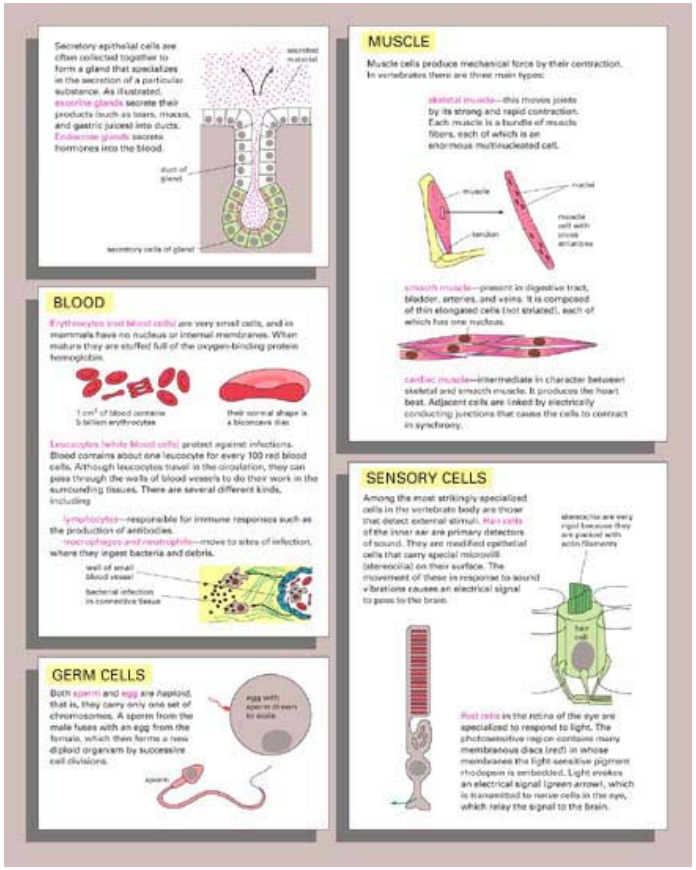
Cell types





www.accessexcellence.com/AB/GG/

Differential expression

- Each cell contains a complete copy of the organism's genome.
- Cells are of many different types and states
  E.g. blood, nerve, and skin cells, dividing cells, cancerous cells, etc.
- What makes the cells different?
- Differential gene expression, i.e., when, where, and how much each gene is expressed.
- On average, 40% of our genes are expressed at any given time.

Central dogma

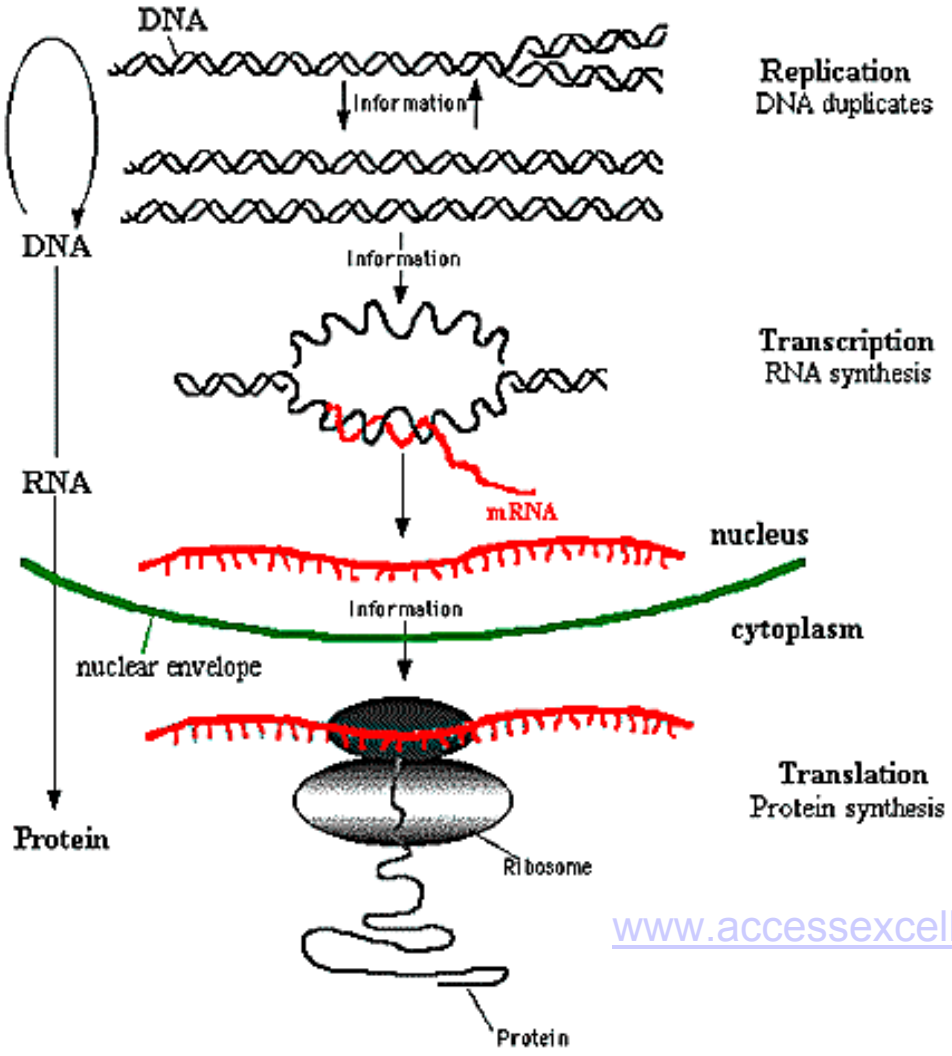
The expression of the genetic information stored in the DNA molecule occurs in two stages:

- (i) transcription, during which DNA is transcribed into mRNA;
- (ii) translation, during which mRNA is translated to produce a protein.

DNA → mRNA → protein

Other important aspects of regulation: methylation, alternative splicing, etc.

Central dogma



www.accessexcellence.com/AB/GG/

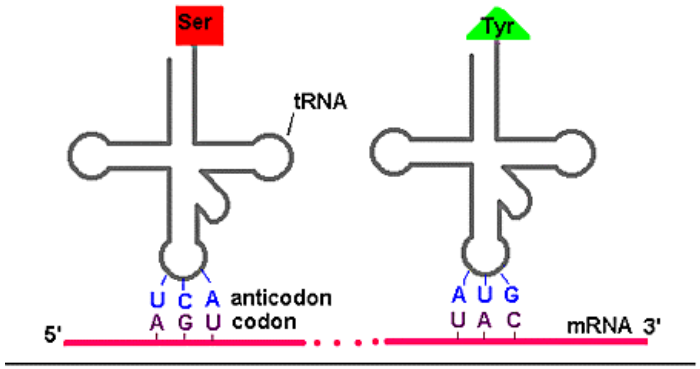
RNA

- A ribonucleic acid or RNA molecule is a nucleic acid similar to DNA, but
  - single-stranded;
  - ribose sugar rather than deoxyribose sugar;
  - uracil (U) replaces thymine (T) as one of the bases.
- RNA plays an important role in protein synthesis and other chemical activities of the cell.
- Several classes of RNA molecules, including messenger RNA (mRNA), transfer RNA (tRNA), ribosomal RNA (rRNA), and other small RNAs.

The genetic code

- DNA: sequence of four different nucleotides.
- Proteins: sequence of twenty different amino acids.
- The correspondence between DNA's four-letter alphabet and a protein's twenty-letter alphabet is specified by the genetic code, which relates nucleotide triplets or codons to amino acids.

The genetic code



2nd base in codon

1st base in codon	U	C	A	G	3rd base in codon
U	Phe	Ser	Tyr	Cys	U
	Phe	Ser	Tyr	Cys	C
	Leu	Ser	STOP	STOP	A
	Leu	Ser	STOP	Trp	G
C	Leu	Pro	His	Arg	U
	Leu	Pro	His	Arg	C
	Leu	Pro	Gln	Arg	A
	Leu	Pro	Gln	Arg	G
A	Ile	Thr	Asn	Ser	U
	Ile	Thr	Asn	Ser	C
	Ile	Thr	Lys	Arg	A
	Met	Thr	Lys	Arg	G
G	Val	Ala	Asp	Gly	U
	Val	Ala	Asp	Gly	C
	Val	Ala	Glu	Gly	A
	Val	Ala	Glu	Gly	G

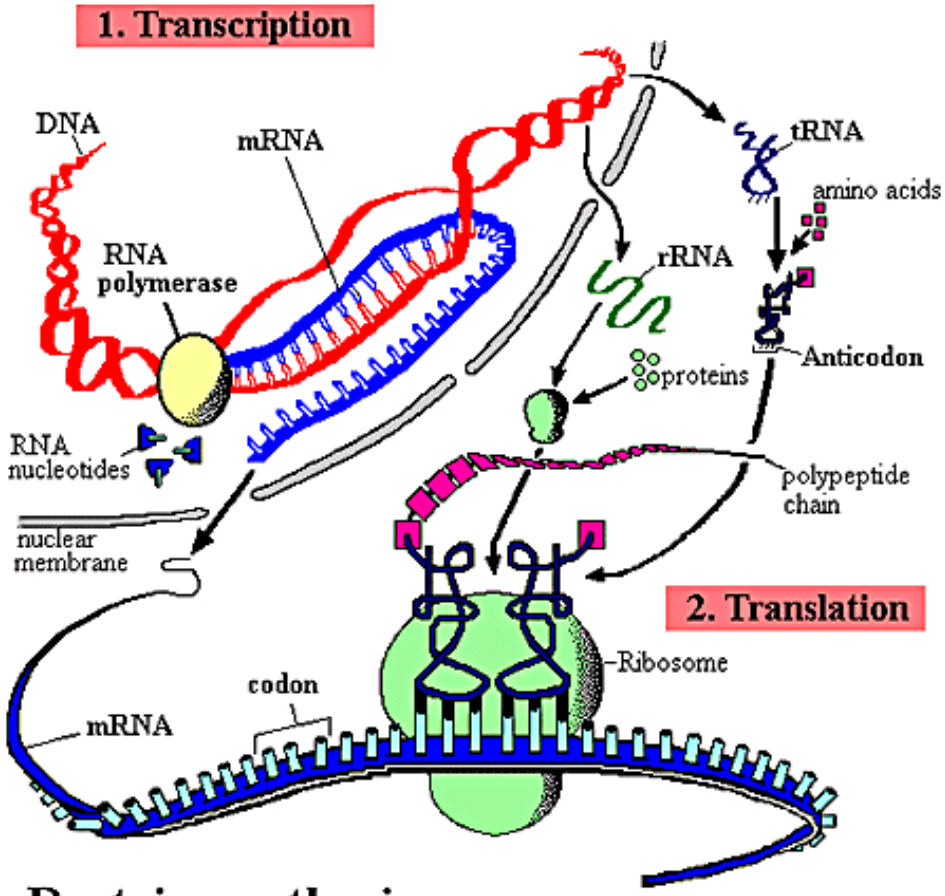
The Genetic Code

www.accessexcellence.com/AB/GG/

Start codon: initiation of translation (AUG, Met).
Stop codons: termination of translation.

Mapping between codons and amino acids is **many-to-one**: 64 codons but only 20 a.a..
Third base in codon is often redundant,
e.g., stop codons.

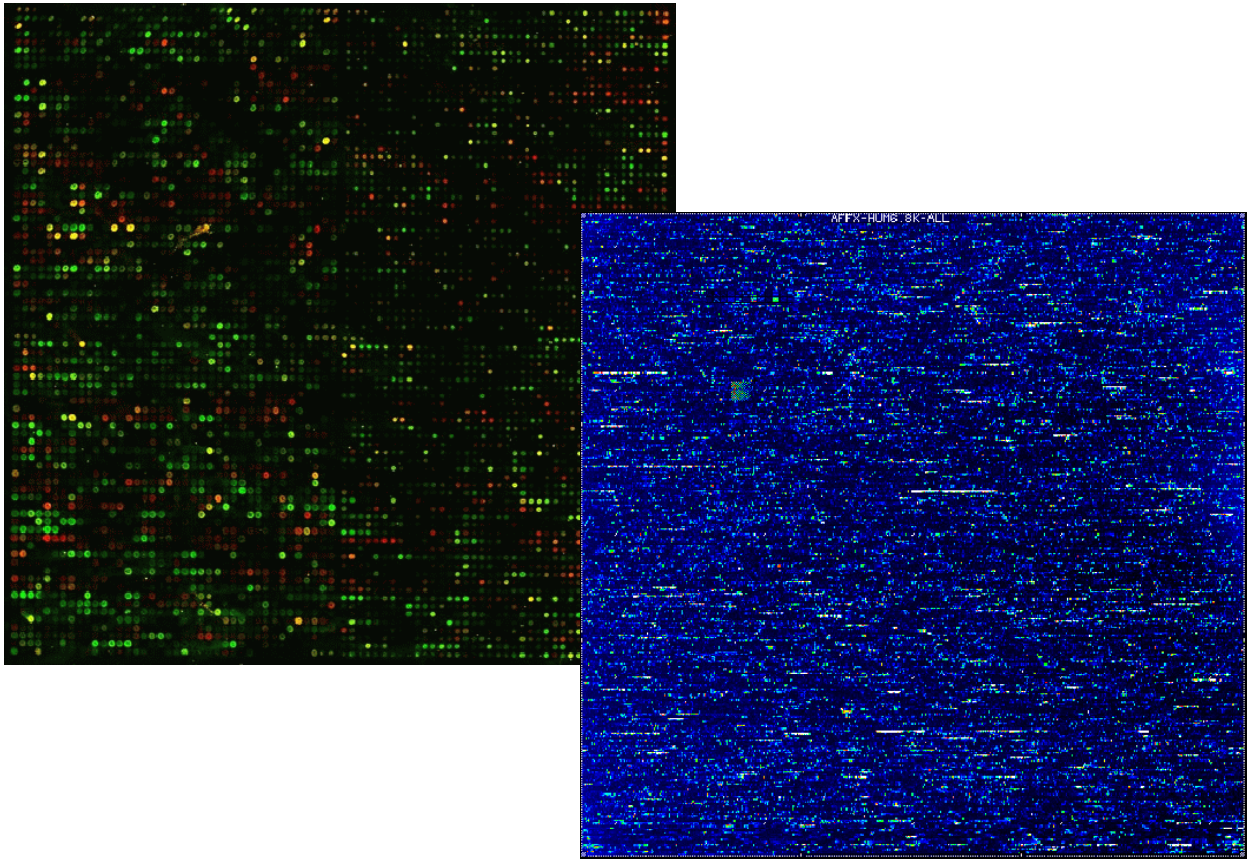
Protein synthesis



Functional genomics

- The various genome projects have yielded the complete DNA sequences of many organisms.
  - E.g. human, mouse, yeast, fruitfly, etc.
  - Human: 3 billion base-pairs, 30-40 thousand genes.
- Challenge: **go from sequence to function**, i.e., define the role of each gene and understand how the genome functions as a whole.

DNA microarrays



DNA microarrays

- Basic principles
- Spotted DNA microarrays
- Affymetrix oligonucleotide chips

DNA microarrays

- DNA microarray experiments are high-throughput biological assays for measuring the abundance of DNA or RNA sequences in different types of cell samples for thousands of sequences simultaneously.
- DNA microarray experiments exploit the availability of sequence data to get information on gene expression in different types of cells.

DNA microarrays

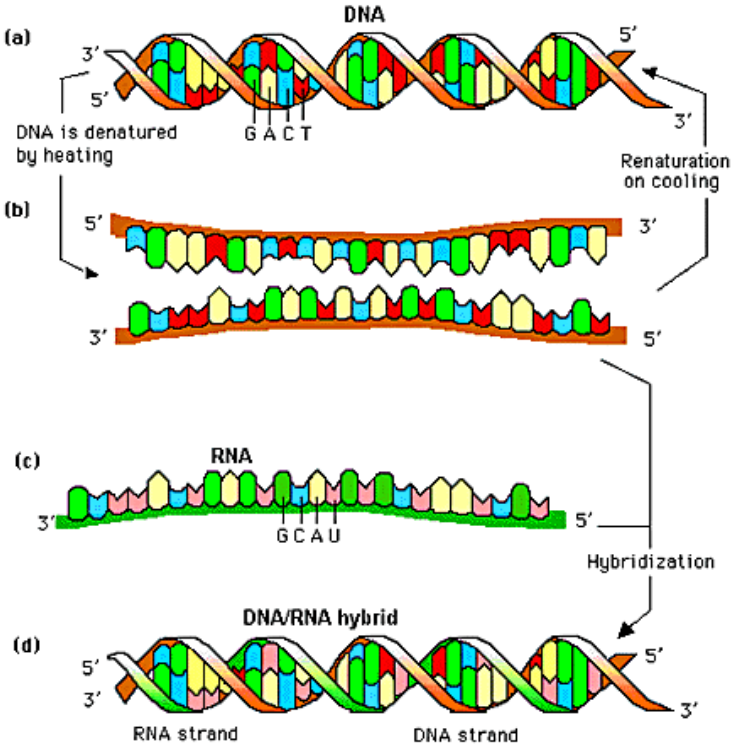
- DNA microarrays rely on the hybridization properties of nucleic acids to monitor DNA or RNA abundance on a genomic scale in different types of cells.
- The ancestor of cDNA microarrays: the Northern blot.

Hybridization

- Hybridization refers to the annealing of two nucleic acid strands following the base-pairing rules.

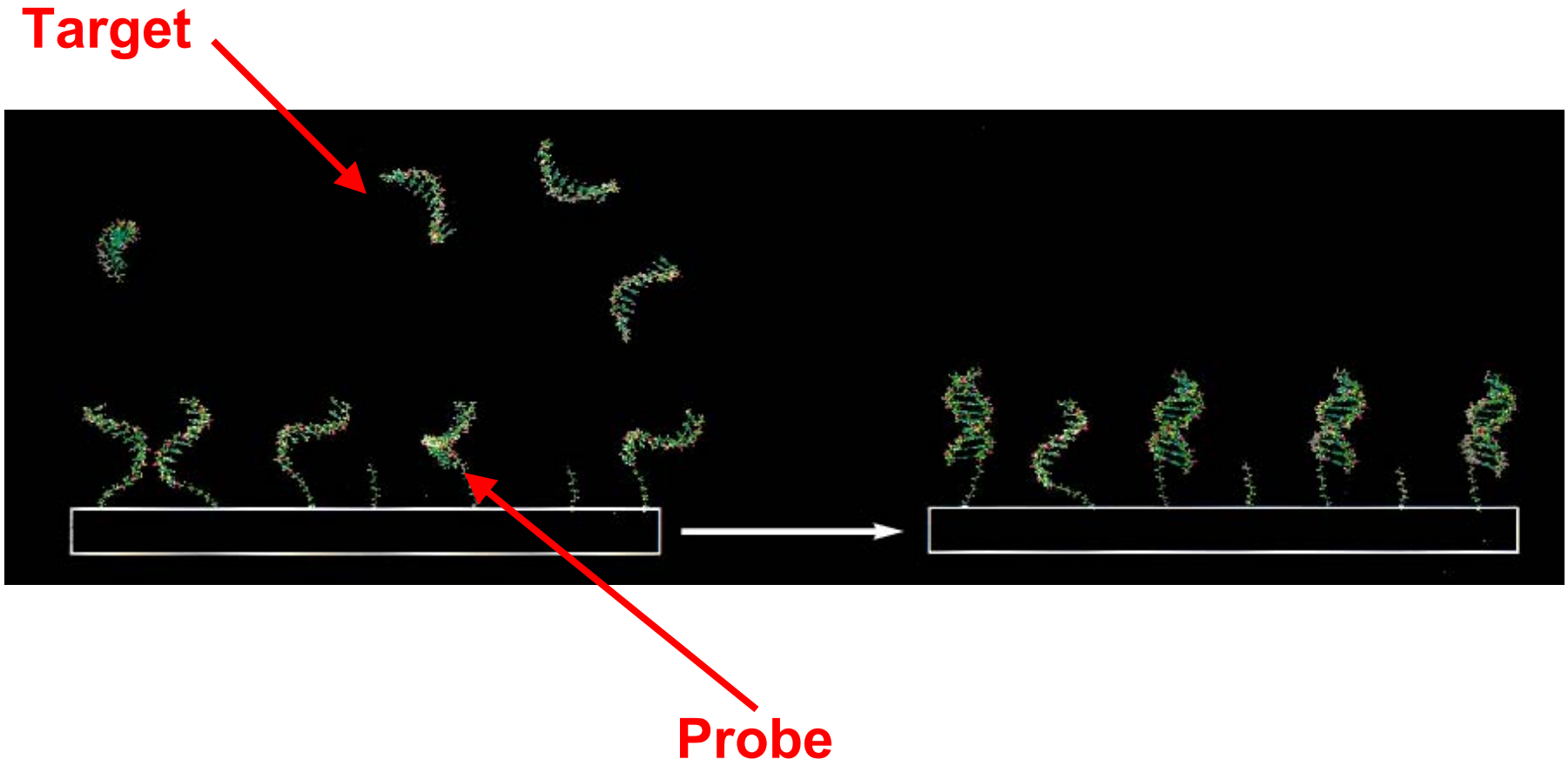
- Nucleic acid strands in a duplex can be separated, or denatured, by heating to destroy the hydrogen bonds.

