# Package 'OMICsPCA'

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Type Package

**Title** An R package for quantitative integration and analysis of multiple omics assays from heterogeneous samples

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Depends R (>= 3.5.0), OMICsPCAdata

Suggests knitr, RUnit, BiocGenerics

**Description** OMICsPCA is an analysis pipeline designed to integrate multi OMICs experiments done on various subjects (e.g. Cell lines, individuals), treatments (e.g. disease/control) or time points and to analyse such integrated data from various various angles and perspectives. In it's core OMICsPCA uses Principal Component Analysis (PCA) to integrate multiomics experiments from various sources and thus has ability to over data insufficiency issues by using the ingegrated data as representatives. OMICsPCA can be used in various application including analysis of overall distribution of OMICs assays across various samples /individuals /time points; grouping assays by user-defined conditions; identification of source of variation, similarity/dissimilarity between assays, variables or individuals.

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**Encoding** UTF-8

biocViews ImmunoOncology, MultipleComparison, PrincipalComponent, DataRepresentation, Workflow, Visualization, DimensionReduction, Clustering, BiologicalQuestion, EpigeneticsWorkflow, Transcription, GeneticVariability, GUI, BiomedicalInformatics, Epigenetics, FunctionalGenomics, SingleCell

**Imports** HelloRanges, fpc, stats, MultiAssayExperiment, pdftools, methods, grDevices, utils,clValid, NbClust, cowplot, rmarkdown, kableExtra, rtracklayer, IRanges, GenomeInfoDb, reshape2, ggplot2, factoextra, rgl, corrplot, MASS, graphics, FactoMineR, PerformanceAnalytics, tidyr, data.table, cluster, magick VignetteBuilder knitr LazyData TRUE RoxygenNote 6.0.1 git\_url https://git.bioconductor.org/packages/OMICsPCA git\_branch RELEASE\_3\_13 git\_last\_commit\_d08002d git\_last\_commit\_date 2021-05-19 Date/Publication 2021-10-14

# **R** topics documented:

analyse_individuals	2
analyse_integrated_individuals	3
analyse_integrated_variables	5
analyse_variables	6
chart_correlation	7
cluster	8
cluster_boxplot	10
cluster_parameters	
create_group	
descriptor	14
extract	
extract_assay	
integrate_pca	
integrate_variables	
intersect	
merge_cells	20
F	21
plot_density_3D	22
prograded_density	23
F	24
prepare_dataset	25
	27

# Index

analyse\_individuals

Quick analysis and visualization of the individuals/annotations/rows (e.g. Tss, gene) from integrated Assay.

# Description

This function displays scatterplot of the individuals from integrated assays. This allows the display of scatterplots both in 2D and 3D.

#### Usage

```
analyse_individuals(name,Assay, choice, PC, group,
groupinfo = NULL, ...)
```

#### Arguments

name	Name of the "PCA" object containing the assay data
Assay	Name of an Assay
choice	<pre>1 = 2D scatterplot on selected PCs through PC argument. 2 = 3D scatterplot on selected PCs through PC argument</pre>
	for details of the choices see the vignette
PC	A vector of numbers corresponding to principal components
group	A vector of group names. Required only if choice = $2$
groupinfo	output of create_group, or similar object containing group information.
	additional arguments allowed to base function "plot3d" of package "rgl"

# Value

Displays various plots and tables as per the combination of input arguments

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Examples

```
PCAlist <- integrate_variables(Assays = c("H2az","H3k4me1",
 "H3k9ac"), name = multi_assay,
groups = c("WE","RE"), groupinfo = groupinfo_ext,
scale.unit = FALSE, graph = FALSE)
analyse_individuals(name = PCAlist,
Assay = "H3k9ac", groupinfo = groupinfo_ext,
choice = 1, PC = c(1,2))
```

analyse\_integrated\_individuals

*Quick analysis and visualization of the individuals/annotations/rows (e.g. Tss, gene)* 