Hybridization



Nucleic Acid Hybridization

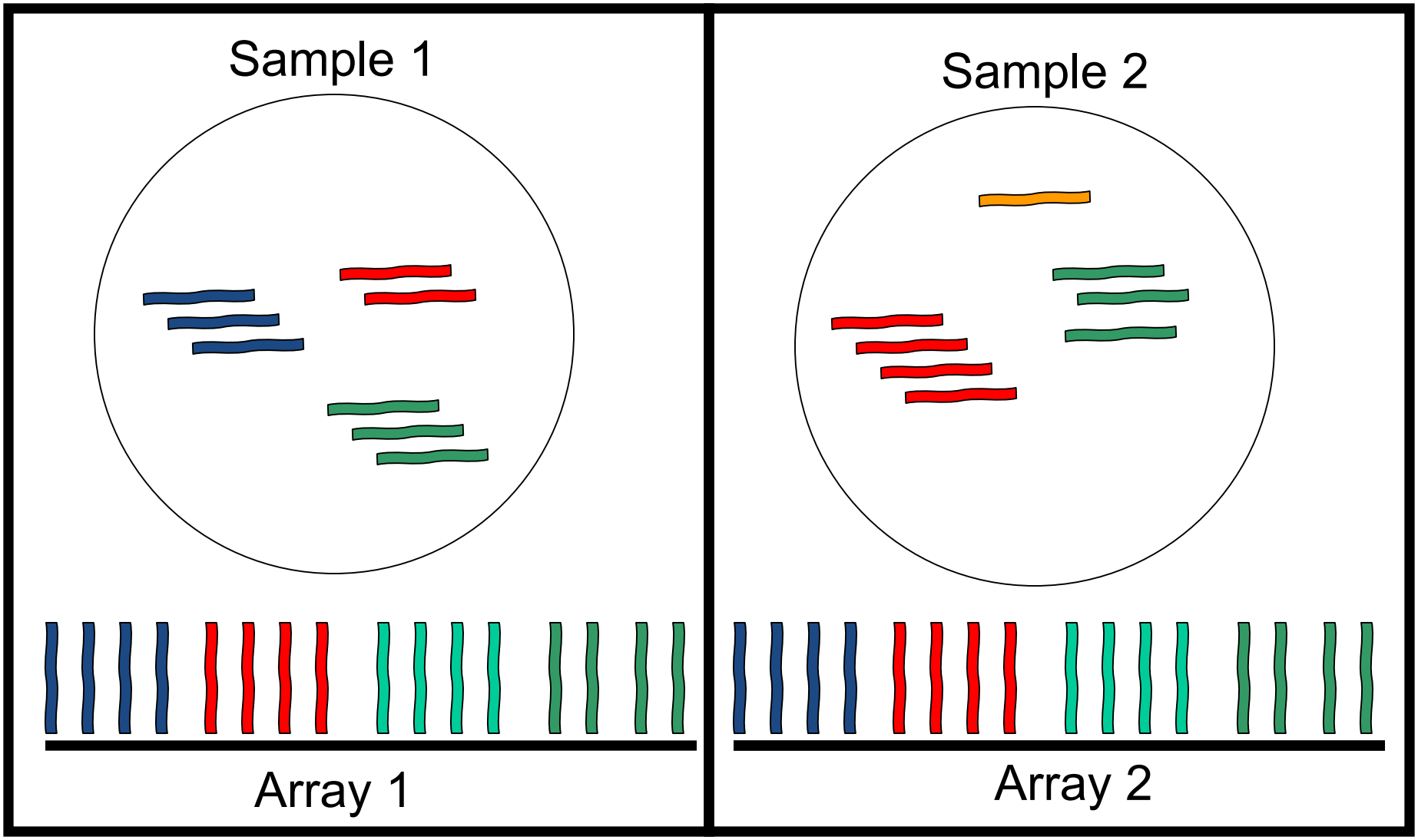
DNA microarrays



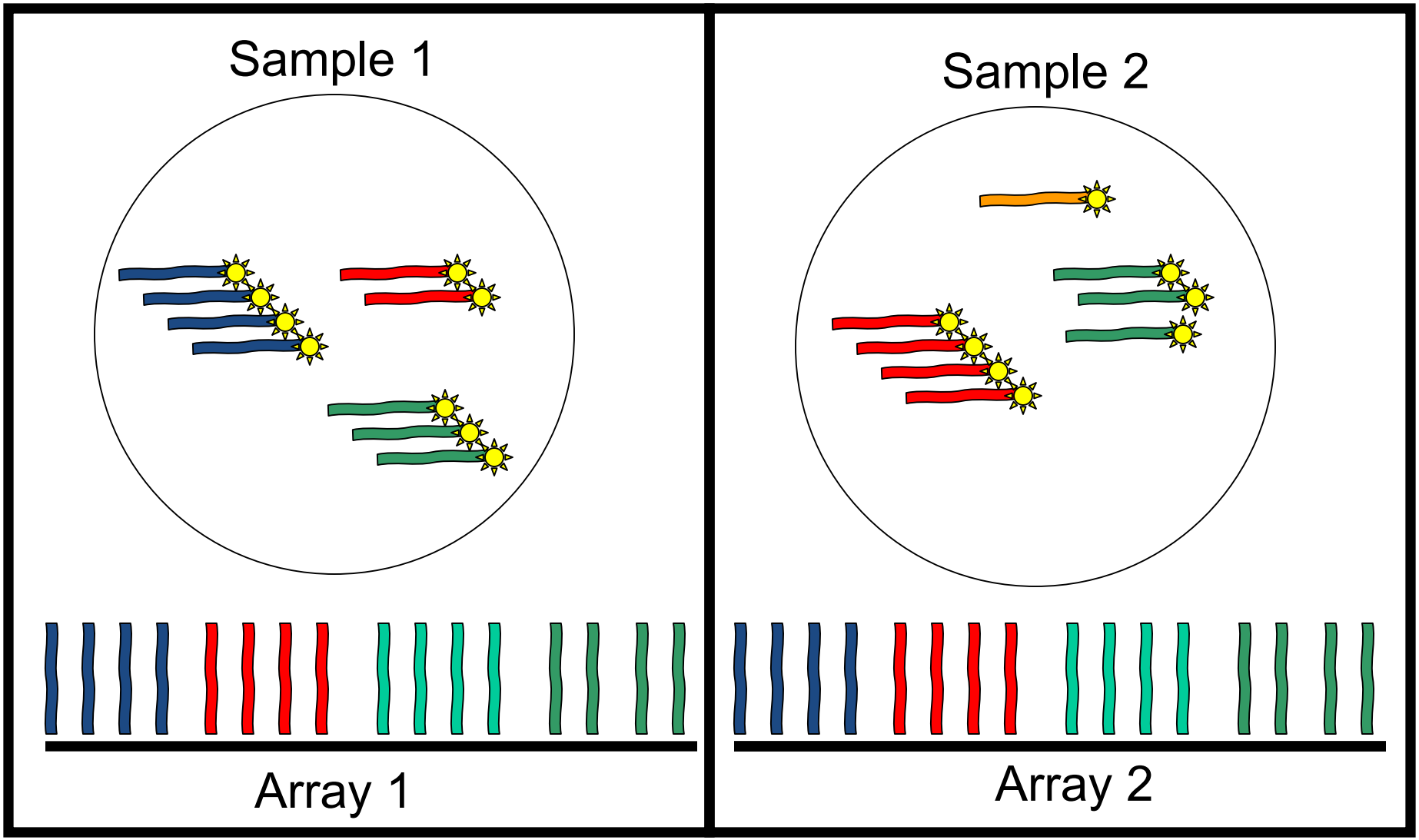
DNA microarrays

- The extent of hybridization of DNA sequences in the target sample to probe sequences on the array reflects the abundance of the probe sequences in the target sample.
- To quantify the extent of hybridization, the target sequences are fluorescently labeled.
- The hybridized arrays are scanned and the measured fluorescence intensities are used as measures of DNA/RNA abundance.

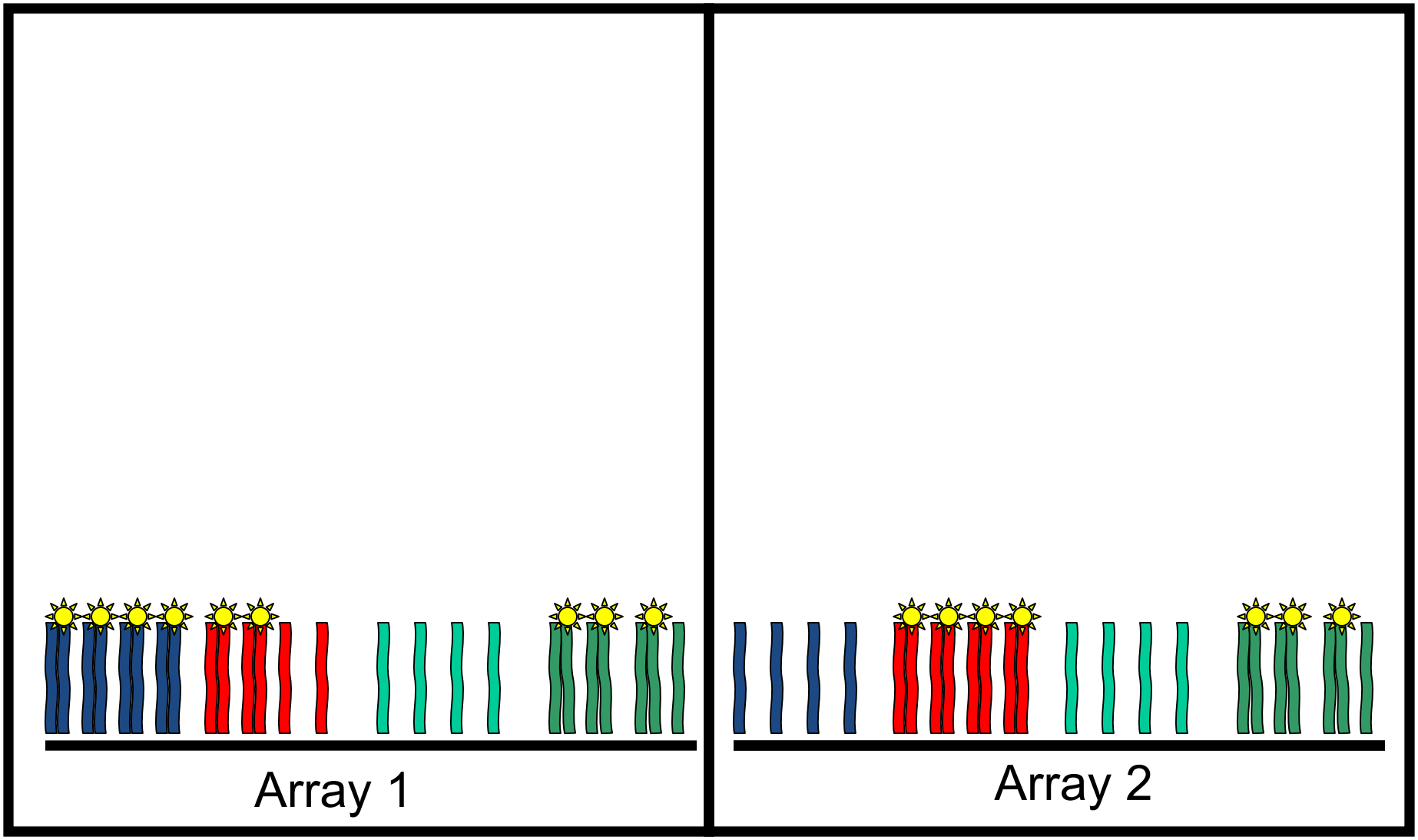
Before labeling



Before hybridization



After hybridization



Scanner image

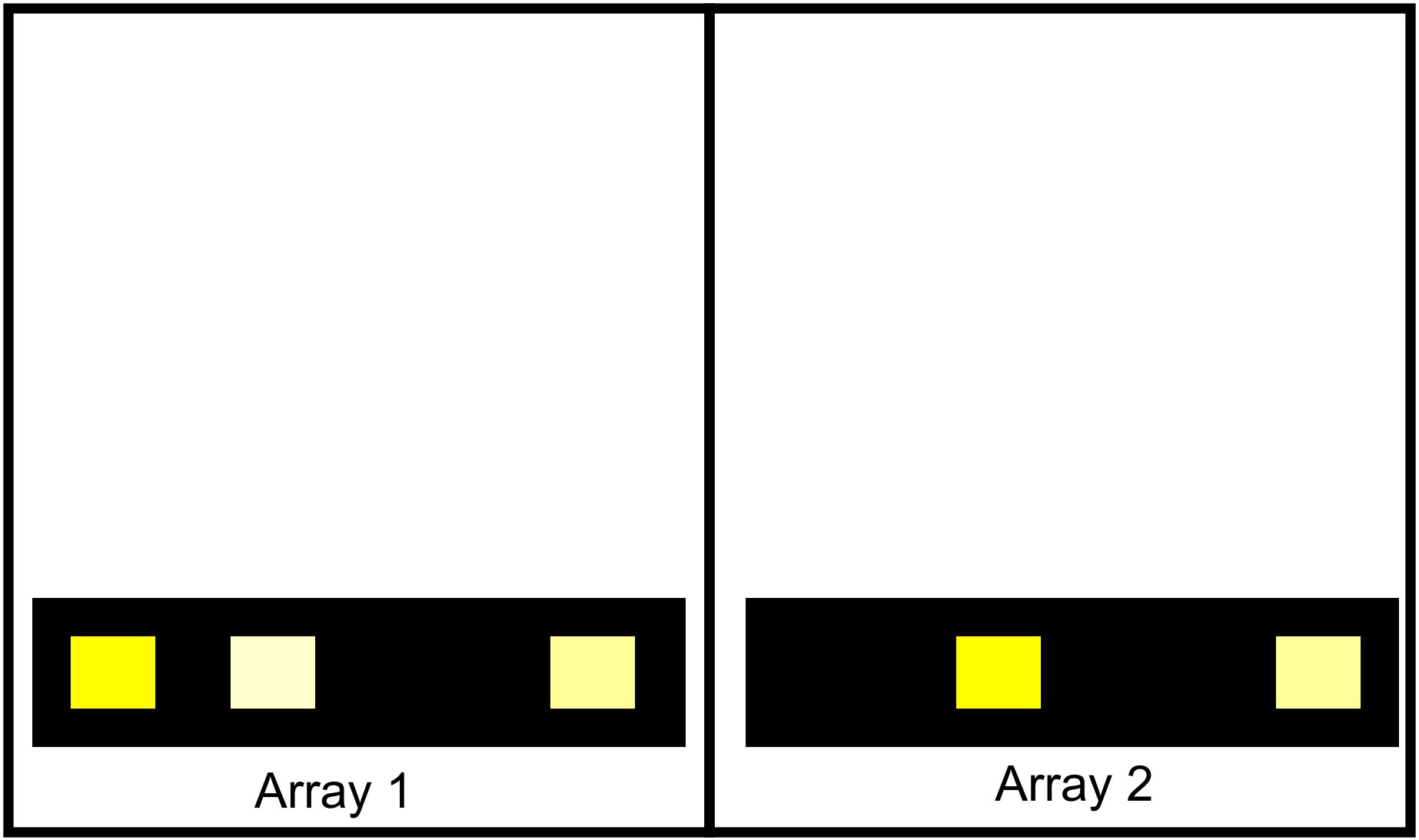
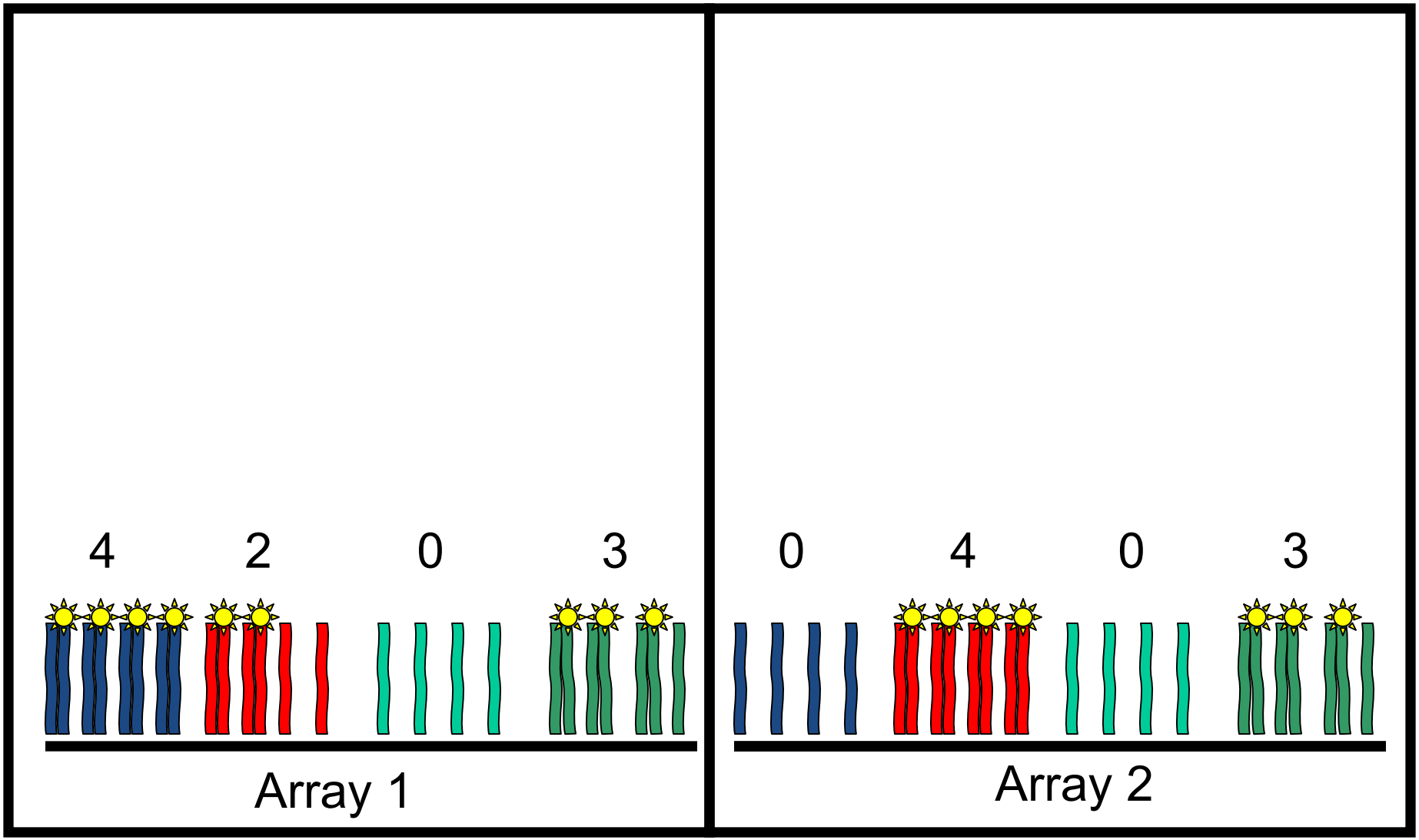


Image quantification



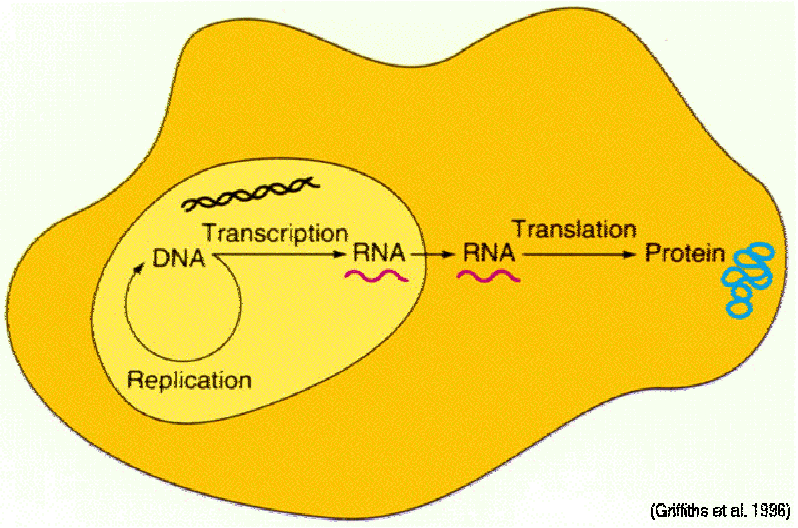
Gene expression assays

- Spotted cDNA arrays (Brown/Botstein);
- Short oligonucleotide arrays (Affymetrix);
- Long oligonucleotide arrays (Agilent Inkjet);
- Fibre optic arrays (Illumina);
- Serial analysis of gene expression (SAGE);
- …

Applications of microarrays

- Measuring transcript abundance (cDNA arrays);
- Genotyping;
- Estimating DNA copy number (CGH);
- Determining identity by descent (GMS);
- Measuring mRNA decay rates;
- Identifying protein binding sites;
- Determining sub-cellular localization of gene products;
- …

Transcriptome



- mRNA or transcript levels sensitively reflect the state of a cell.
- Measuring protein levels (translation) would be more direct but more difficult.

Transcriptome

- The transcriptome reflects
  - Tissue source: cell type, organ.
  - Tissue activity and state:
    - Stage of development, growth, death.
    - Cell cycle.
    - Disease vs. healthy.
    - Response to therapy, stress.

Applications of microarrays

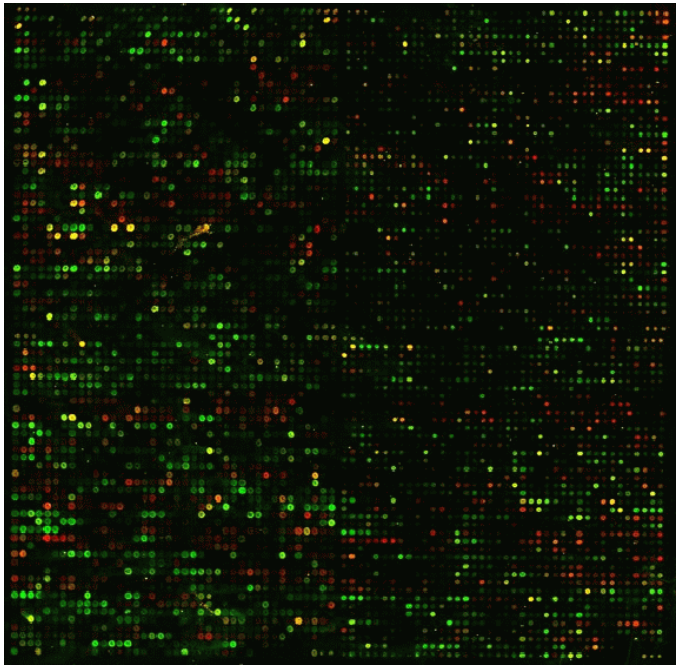
- **Cancer research:** Molecular characterization of tumors on a genomic scale

  → more reliable diagnosis and effective treatment of cancer.
- **Immunology:** Study of host genomic responses to bacterial infections.
- …

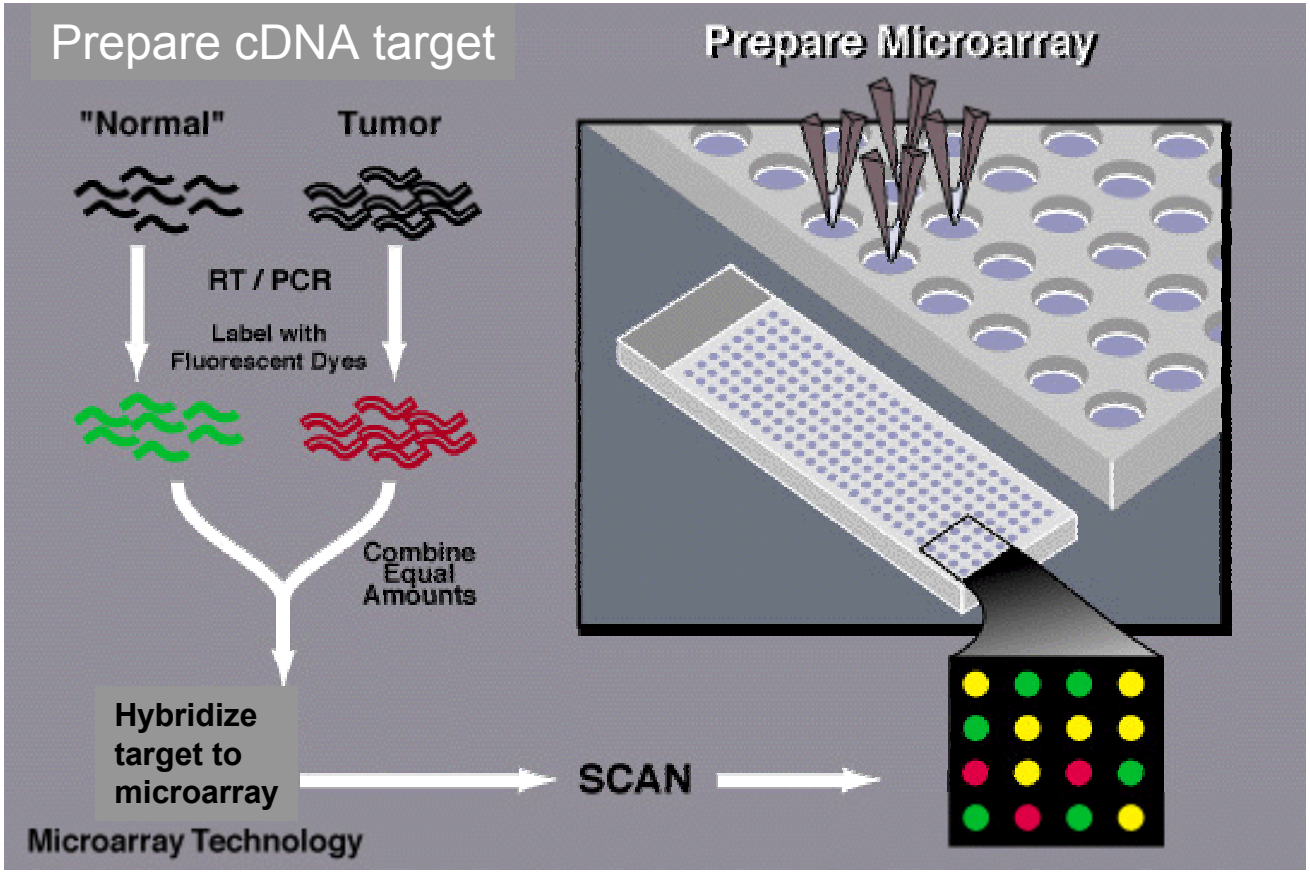
Applications of microarrays

- Compare mRNA (transcript) levels in different types of cells, i.e., vary
  - Tissue: liver vs. brain;
  - Treatment: drugs A, B, and C;
  - State: tumor vs. non-tumor, development;
  - Organism: different yeast strains;
  - Timepoint;
  - etc.

Spotted DNA microarrays



Spotted DNA microarrays



Spotted DNA microarrays

- The relative abundance of a spotted DNA sequence in two DNA or RNA samples may be assessed by monitoring the differential hybridization of these two samples to the sequence on the array.
- Probes: DNA sequences spotted on the array, immobile substrate.
- Targets: Nucleic acid samples hybridized to the array, mobile substrate.

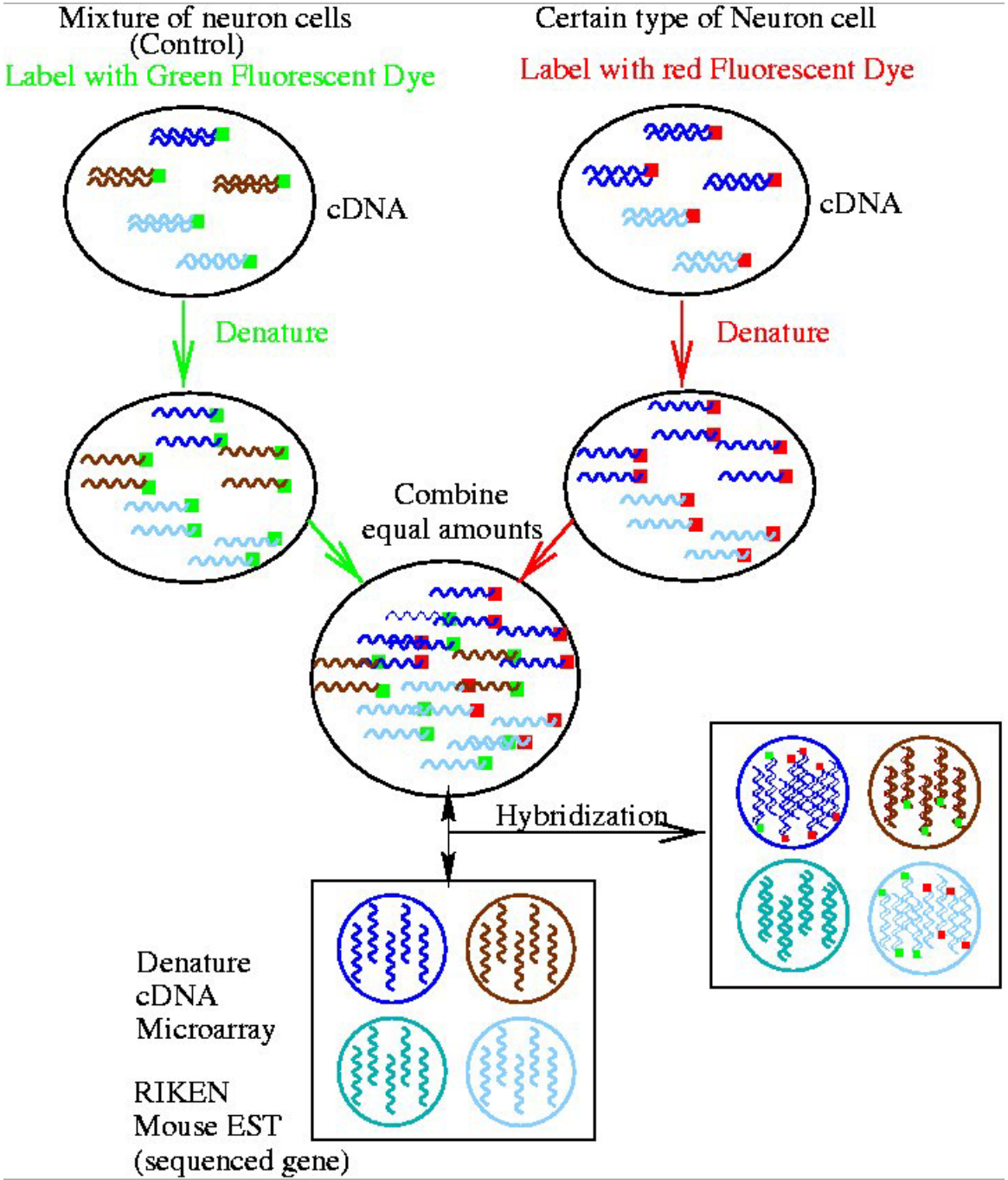
Spotted DNA microarrays

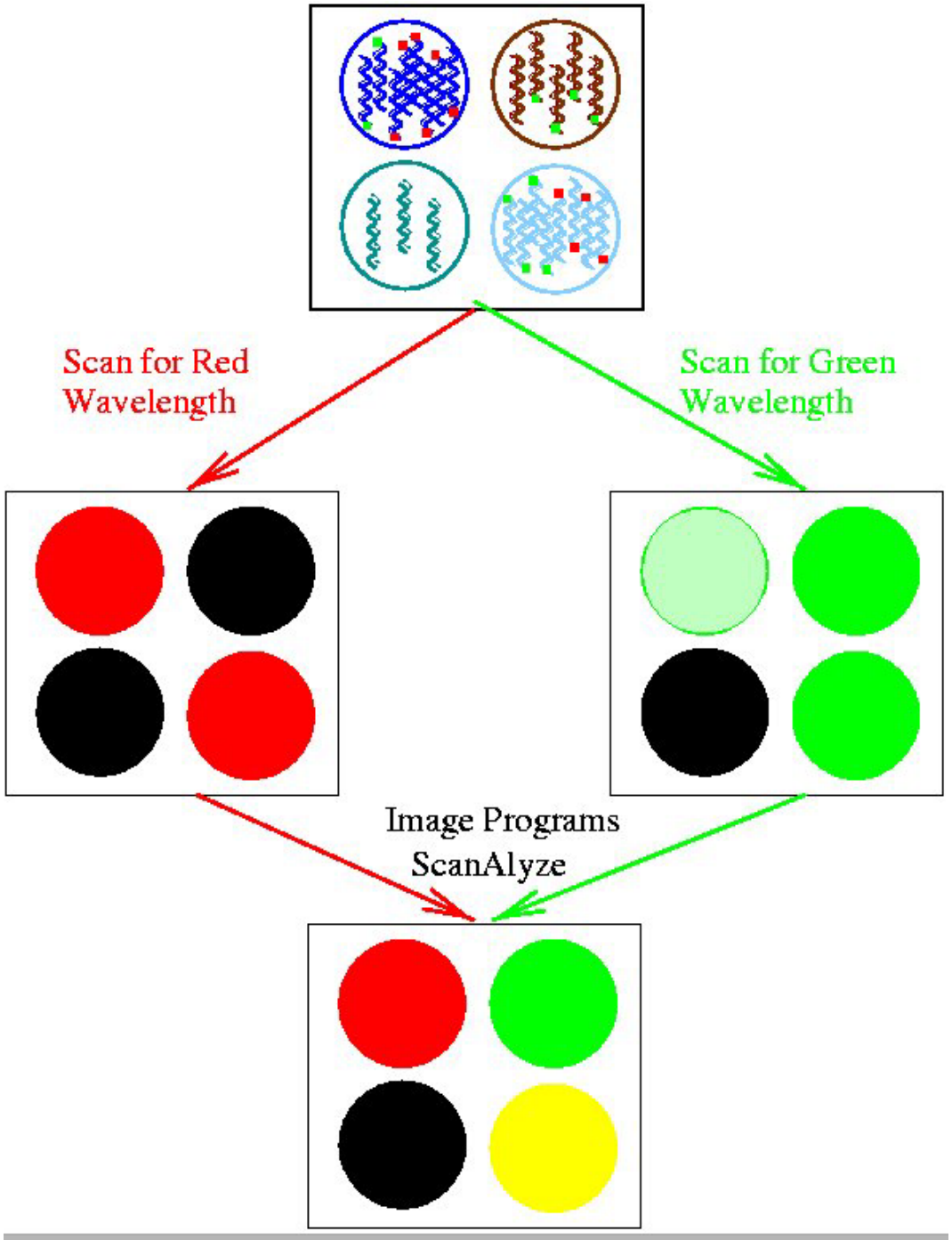
- The ratio of the red and green fluorescence intensities for each spot is indicative of the relative abundance of the corresponding DNA probe in the two nucleic acid target samples.

Spotted DNA microarrays

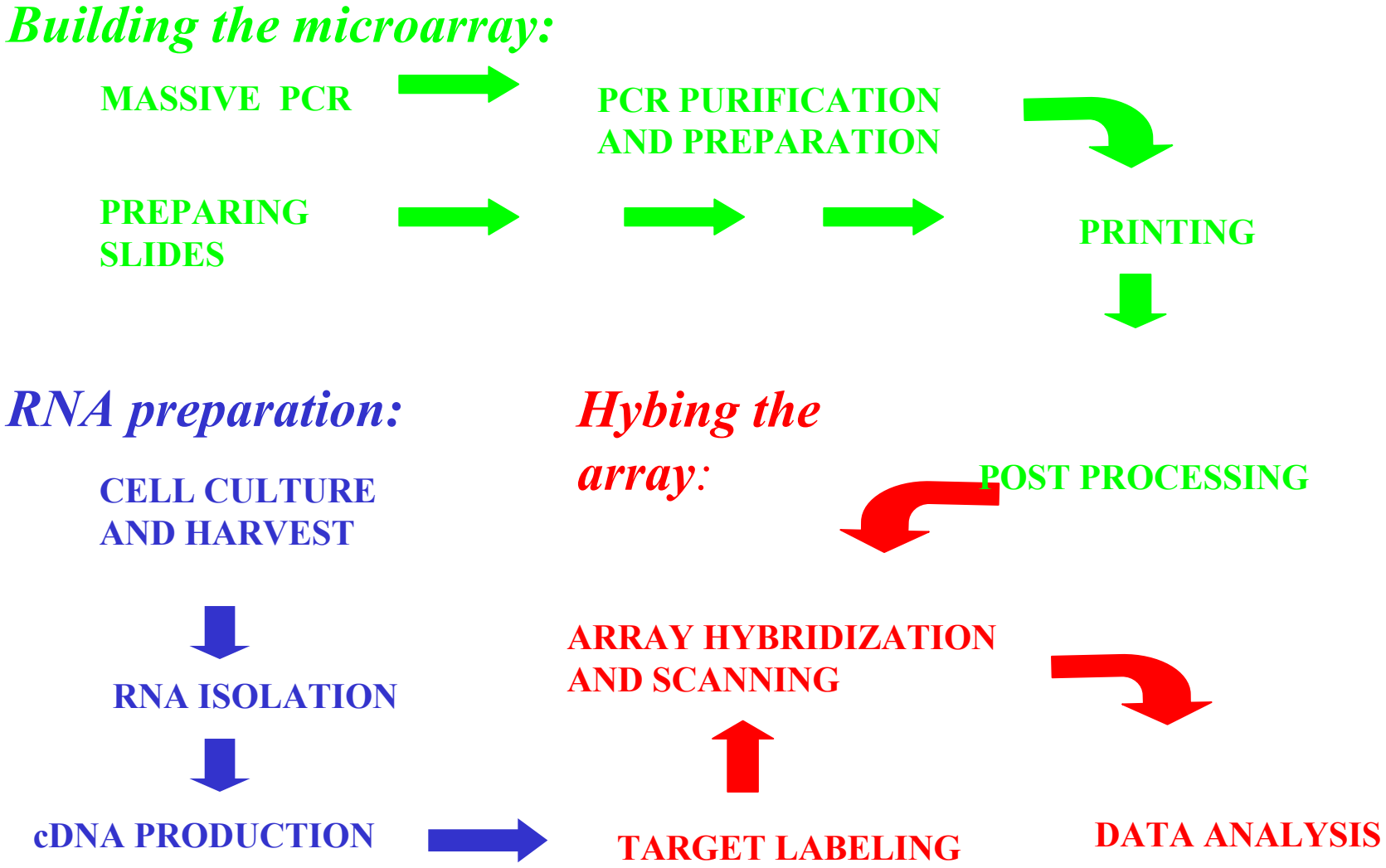
$$M = \log_2 R/G = \log_2 R - \log_2 G$$

- **M < 0**, gene is over-expressed in green-labeled sample compared to red-labeled sample.
- **M = 0**, gene is equally expressed in both samples.
- **M > 0**, gene is over-expressed in red-labeled sample compared to green-labeled sample.

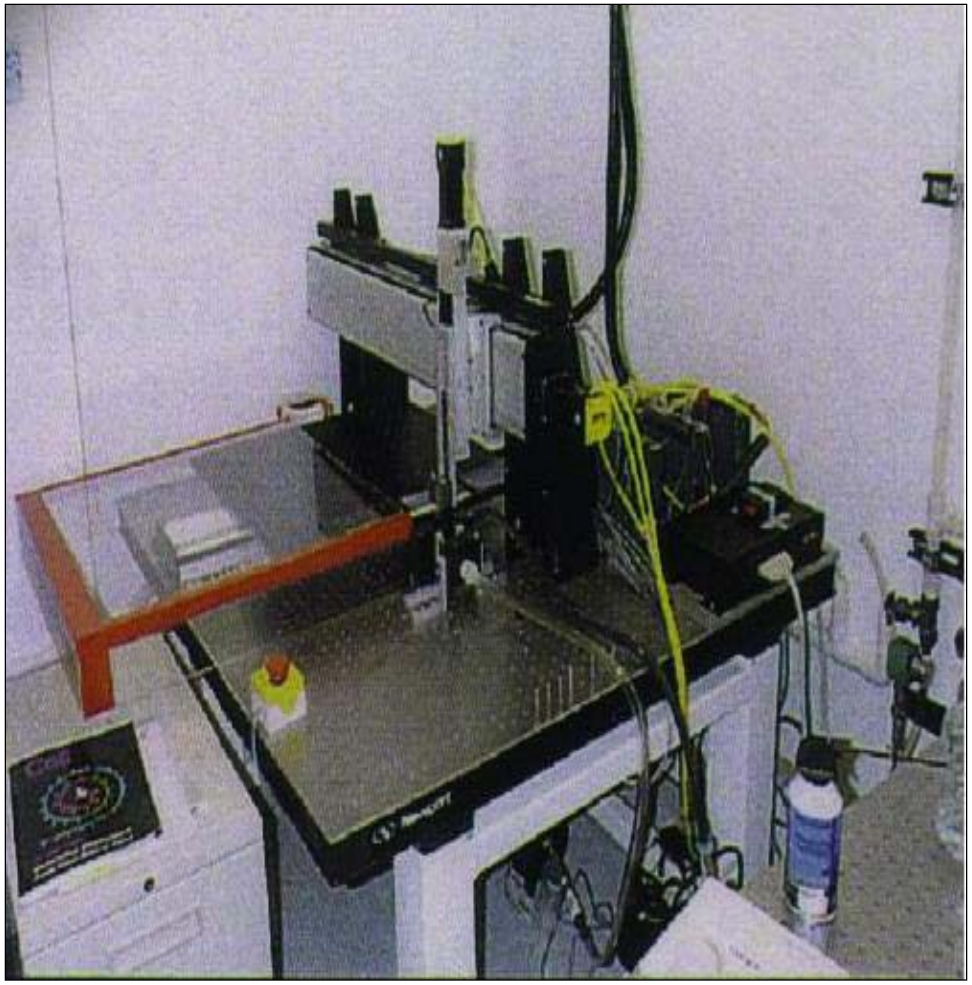

Mixture of neuron cells
(Control)
Label with Green Fluorescent Dye
Certain type of Neuron cell
Label with red Fluorescent Dye
cDNA
cDNA
Denature
Denature
Combine
equal amounts
Hybridization
Denature
cDNA
Microarray
RIKEN
Mouse EST
(sequenced gene)


Scan for Red
Wavelength
Scan for Green
Wavelength
Image Programs
ScanAlyze

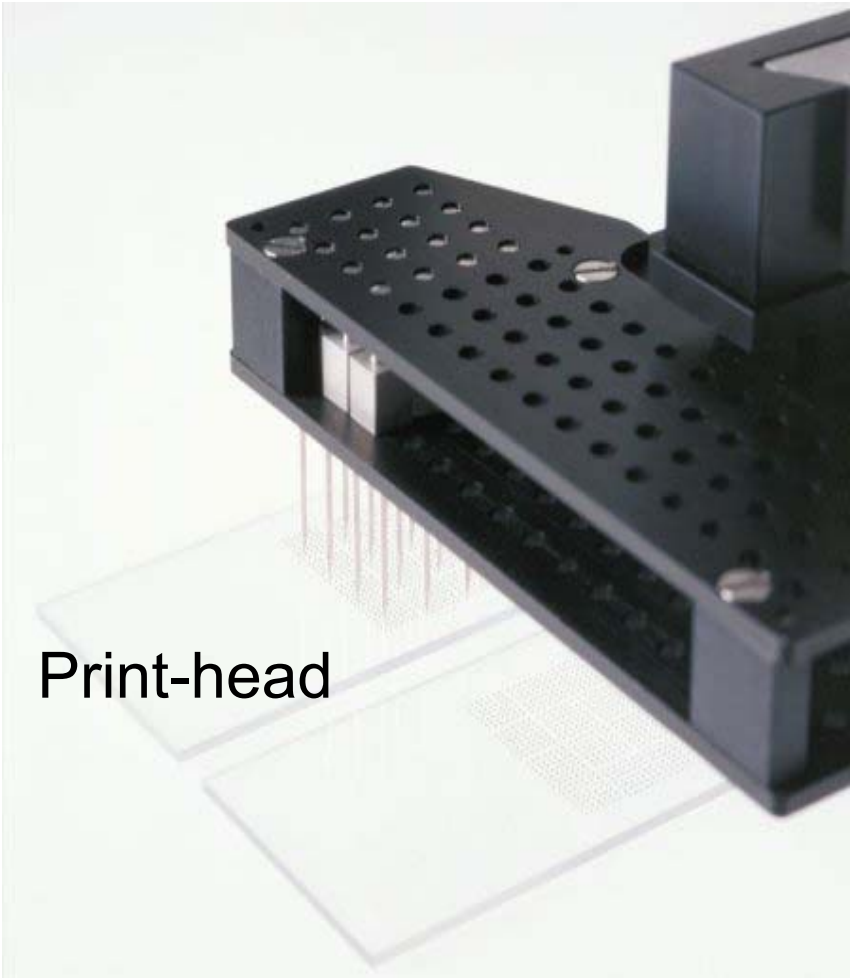
The process

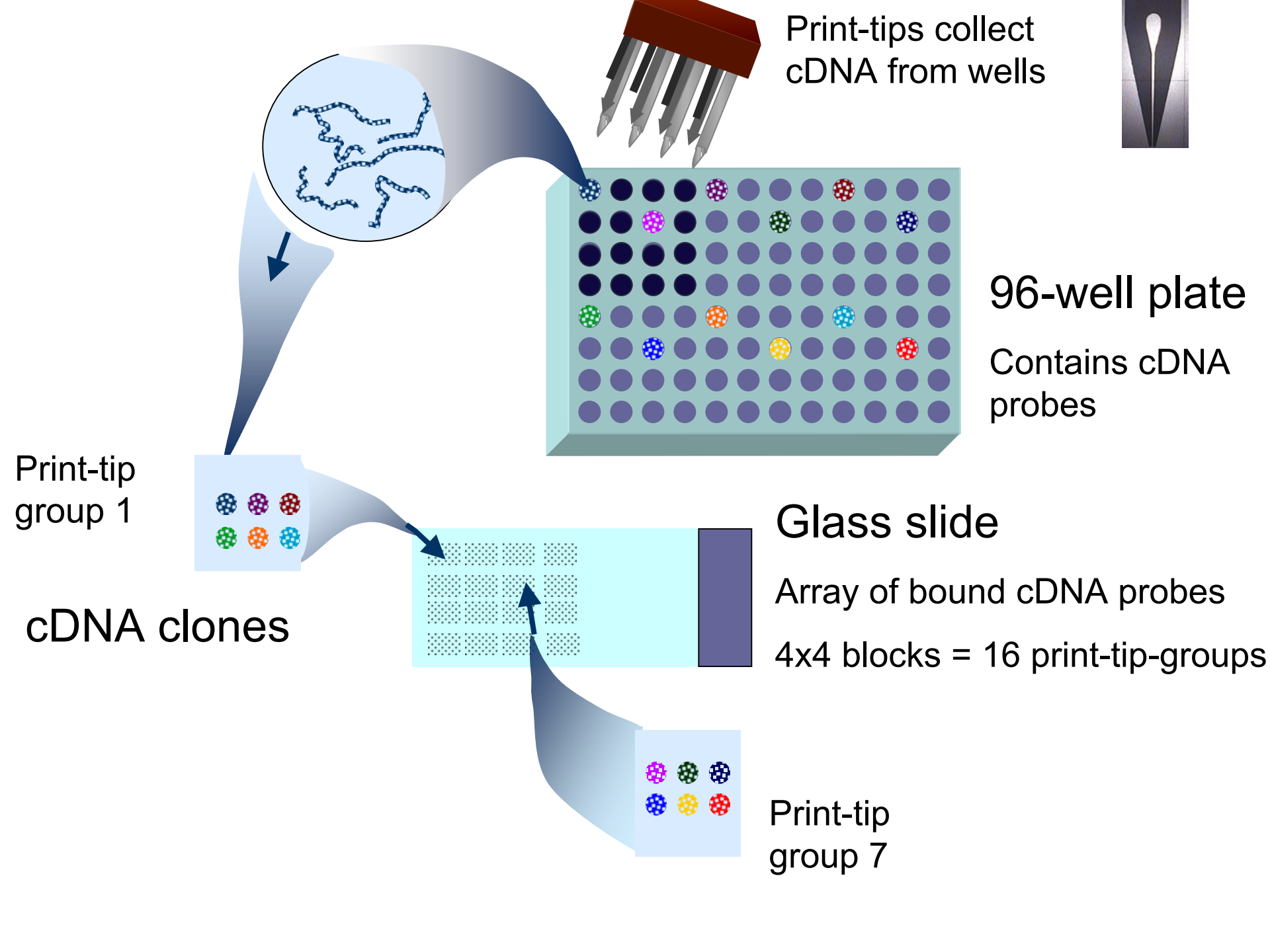

Building the microarray:
MASSIVE PCR
PCR PURIFICATION AND PREPARATION
PREPARING SLIDES
PRINTING
RNA preparation:
CELL CULTURE AND HARVEST
RNA ISOLATION
cDNA PRODUCTION
Hybing the array:
TARGET LABELING
ARRAY HYBRIDIZATION AND SCANNING
POST PROCESSING
DATA ANALYSIS

The arrayer

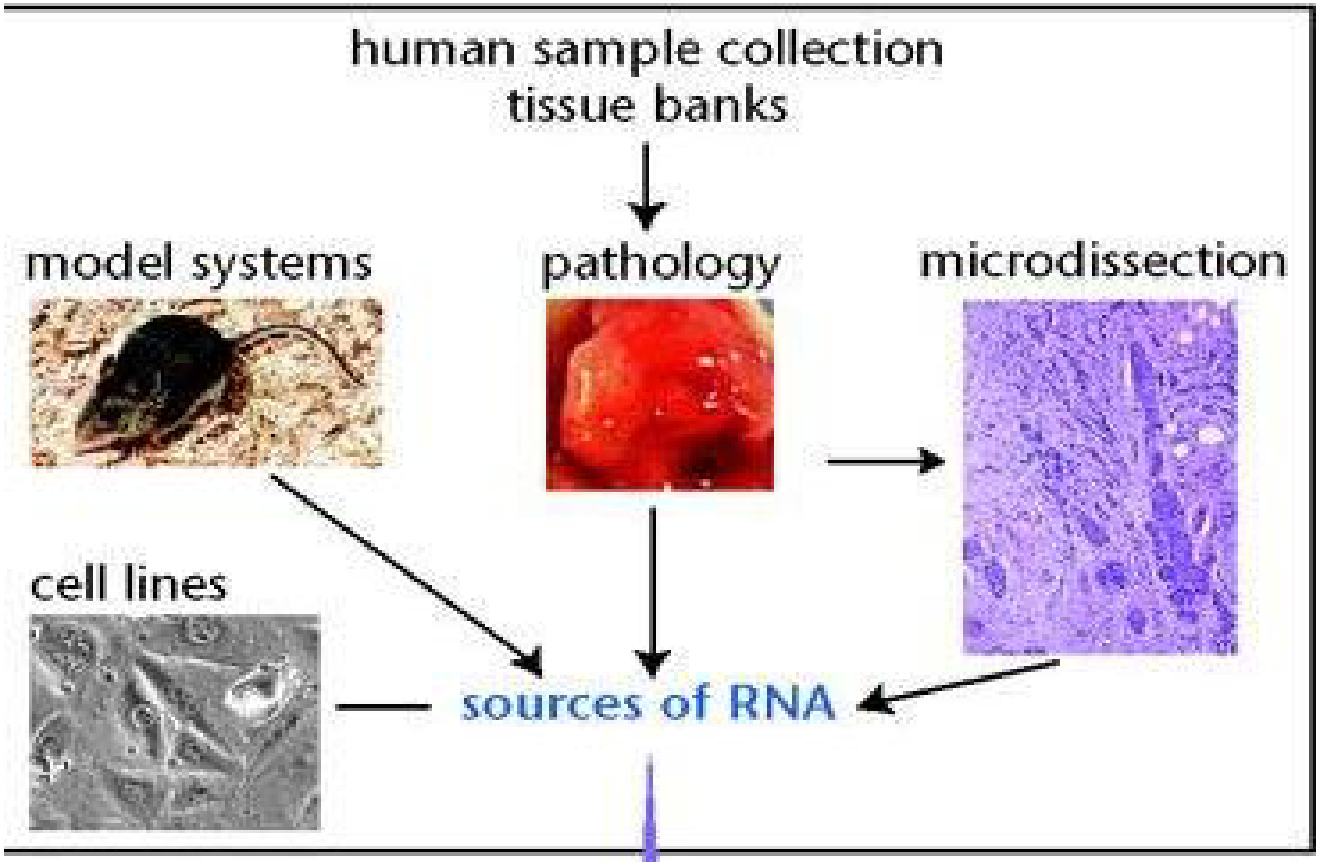


Ngai Lab arrayer, UC Berkeley

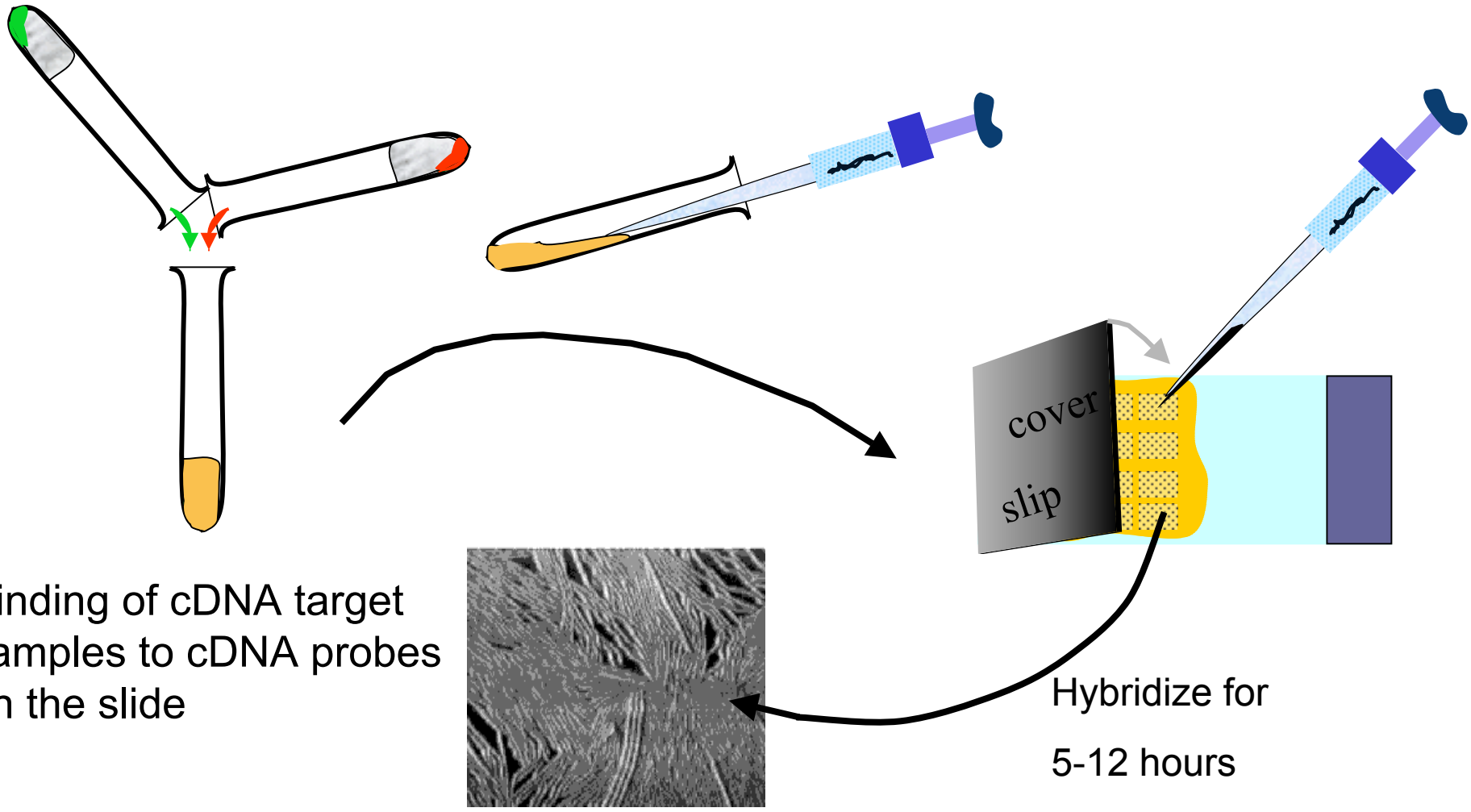



Print-tips collect cDNA from wells
96-well plate
Contains cDNA probes
Print-tip group 1
cDNA clones
Glass slide
Array of bound cDNA probes
4x4 blocks = 16 print-tip-groups
Print-tip group 7

Sample preparation



Hybridization

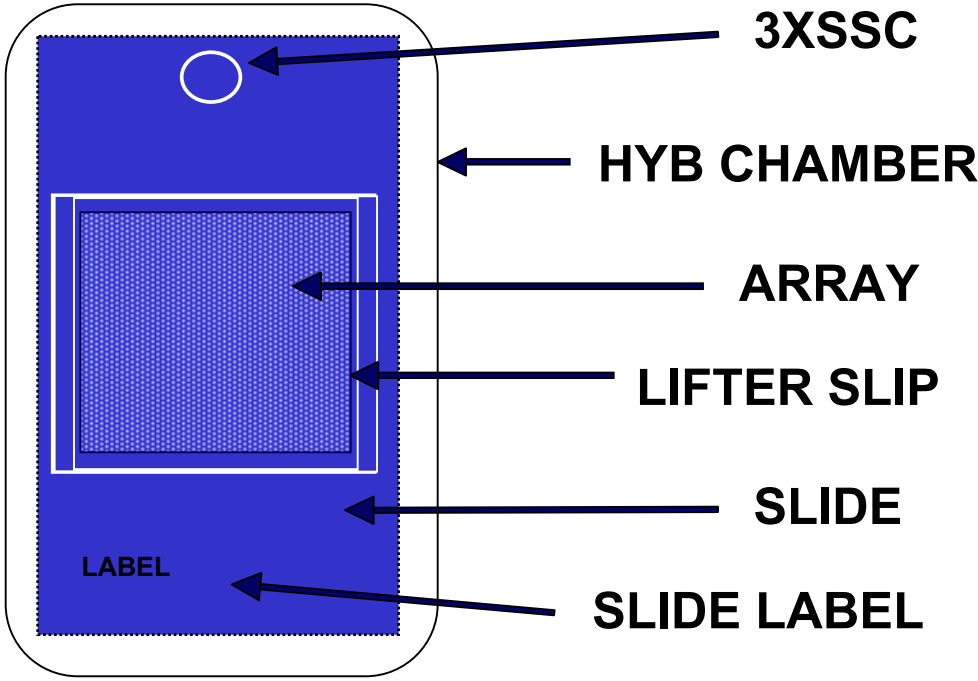


Binding of cDNA target samples to cDNA probes on the slide

Hybridize for

5-12 hours

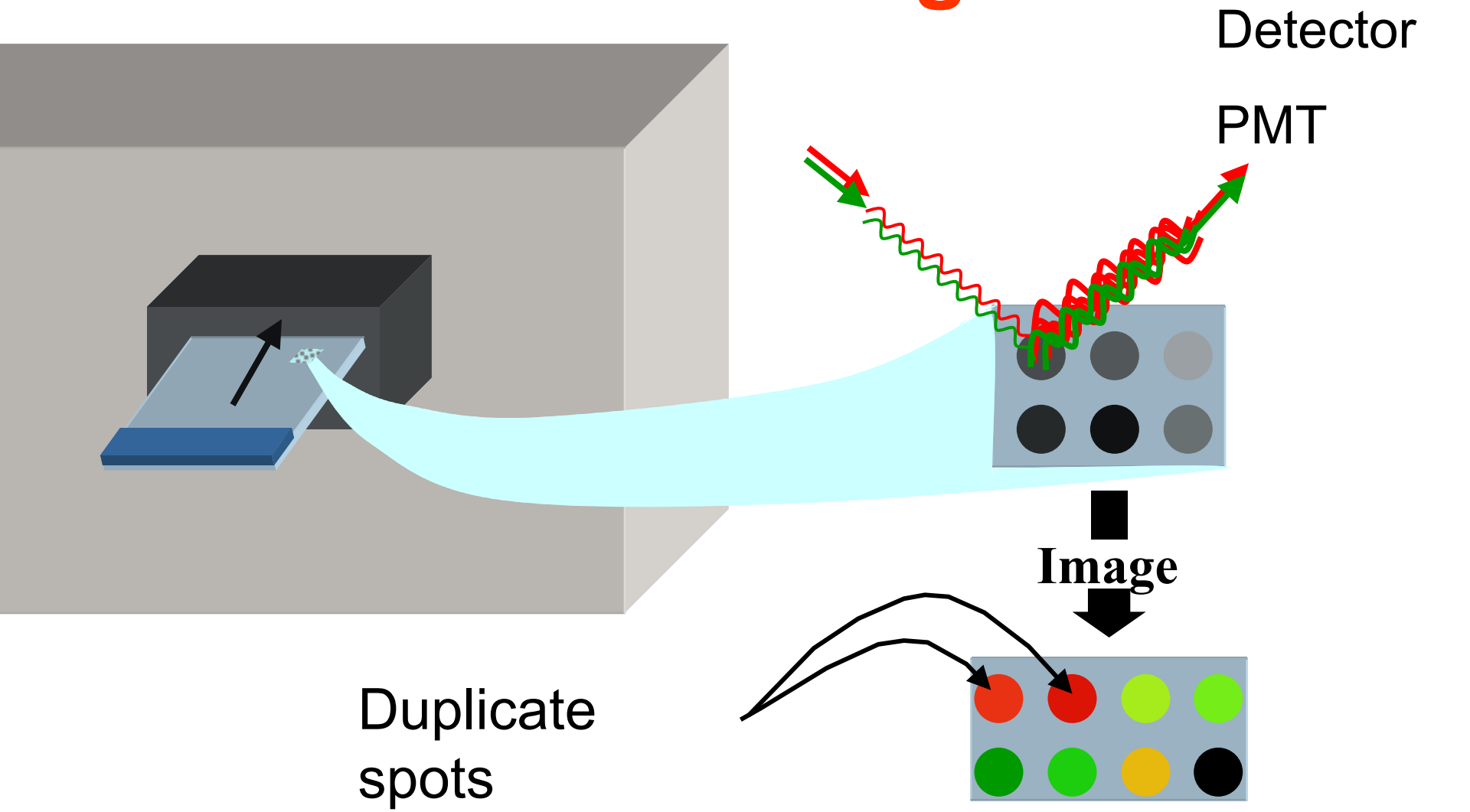
Hybridization chamber



- Humidity
- Temperature
- Formamide

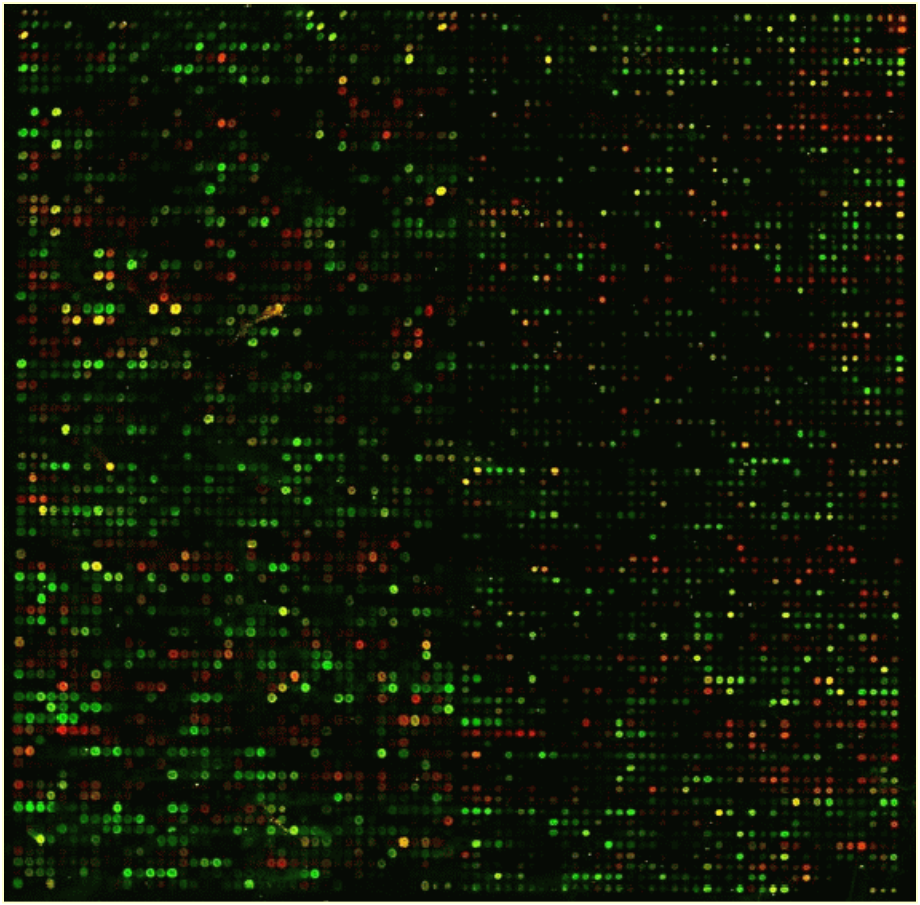
(Lowers the Tmp)

Scanning



Cy5: 635nm
Cy3: 532nm

RGB overlay of Cy3 and Cy5 images



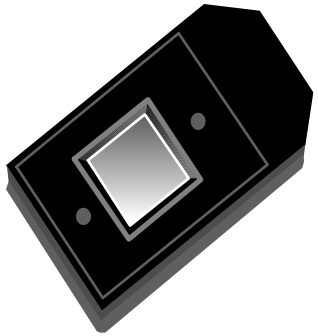
Raw data

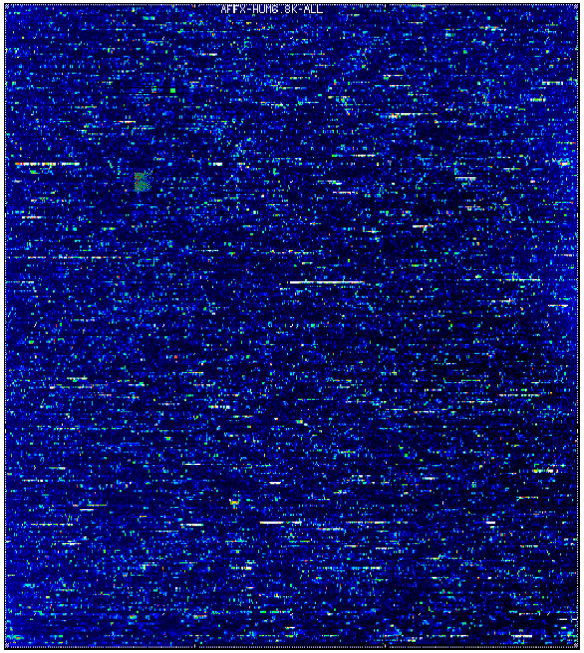
- Pairs of 16–bit TIFFs, one for each dye.
- E.g. Human cDNA arrays:
  - ~43K spots;
  - ~ 20Mb per channel;
  - ~ 2,000 x 5,500 pixels per image;
  - spot separation: ~ 136um.
- For a “typical” array, the spot area has
  - mean = 43 pixels,
  - med = 32 pixels,
  - SD = 26 pixels.

Animation

www.bio.davidson.edu/courses/genomics/chip/chip.html

Oligonucleotide chips





Terminology

- Each gene or portion of a gene is represented by 16 to 20 oligonucleotides of 25 base-pairs.

- Probe: an oligonucleotide of 25 base-pairs, i.e., a 25-mer.
- Perfect match (PM): A 25-mer complementary to a reference sequence of interest (e.g., part of a gene).
- Mismatch (MM): same as PM but with a single homomeric base change for the middle (13^{th}) base (transversion purine <-> pyrimidine, G <->C, A <->T) .
- Probe-pair: a (PM,MM) pair.
- Probe-pair set: a collection of probe-pairs (16 to 20) related to a common gene or fraction of a gene.
- Affy ID: an identifier for a probe-pair set.
- The purpose of the MM probe design is to measure non-specific binding and background noise.

Probe-pair set

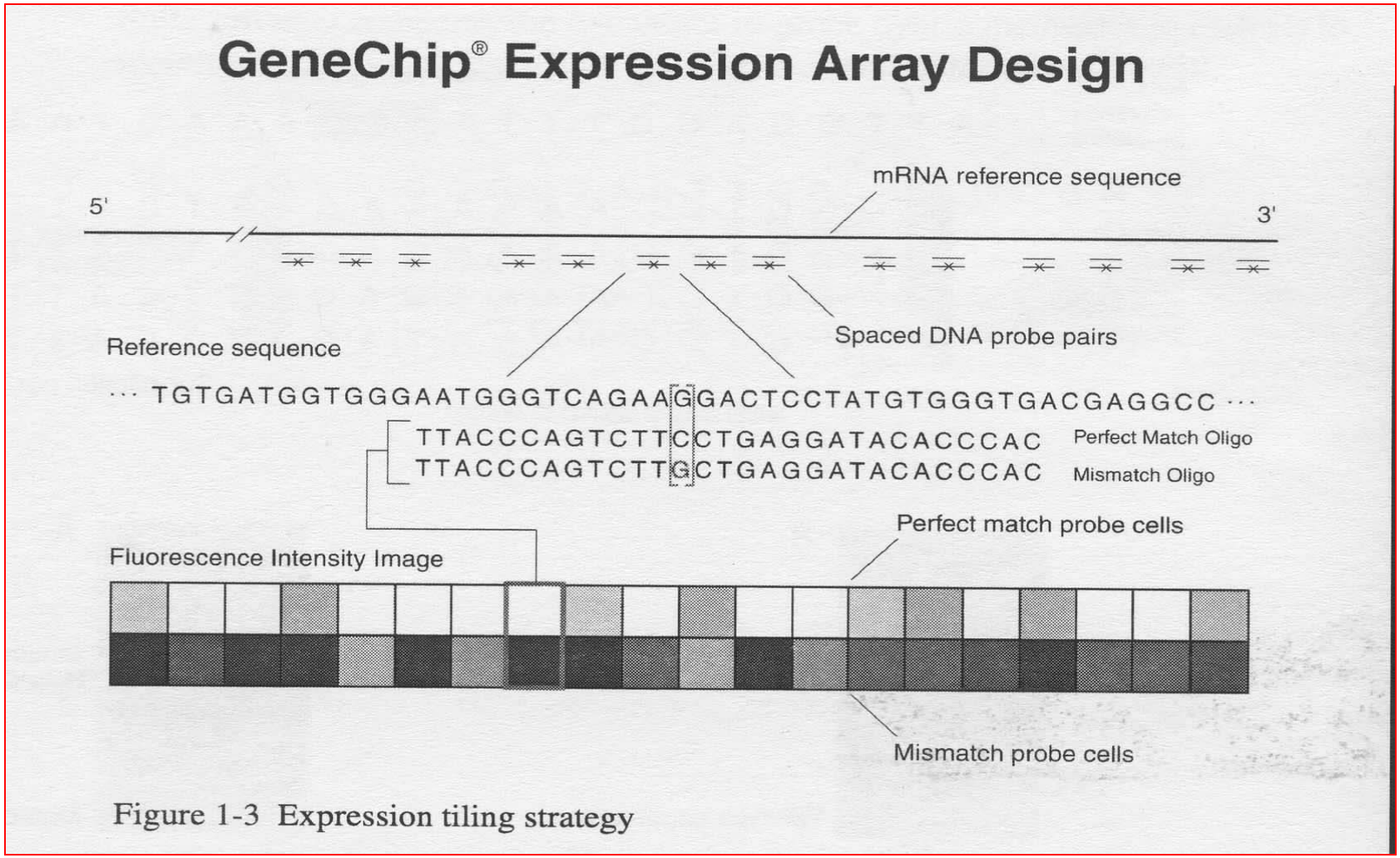
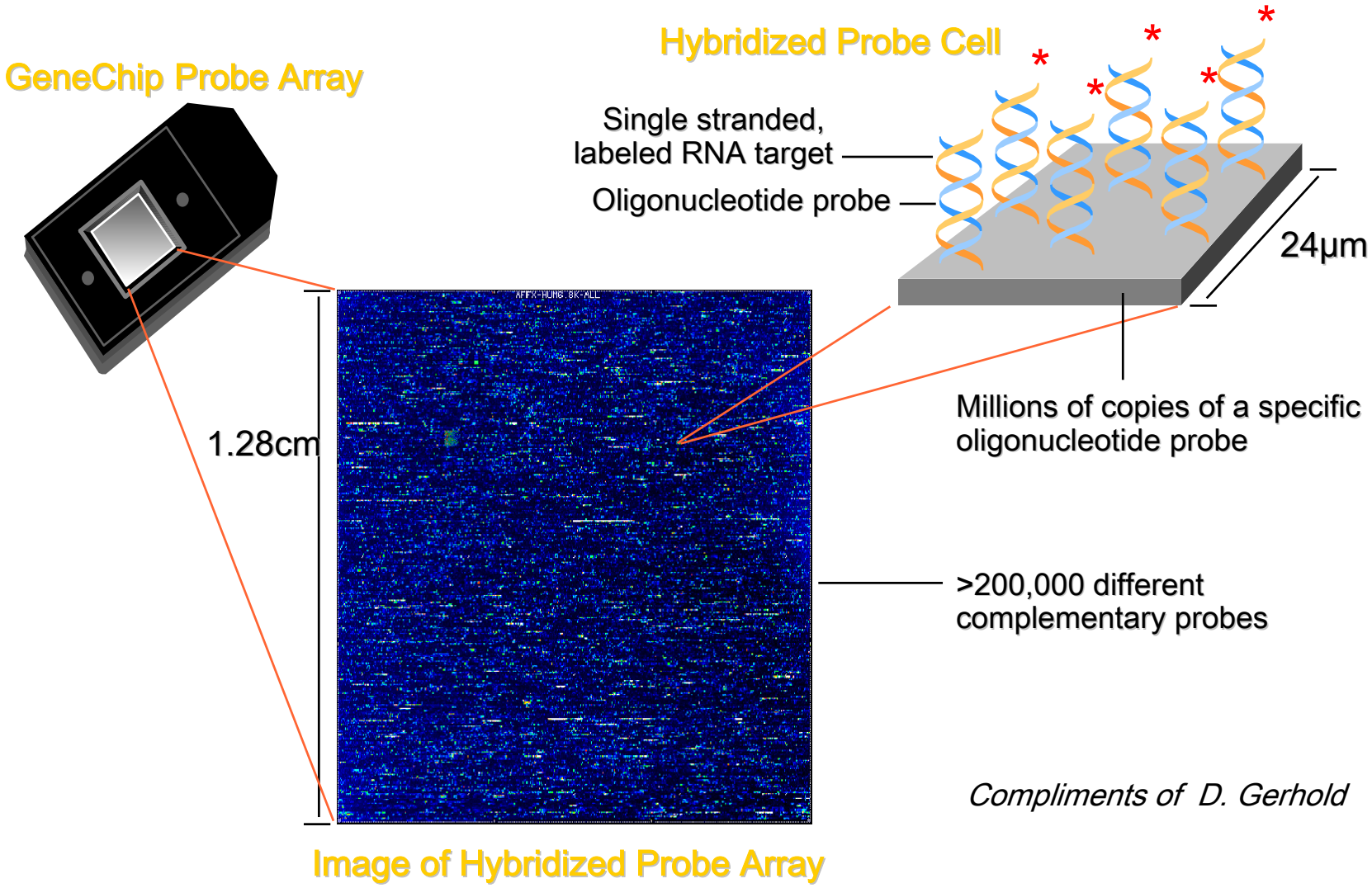


Figure 1-3 Expression tiling strategy

Spotted vs. Affymetrix arrays

Spotted arrays	Affymetrix arrays
One probe per gene	16 – 20 probe-pairs per gene
Probes of varying length	Probes are 25-mers
Two target samples per array	One target sample per array

Oligonucleotide chips

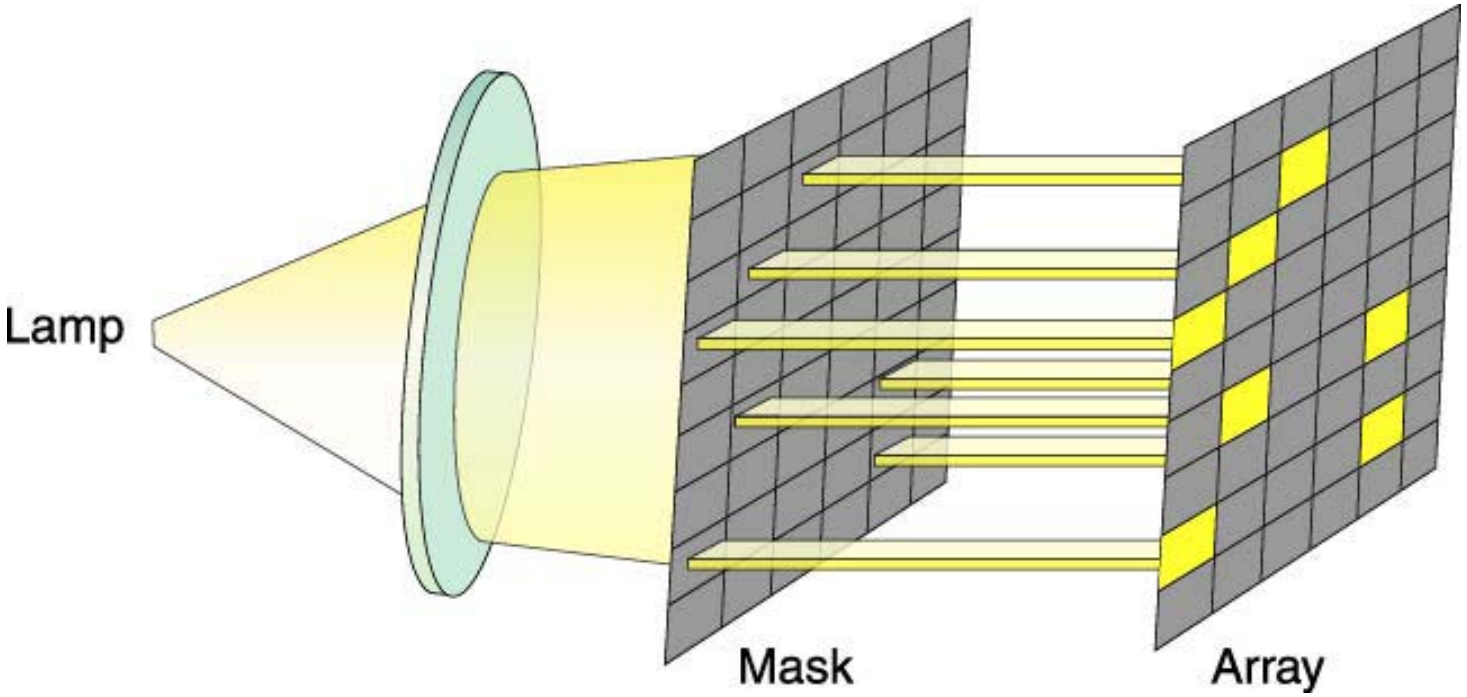


Compliments of D. Gerhold

Oligonucleotide chips

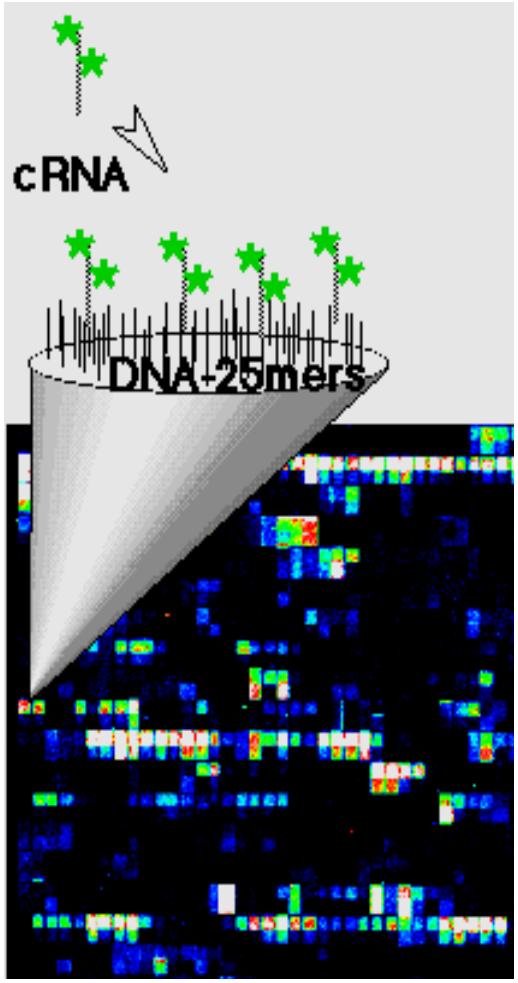
- The probes are synthesized *in situ*, using combinatorial chemistry and photolithography.
- Probe cells are square-shaped features on the chip containing millions of copies of a single 25-mer probe. Sides are 18-50 microns.

Oligonucleotide chips



The manufacturing of GeneChip® probe arrays is a combination of photolithography and combinatorial chemistry.

Image analysis



- About 100 pixels per probe cell.
- These intensities are combined to form one number representing the expression level for the probe cell oligo.
- → CEL file with PM or MM intensity for each cell.

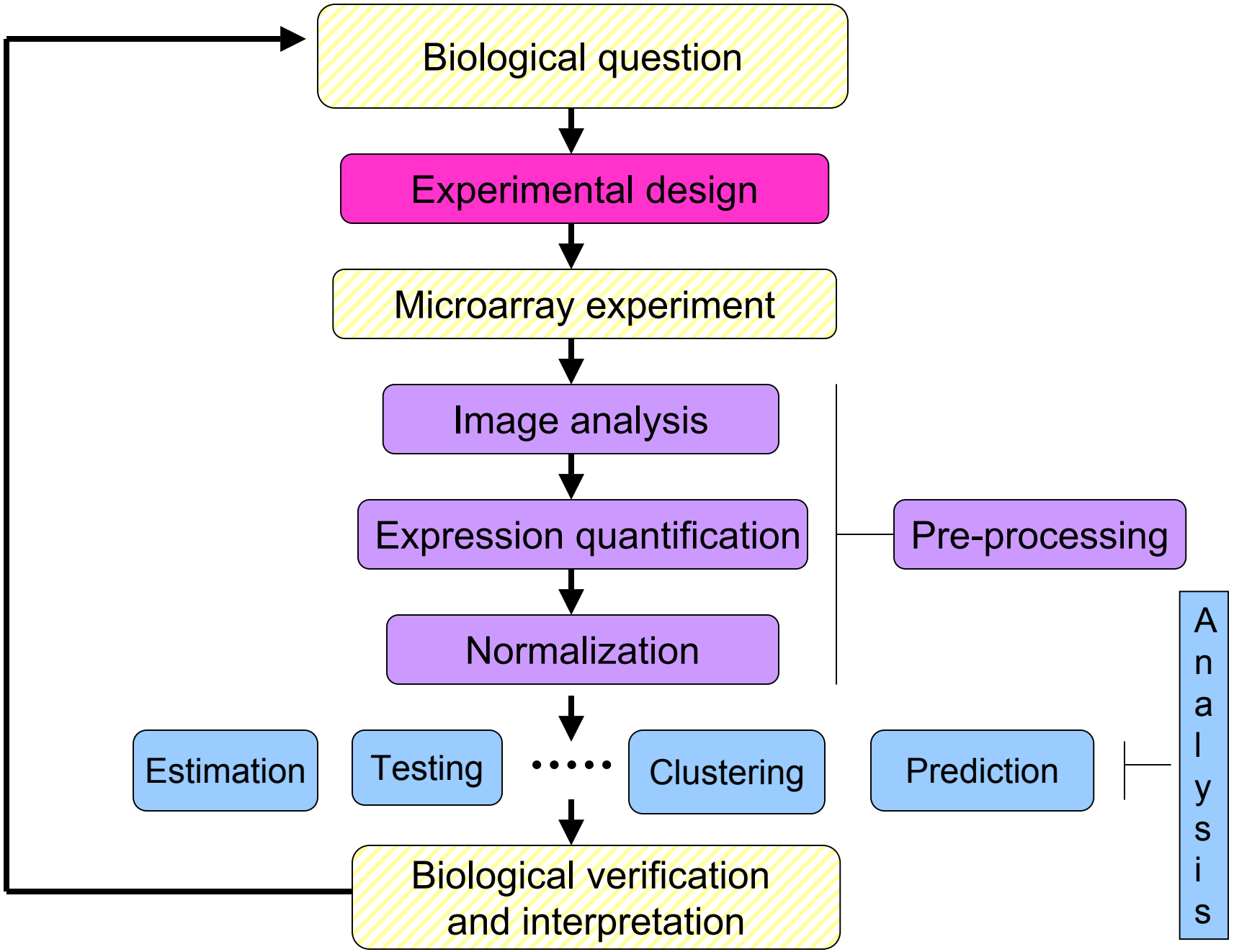
Expression measures

- Most expression measures are based on differences of **PM-MM**.
- The intention is to correct for background and non-specific binding.
- E.g. *MarrayArray Suite*® (MAS) v. 4.0 uses Average Difference Intensity (ADI) or

  AvDiff = average of PM-MM.
- Problem: MM may also measure signal.
- More on this in lecture *Pre-processing DNA Microarray Data.*

WWW resources

- **Complete guide to "microarraying"**
  http://cmgm.stanford.edu/pbrown/mguide/
  http://www.microarrays.org
  - Parts and assembly instructions for printer and scanner;
  - Protocols for sample prep;
  - Software;
  - Forum, etc.
- **cDNA microarray animation**
  http://www.bio.davidson.edu/courses/genomics/chip/chip.html
- **Affymetrix**
  http://www.affymetrix.com


Biological question
Experimental design
Microarray experiment
Image analysis
Expression quantification
Normalization
Pre-processing
Estimation
Testing
.....
Clustering
Prediction
Analysis
Biological verification
and interpretation

Statistical computing

Everywhere …

- Statistical design and analysis:
  - image analysis, normalization, estimation, testing, clustering, prediction, etc.

- Integration of experimental data with biological metadata from WWW-resources
  - gene annotation (GenBank, LocusLink);
  - literature (PubMed);
  - graphical (pathways, chromosome maps).

Outline

- Introduction to the biology and technology of DNA microarrays
- Overview of the Bioconductor project
- Annotation
- Visualization
- Pre-processing: spotted and Affymetrix arrays
- Differential gene expression
- Software demo

Overview of the Bioconductor Project

Bioconductor

- Bioconductor is an open source and open development software project for the analysis and comprehension of biomedical and genomic data.

- Software, data, and documentation are available from www.bioconductor.org.

Bioconductor

- The project was started in the Fall of 2001 by Robert Gentleman, at the Biostatistics Unit of the Dana Farber Cancer Institute.
- There are currently 21 core developers, at various institutions in the US and Europe.
- R and the R package system are used to design and distribute software (www.r-project.org).
- First release (v 1.0): May 2nd, 2002, 15 packages.
- Second release (v 1.1): November 18th, 2002, 5 new packages.

Bioconductor

There are two main classes of packages

- End-user packages:
  - aimed at users unfamiliar with R or computer programming;
  - polished and easy to use interfaces to a wide variety of computational and statistical methods for the analysis of genomic data.
- Developer packages: aimed at software developers, in the sense that they provide ``software to write software''.

Bioconductor packages

Release 1.1, November 18th, 2002

- General infrastructure:
  Biobase, **reposTools**, **rhdf5**, **tkWidgets**.
- Annotation:
  annotate, **AnnBuilder** → data packages.
- Graphics:
  geneplotter, **hexbin**.
- Pre-processing for Affymetrix oligonucleotide chip data:
  affy, **vsn**, CDF packages.
- Pre-processing for spotted DNA microarray data:
  marrayClasses, **marrayInput**, **marrayNorm**, **marrayPlots**, **marrayTools**, **vsn**.
- Differential gene expression:
  edd, **genefilter**, **multtest**, **ROC**.
- Graphs:
  graph.

Ongoing efforts

- Variable (feature) selection;
- Prediction;
- Cluster analysis;
- Cross-validation;
- Multiple testing;
- Quality measures for microarray data;
- Interactions with MAGE-ML;
- Biological sequence analysis;
- Etc.

Computing needs

- Mechanisms for facilitating the design and deployment of portable, extensible, and scalable software.
- Support for interoperability with software written in other languages.
- Tools for integrating biological metadata from the WWW in the analysis of experimental metadata.
- Access to a broad range of statistical and numerical methods.
- High-quality visualization and graphics tools that support interactivity.
- An effective, extensible user interface.
- Tools for producing innovative, high-quality documentation and training materials.
- Methodology that supports the creation, testing, and distribution of software and data modules.

Bioconductor

- Interactive tools for linking experimental data in real time, to biological metadata from WWW resources.

  E.g. PubMed, GenBank, LocusLink.
- Scenario. Normalize spotted array data with **`marrayNorm`**, obtain list of differentially expressed genes from **`multtest`** or **`genefilter`**, use the **`annotate`** package
  - to retrieve and search PubMed abstracts for these genes;
  - to generate an HTML report with links to LocusLink for each gene.

Bioconductor

- Widgets. Small-scale graphical user interfaces (GUI), providing point & click access for specific tasks (`tkWidgets`).
- E.g. File browsing and selection for data input, basic analyses.

- Object-oriented class/method design. Allows efficient representation and manipulation of large and complex biological datasets of multiple types (cf. MIAME standards).

Object-oriented programming

- The Bioconductor project has adopted the object-oriented programming – OOP – paradigm presented in

  J. M. Chambers (1998). *Programming with Data*.
- Tools for programming using the class/method mechanism are provided in the R **`methods`** package.
- Tutorial: www.omegahat.org/RSMethods/index.html

OOP

- A class provides a software abstraction of a real world object. It reflects how we think of certain objects and what information these objects should contain.
- Classes are defined in terms of slots which contain the relevant data.
- An object is an instance of a class.
- A class defines the structure, inheritance, and initialization of objects.

OOP

- A method is a function that performs an action on data (objects).
- Methods define how a particular function should behave depending on the class of its arguments.
- Methods allow computations to be adapted to particular data types, i.e., classes.
- A generic function is a dispatcher, it examines its arguments and determines the appropriate method to invoke.
- Examples of generic functions include **`plot`**, **`summary`**, **`print`**.

Data

- Issues:
  - complexity;
  - size;
  - evolution.

- We distinguish between biological metadata and experimental metadata.

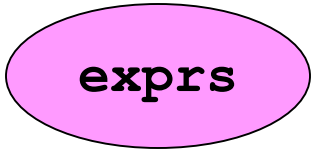
Experimental metadata

- Gene expression measures
  - scanned images, i.e., raw data;
  - image quantitation data, i.e., output from image analysis;
  - normalized expression measures, i.e., log ratios M or Affy measures.
- Reliability information for the expression measures.
- Information on the probe sequences printed on the arrays (array layout).
- Information on the target samples hybridized to the arrays.
- See *Minimum Information About a Microarray Experiment – MIAME* – standards.

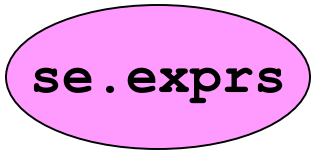
Biological metadata

- Biological attributes that can be applied to the experimental data.
- E.g. for genes
  - chromosomal location;
  - gene annotation (LocusLink, GO);
  - relevant literature (PubMed).
- Biological metadata sources are large, complex, evolving rapidly, and typically distributed via the WWW.

exprSet class

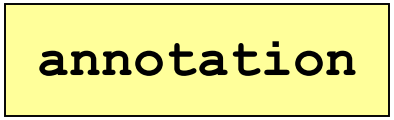


Matrix of expression measures, genes x samples



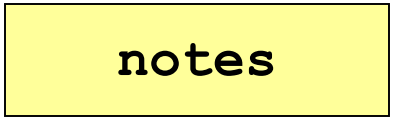
Matrix of SEs for expression measures, genes x samples

Sample level covariates, instance of class **phenoData**



Name of annotation data

MIAME information



Any notes

`marrayRaw` class

Pre-normalization intensity data for a batch of arrays

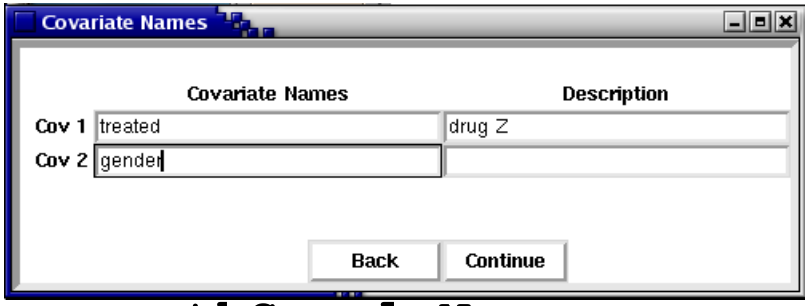
`maRf`	`maGf`	Matrix of red and green foreground intensities
`maRb`	`maGb`	Matrix of red and green background intensities
`maW`		Matrix of spot quality weights
`maLayout`		Array layout parameters - `marrayLayout`
`maGnames`		Description of spotted probe sequences - `marrayInfo`
`maTargets`		Description of target samples - `marrayInfo`
`maNotes`		Any notes

AffyBatch class

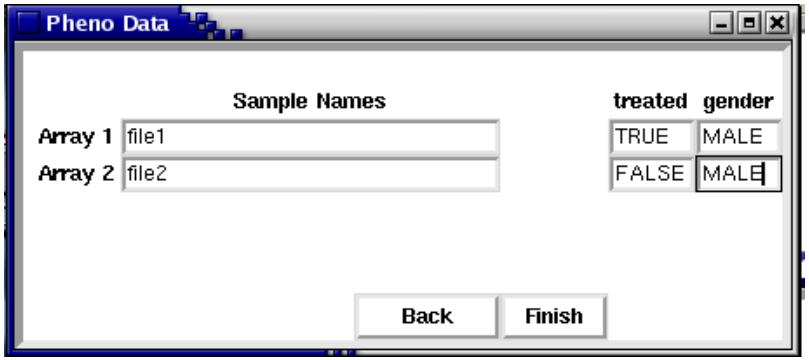
Probe-level intensity data for a batch of arrays (same CDF)

cdfName		Name of **CDF** file for arrays in the batch
nrow	**ncol**	Dimensions of the array
exprs	**se.exprs**	Matrices of probe-level intensities and SEs rows → probe cells, columns → arrays.
phenoData		Sample level covariates, instance of class **phenoData**
annotation		Name of annotation data
description		MIAME information
notes		Any notes

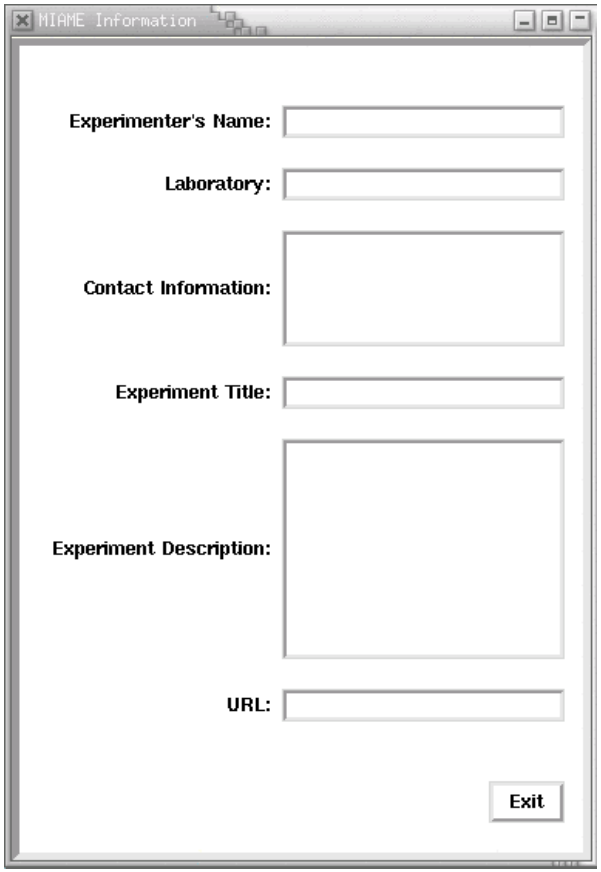
Reading in `phenoData`



`tkSampleNames`



`tkphenoData`



`tkMIAME`

Pedagogy

Extensive documentation and training resources for R and Bioconductor are available on the WWW.

- R manuals and tutorials are available from the R website.
- R help system
  - detailed on-line documentation, available in text, HTML, PDF, and LaTeX formats;
  - e.g. **`help(genefilter), ?pubmed`**.
- R demo system
  - user-friendly interface for running demonstrations of R scripts;
  - e.g. **`demo(marrayPlots)`**, **`demo(affy)`**.
- Bioconductor short courses
  - modular training segments on software and statistical methodology;
  - lectures and computer labs available on WWW for self-instruction.

Vignettes

- Bioconductor has adopted a new documentation paradigm, the vignette.
- A vignette is an executable document consisting of a collection of documentation text and code chunks.
- Vignettes form dynamic, integrated, and reproducible statistical documents that can be automatically updated if either data or analyses are changed.
- Vignettes can be generated using the **`Sweave`** function from the R **`tools`** package.

Vignettes

- Each Bioconductor package contains at least one vignette, located in the `doc` subdirectory of an installed package and accessible from the help browser.
- Vignettes provide task-oriented descriptions of the package's functionality and can be used interactively.
- Vignettes are available separately from the Bioconductor website or as part of the packages.

Vignettes

- Tools are being developed for managing and using this repository of step-by-step tutorials
  - **`Biobase`**: **`openVignette`** – Menu of available vignettes and interface for viewing vignettes (PDF).
  - **`tkWidgets`**: **`vExplorer`** – Interactive use of vignettes.
  - **`reposTools`**.

Sweave

- The Sweave system allows the generation of integrated statistical documents intermixing text, code, and code output (textual and graphical).
- Functions are available in the R `tools` package.
- See `? Sweave` and manual www.ci.tuwien.ac.at/~leisch/Sweave/

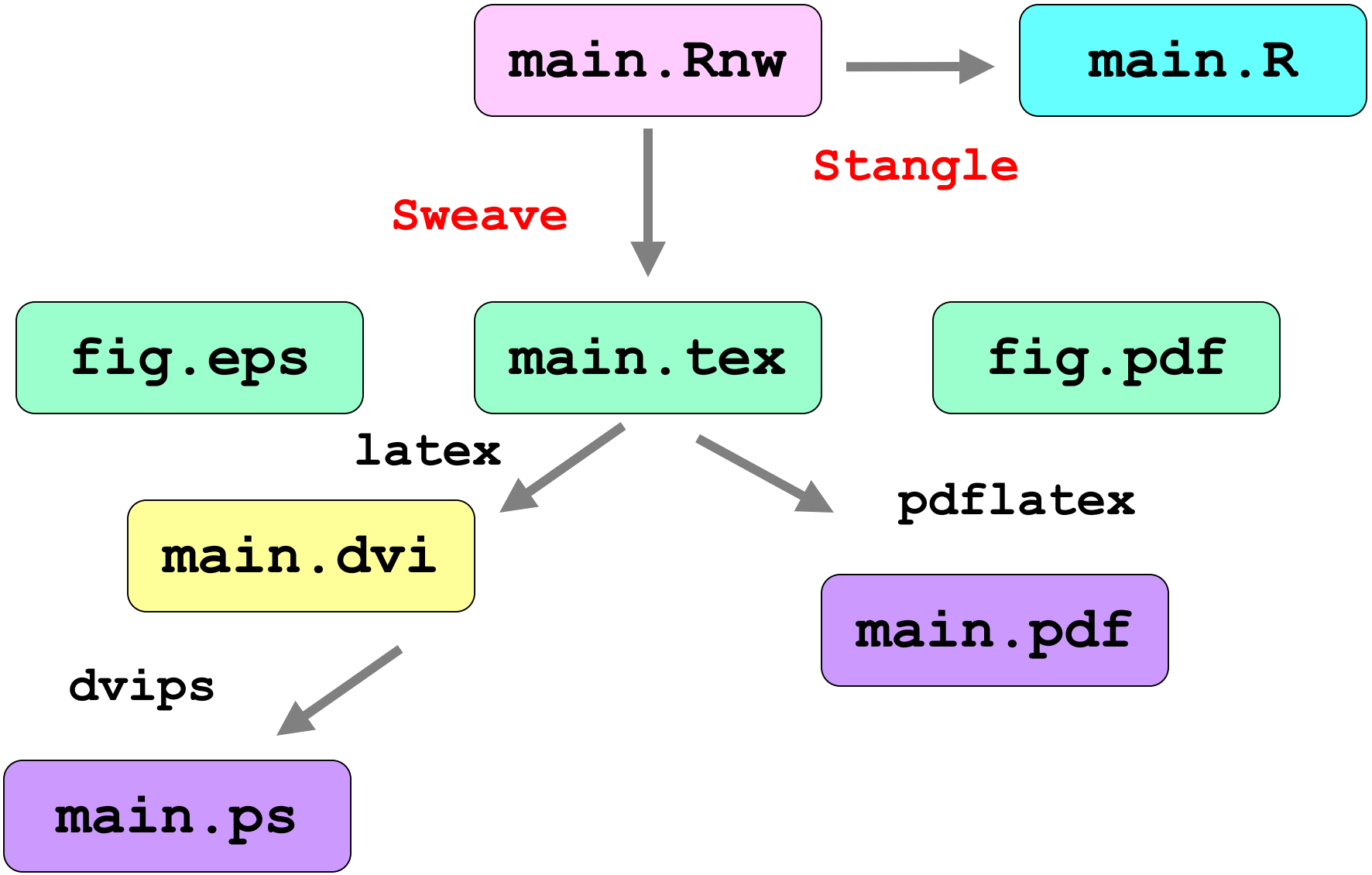
Sweave input

- Input: a text file which consists of a sequence of code and documentation chunks, or segments (noweb file).
  - Documentation chunks
    - start with @
    - can be text in a markup language like LaTeX.
  - Code chunks
    - start with <<*name*>>=
    - can be R or S-Plus code.
  - File extension: **`.rnw`**, **`.Rnw`**, **`.snw`**, **`.Snw`**.

Sweave output

- Output: a single document, e.g., **`.tex`** file or **`.pdf`** file containing
  - the documentation text,
  - the R code,
  - the code output: text and graphs.
- The document can be automatically regenerated whenever the data, code, or documentation text change.
- **`Stangle`** or **`tangleToR`**: extract only the code.

Sweave


main.Rnw
main.R
Stangle
Sweave
fig.eps
main.tex
fig.pdf
latex
pdflatex
main.dvi
main.pdf
dvips
main.ps

Annotation

`annotate` package

- One of the largest challenges in analyzing genomic data is associating the experimental data with the available biological metadata, e.g., sequence, gene annotation, chromosomal maps, literature.

- Bioconductor provides two main packages for this purpose:
  - `annotate` (end-user);
  - `AnnBuilder` (developer).

WWW resources

- Nucleotide databases: e.g. GenBank.
- Gene databases: e.g. LocusLink, UniGene.
- Protein sequence and structure databases: e.g. SwissProt, Protein DataBank (PDB).
- Literature databases: e.g. PubMed, OMIM.
- Chromosome maps: e.g. NCBI Map Viewer.
- Pathways: e.g. KEGG.
- Entrez is a search and retrieval system that integrates information from databases at NCBI (National Center for Biotechnology Information).

`annotate`: matching IDs

Important tasks

- Associate manufacturers or in-house probe identifiers to other available identifiers.

  E.g.

  Affymetrix IDs → LocusLink LocusID

  Affymetrix IDs → GenBank accession number.
- Associate probes with biological data such as chromosomal position, pathways.
- Associate probes with published literature data via PubMed (need PMID).

`annotate`: matching IDs

Affymetrix identifier HGU95A chips	`"41046_s_at"`
LocusLink, LocusID	`"9203"`
GenBank accession #	`"X95808"`
Gene symbol	`"ZNF261"`

PubMed, PMID	`"10486218"`
	`"9205841"`
	`"8817323"`

Chromosomal location	`"X", "Xq13.1"`

Annotation data packages

- The Bioconductor project provides packages that contain only data.
- These data packages are built using **`AnnBuilder`**.
- They can be downloaded from the Bioconductor website and also using **`update.packages`**.*** installDataPackage.
- Data packages contain many different mappings to interesting data.
  - Mappings between Affy IDs and other probe IDs: **`hgu95a`** for HGU95A GeneChip series, also, **`hgu133a`**, **`hu6800`**, **`mgu74a`**, **`rgu34a`**.
  - Affy CDF data packages.
- The packages are updated and expanded regularly as updated and new data become available.

`annotate`: matching IDs

- Much of what `annotate` does relies on matching symbols.
- This is basically the role of a hash table in most programming languages.
- In R, we rely on environments.
- The annotation data packages provide R environment objects containing key and value pairs for the mappings between two sets of probe identifiers.
- Keys can be accessed using the R `ls` function.
- Matching values in different environments can be accessed using the `get` or `multiget` functions.

annotate: matching IDs

```
> library(hgu95a)
> get("41046_s_at", env = hgu95aACCNUM)
[1] "X95808"
> get("41046_s_at", env = hgu95aLOCUSID)
[1] "9203"
> get("41046_s_at", env = hgu95aSYMBOL)
[1] "ZNF261"
> get("41046_s_at", env = hgu95aGENENAME)
[1] "zinc finger protein 261"
> get("41046_s_at", env = hgu95aSUMFUNC)
[1] "Contains a putative zinc-binding
  motif (MYM)|Proteome"
> get("41046_s_at", env = hgu95aUNIGENE)
[1] "Hs.9568"
```

annotate: matching IDs

```
> get("41046_s_at", env = hgu95aCHR)
[1] "X"
> get("41046_s_at", env = hgu95aCHRLOC)
[1] "66457019@X"
> get("41046_s_at", env = hgu95aCHRORI)
[1] "-@X"
> get("41046_s_at", env = hgu95aMAP)
[1] "Xq13.1”
> get("41046_s_at", env = hgu95aPMID)
[1] "10486218" "9205841"  "8817323"
> get("41046_s_at", env = hgu95aGO)
[1] "GO:0003677" "GO:0007275"
```

`annotate`: matching IDs

- Instead of relying on the general R functions for environments, new user-friendly functions have been written for accessing and working with specific identifiers.

- E.g. **`getGO`**, **`getGOdesc`**, **`getLL`**, **`getPMID`**, **`getSYMBOL`**.

annotate: matching IDs

```
> getSYMBOL("41046_s_at",data="hgu95a")
  41046_s_at
    "ZNF261"
> gg<- getGO("41046_s_at",data="hgu95a")
> getGOdesc(gg, "MF")
  $"c("GO:0003677", "GO:0007275")"
  [1] "DNA binding"
> getLL("41046_s_at",data="hgu95a")
  41046_s_at
      9203
> getPMID("41046_s_at",data="hgu95a")
  $"41046_s_at"
  [1] 10486218  9205841  8817323
```

`annotate`: querying databases

The `annotate` package provides tools for

- Searching and processing information from various WWW biological databases
  - GenBank,
  - LocusLink,
  - PubMed.
- Regular expression searching of PubMed abstracts.
- Generating nice HTML reports of analyses, with links to biological databases.

`annotate`: WWW queries

- Functions for querying WWW databases from R rely on the **`browseURL`** function

```
browseURL("www.r-project.org")
```

- The **`XML`** package is used to parse query results.

`annotate`: querying GenBank

www.ncbi.nlm.nih.gov/Genbank/index.html

- Given a vector of GenBank accession numbers or NCBI UIDs, the **`genbank`** function
  - opens a browser at the URLs for the corresponding GenBank queries;
  - returns an **`XMLdoc`** object with the same data.

```
genbank("X95808",disp="browser")
```

http://www.ncbi.nih.gov/entrez/query.fcgi?tool=bioconductor&cmd=Search&db=Nucleotide&term=X95808

```
genbank(1430782,disp="data",
    type="uid")
```

`annotate`: querying LocusLink

www.ncbi.nlm.nih.gov/LocusLink/

- **`locuslinkByID`**: given one or more LocusIDs, the browser is opened at the URL corresponding to the first gene.

  ```
  locuslinkByID("9203")
  ```
  http://www.ncbi.nih.gov/LocusLink/LocRpt.cgi?l=9203

- **`locuslinkQuery`**: given a search string, the results of the LocusLink query are displayed in the browser.

  ```
  locuslinkQuery("zinc finger")
  ```
  http://www.ncbi.nih.gov/LocusLink/list.cgi?Q=zinc finger&ORG=Hs&V=0

`annotate`: querying PubMed

www.ncbi.nlm.nih.gov

- For any gene there is often a large amount of data available from PubMed.
- The **`annotate`** package provides the following tools for interacting with PubMed
  - **`pubMedAbst`**: a class structure for PubMed abstracts in R.
  - **`pubmed`**: the basic engine for talking to PubMed.

annotate: pubMedAbst class

Class structure for storing and processing PubMed abstracts in R

- **pmid**
- **authors**
- **abstText**
- **articleTitle**
- **journal**
- **pubDate**
- **abstUrl**

`annotate`: high-level tools for querying PubMed

- **`pm.getabst`**: download the specified PubMed abstracts (stored in XML) and create a list of **`pubMedAbst`** objects.
- **`pm.titles`**: extract the titles from a list of PubMed abstracts.
- **`pm.abstGrep`**: regular expression matching on the abstracts.

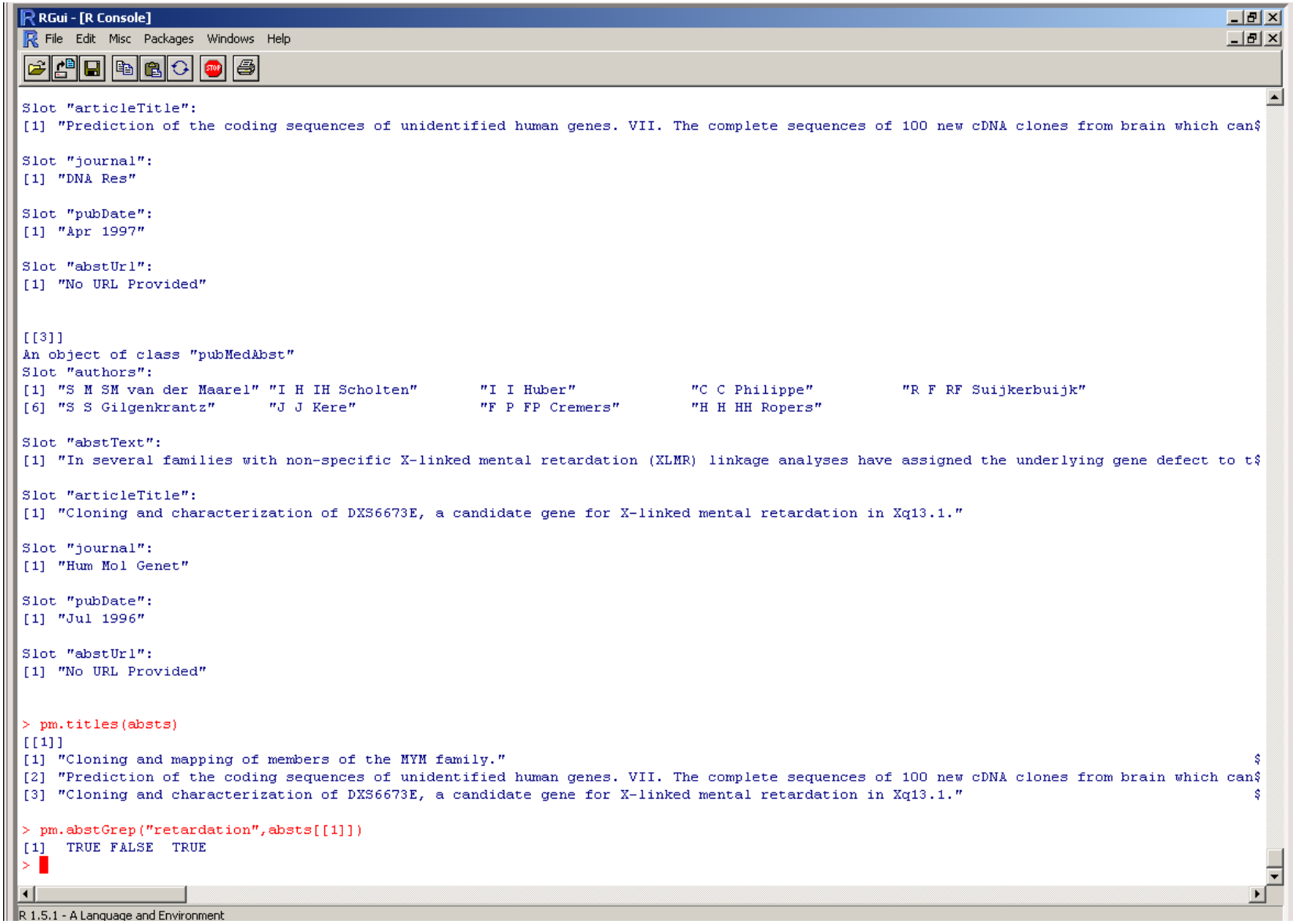
annotate: PubMed example

```
pmid <-get("41046_s_at", env=hgu95aPMID)
pubmed(pmid, disp="browser")
```

http://www.ncbi.nih.gov/entrez/query.fcgi?tool=bioconductor&cmd=Retrieve&db=PubMed&list_uids=10486218%2c9205841%2c8817323

```
absts <- pm.getabst("41046_s_at",
  base="hgu95a")
pm.titles(absts)
pm.abstGrep("retardation",absts[[1]])
```

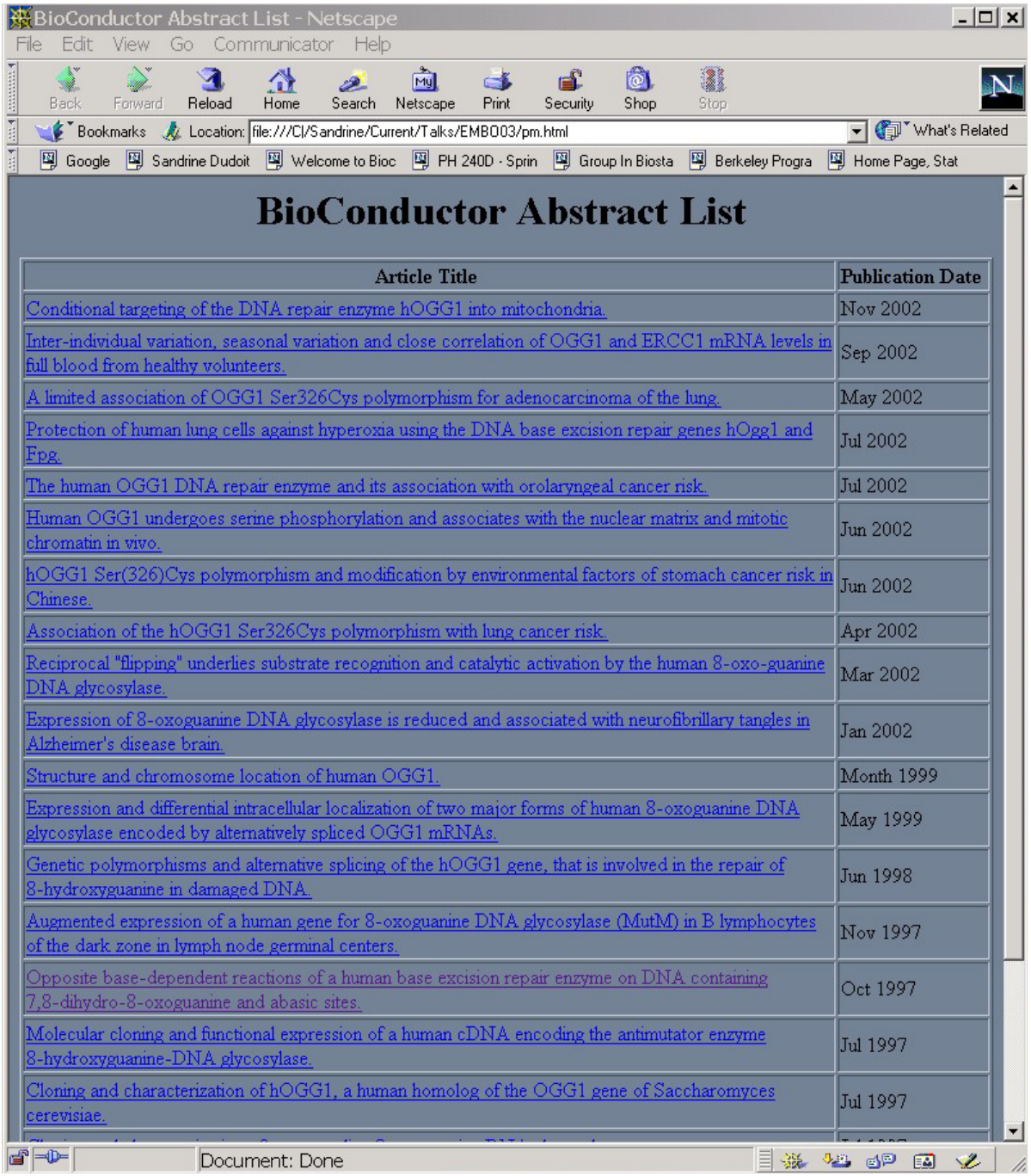
annotate: PubMed example



`annotate`: PubMed HTML report

- The new function **`pmAbst2HTML`** takes a list of **`pubMedAbst`** objects and generates an HTML report with the titles of the abstracts and links to their full page on PubMed.

```
pmAbst2HTML(absts[[1]],filename="pm.html")
```

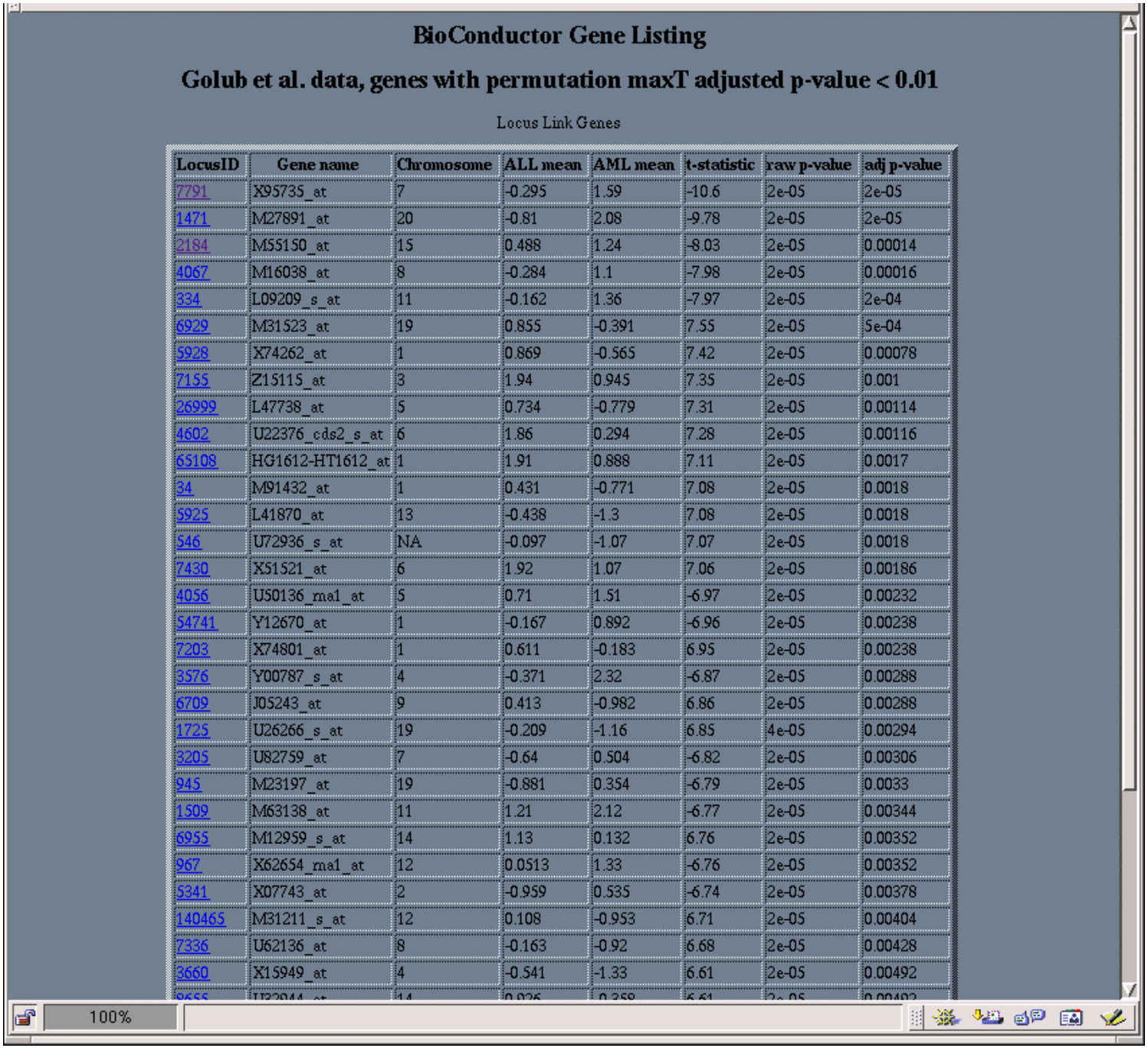


`pmAbst2html` function from **`annotate`** package

pm.html

`annotate`: analysis reports

- A simple interface, `ll.htmlpage`, can be used to generate an HTML report of analysis results.
- The page consists of a table with one row per gene, with links to LocusLink.
- Entries can include various gene identifiers and statistics.



BioConductor Gene Listing

Golub et al. data, genes with permutation maxT adjusted p-value < 0.01

Locus Link Genes

LocusID	Gene name	Chromosome	ALL mean	AML mean	t-statistic	raw p-value	adj p-value
7791	X95735_at	7	-0.295	1.59	-10.6	2e-05	2e-05
1471	M27891_at	20	-0.81	2.08	-9.78	2e-05	2e-05
2184	M55150_at	15	0.488	1.24	-8.03	2e-05	0.00014
4067	M16038_at	8	-0.284	1.1	-7.98	2e-05	0.00016
334	L09209_s_at	11	-0.162	1.36	-7.97	2e-05	2e-04
6929	M31523_at	19	0.855	-0.391	7.55	2e-05	5e-04
5928	X74262_at	1	0.869	-0.565	7.42	2e-05	0.00078
7155	Z15115_at	3	1.94	0.945	7.35	2e-05	0.001
26999	L47738_at	5	0.734	-0.779	7.31	2e-05	0.00114
4602	U22376_cds2_s_at	6	1.86	0.294	7.28	2e-05	0.00116
65108	HG1612-HT1612_at	1	1.91	0.888	7.11	2e-05	0.0017
34	M91432_at	1	0.431	-0.771	7.08	2e-05	0.0018
5925	L41870_at	13	-0.438	-1.3	7.08	2e-05	0.0018
546	U72936_s_at	NA	-0.097	-1.07	7.07	2e-05	0.0018
7430	X51521_at	6	1.92	1.07	7.06	2e-05	0.00186
4056	U50136_rna1_at	5	0.71	1.51	-6.97	2e-05	0.00232
54741	Y12670_at	1	-0.167	0.892	-6.96	2e-05	0.00238
7203	X74801_at	1	0.611	-0.183	6.95	2e-05	0.00238
3576	Y00787_s_at	4	-0.371	2.32	-6.87	2e-05	0.00288
6709	J05243_at	9	0.413	-0.982	6.86	2e-05	0.00288
1725	U26266_s_at	19	-0.209	-1.16	6.85	4e-05	0.00294
3205	U82759_at	7	-0.64	0.504	-6.82	2e-05	0.00306
945	M23197_at	19	-0.881	0.354	-6.79	2e-05	0.0033
1509	M63138_at	11	1.21	2.12	-6.77	2e-05	0.00344
6955	M12959_s_at	14	1.13	0.132	6.76	2e-05	0.00352
967	X62654_rna1_at	12	0.0513	1.33	-6.76	2e-05	0.00352
5341	X07743_at	2	-0.959	0.535	-6.74	2e-05	0.00378
140465	M31211_s_at	12	0.108	-0.953	6.71	2e-05	0.00404
7336	U62136_at	8	-0.163	-0.92	6.68	2e-05	0.00428
3660	X15949_at	4	-0.541	-1.33	6.61	2e-05	0.00492
9655	U22944_at	14	0.926	0.268	6.61	2e-05	0.00492

100%

`ll.htmlpage` function from **`annotate`** package

genelist.html

`annotate`: `chromLoc` class

Location information for one gene

- **`chrom`**: chromosome name.
- **`position`**: starting position of the gene in bp.
- **`strand`**: chromosome strand +/-.

annotate: chromLocation class

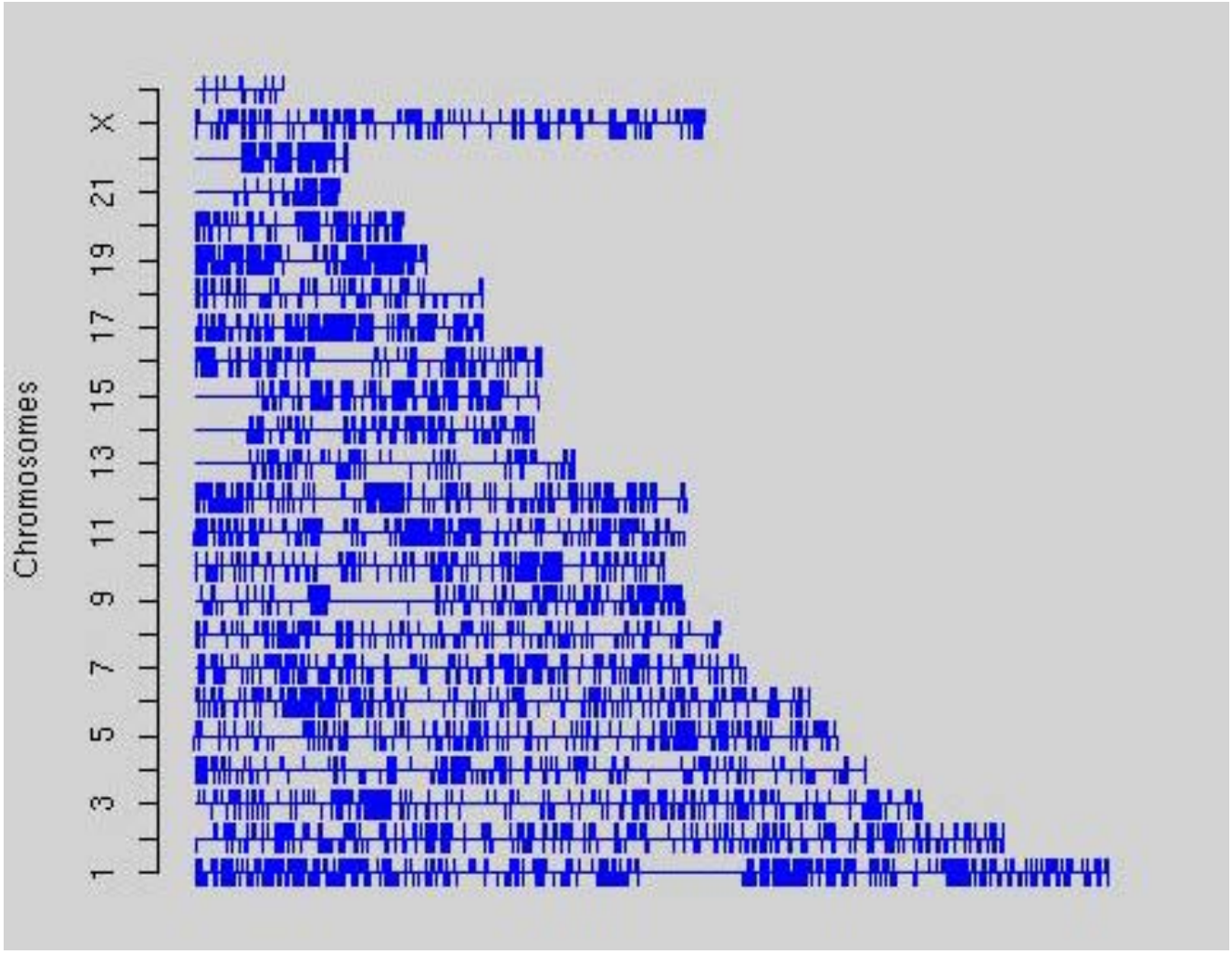
Location information for a set of genes

- **species:** species that the genes correspond to.
- **datSource:** source of the gene location data.
- **nChrom:** number of chromosomes for the species.
- **chromNames:** chromosome names.
- **chromLocs:** starting position of the genes in bp.
- **chromLengths:** length of each chromosome in bp.
- **geneToChrom:** hash table translating gene IDs to location.

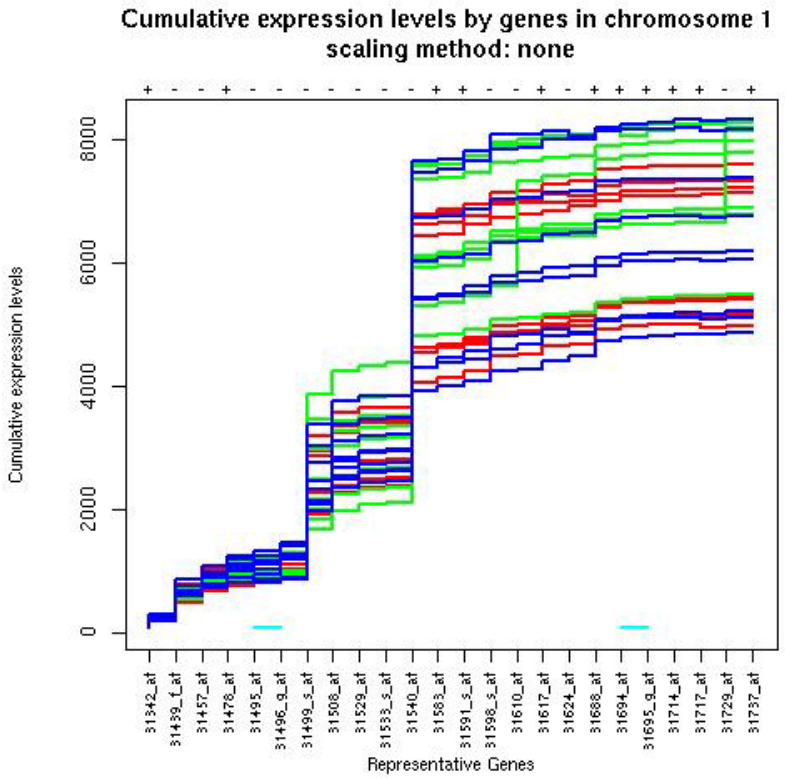
Function **buildChromClass**.

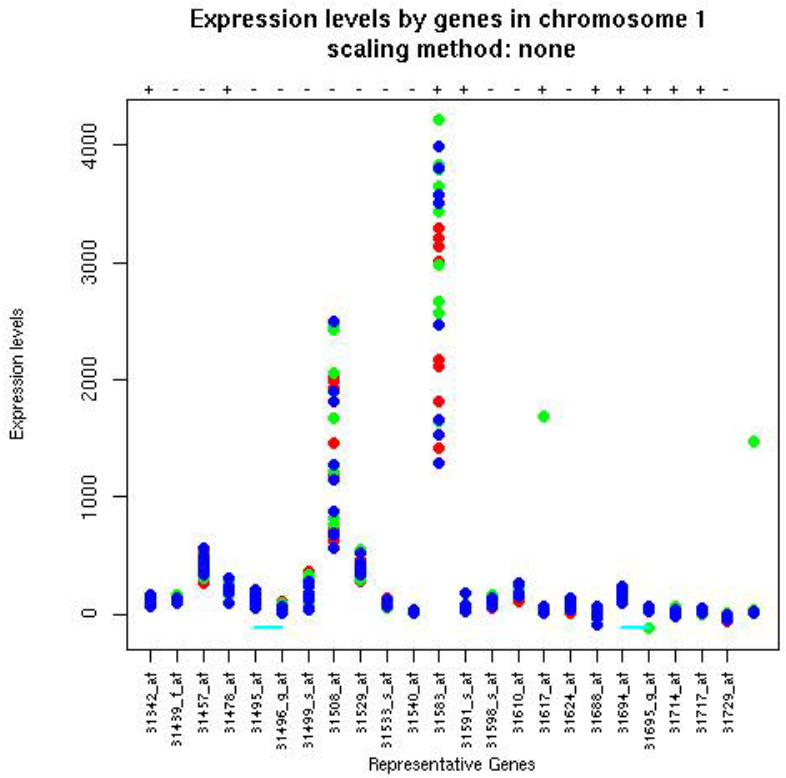
Visualization

geneplotter: cPlot

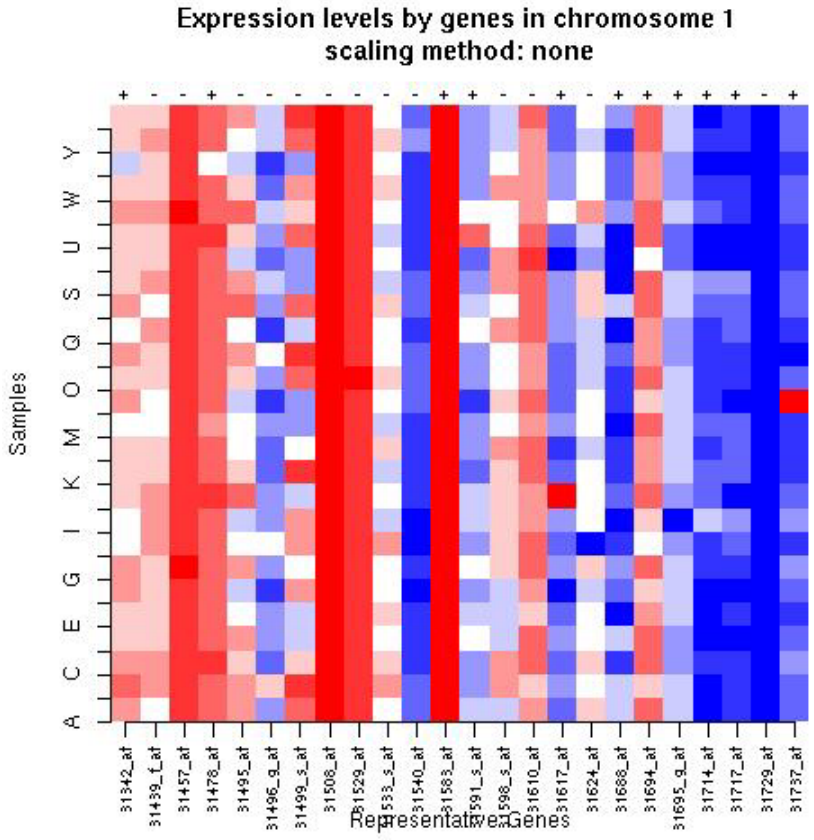


geneplotter: alongChrom

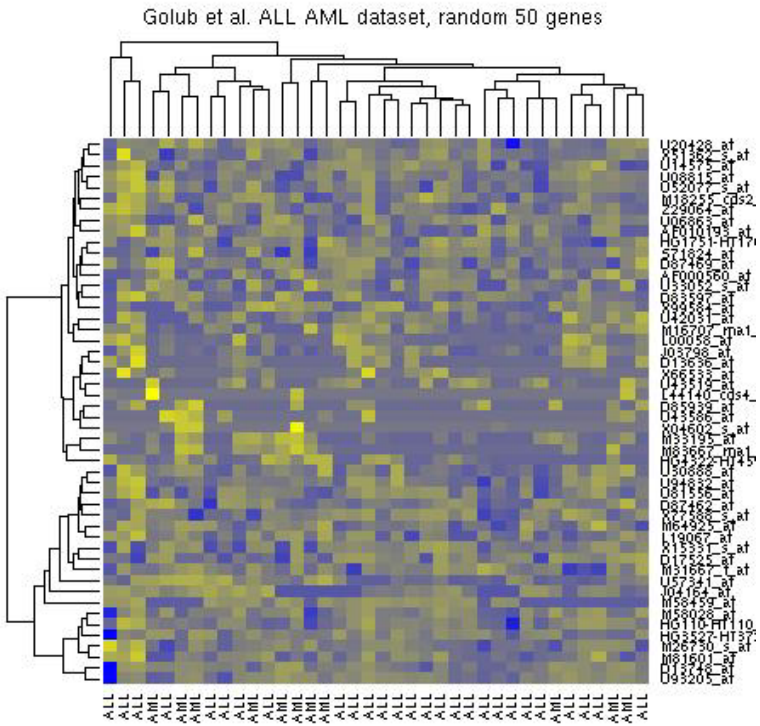


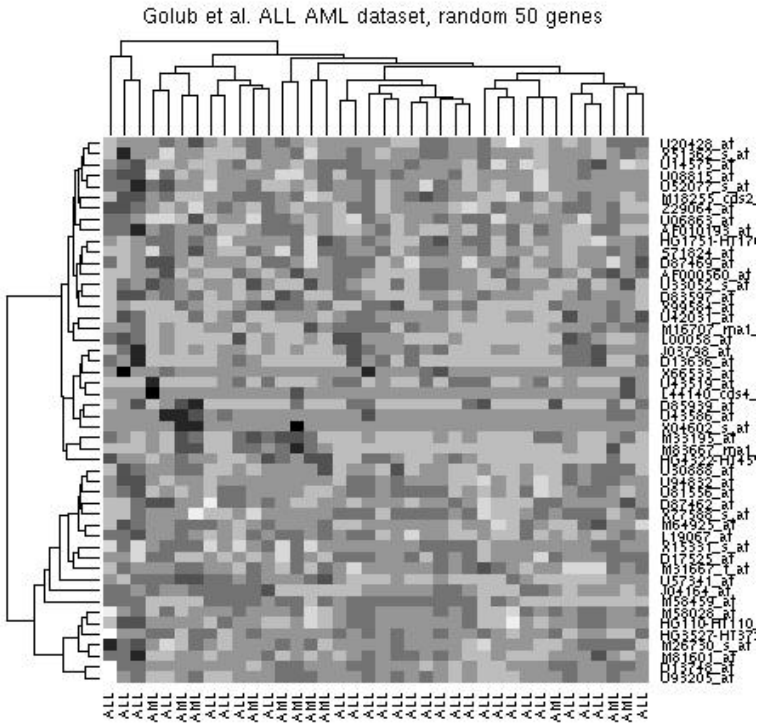


geneplotter: alongChrom



mva: heatmap


Golub et al. ALL AML dataset, random 50 genes


Golub et al. ALL AML dataset, random 50 genes