#### Description

This function works similarly as "analyse\_individuals()". The only extra argument needs to be supplied here is "start\_end", (if choice = 1.) returned by "integrate\_pca()"

#### Usage

```
analyse_integrated_individuals(
name, choice = 1, geom = "point",
PC = c(1,2), groupinfo = NULL, ...)
```

#### Arguments

name	Name of the "PCA" object containing the integrated PCA.
choice	<pre>1 = 2D scatterplot on selected PCs through PC argument. 2 = 3D scatterplot on selected PCs through PC argument</pre>
	for details of the choices see the vignette
geom	Used when choice = 1. Avalilable options are : "point", "text", c("point", "text"). default = "point"
PC	A vector of numbers corresponding to principal components
groupinfo	same as integrate_variables()
	additional arguments allowed to base function "plot3d" of package "rgl"

#### Value

Displays various plots and tables as per the combination of input arguments

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
"H3k9ac"), name = multi_assay, mergetype = 2,
exclude = exclude,
groupinfo = groupinfo_ext, graph = FALSE)</pre>
```

```
name = int_PCA$int_PCA
```

```
analyse_integrated_individuals(
name = name,
choice = 2, PC = c(1,2,3),
col = c("RED", "BLACK","GREEN"),
groupinfo = groupinfo_ext)
```

```
analyse_integrated_variables
```

*Quick analysis and visualization of the integrated Assays by integrate() function.* 

# Description

This function is designed for a quick analysis and visualization of the data integrated by "integrate-Assays()"" function. It works similarly as analyse\_variables. In addition to that it takes an additinal argument "start\_end" returned by "integrateAssays()".

#### Usage

```
analyse_integrated_variables(start_end = start_end, Assay = "all",
name, choice, title = NULL, PC = 1,
var_type = "contrib", ...)
```

# Arguments

start_end	a list as returned by "integrate_pca()"
Assay	Name of an Assay. default = "all"
name	Name of the "PCA" object returned by "integrate_pca()"
choice	<pre>1 = variance barplot 2 = Loadings of cell lines on selected PCs 3 = hCorrelation matrix 4 = Squarred loadings of Cell lines on a PC 5 = Contribution of Cell lines on a PC for details of the choices see the vignette</pre>
title	title of plots
PC	The PC on which the plots will be drawn. used in choice 4 and 5. default = $1$
var_type	Used when choice = 2. available options are: "coord", "cos2" and "contrib"
	additional arguments allowed to base functions "used in various choices. See vignettes for details.

#### Value

Displays various plots and tables as per the combination of input arguments

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Examples

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
    "H3k9ac"), name = multi_assay, mergetype = 2,
exclude = exclude, graph = FALSE)
start_end = int_PCA$start_end
name = int_PCA$int_PCA
analyse_integrated_variables(start_end = start_end, name = name,
choice = 1, title = "variance barplot", Assay = 1, addlabels = TRUE)</pre>
```

```
analyse_variables Quick analysis and visualization of the integrated Assays by integrate()
function.
```

# Description

This function is designed for a quick analysis and visualization of the data integrated by integrate() function. It takes 3 compulsory arguments name, Assay and choice. The type of analysis should be selected through the "choice" argument. This function acts as a wrapper around a collection of functions of package "factoextra" and "corrplot" and thus can take additional arguments specific and non-conflicting to such functions.

#### Usage

```
analyse_variables(name, Assay, choice, title = NULL, PC = 1,
var_type = "contrib", ...)
```

#### Arguments

name	Name of the "PCA" object
Assay	Name of an Assay.
choice	<pre>1 = variance barplot 2 = Loadings of cell lines on selected PCs 3 = hCorrelation matrix 4 = Squarred loadings of Cell lines on a PC 5 = Contribution of Cell lines on a PC</pre>
	for details of the choices see the vignette
title	Title of the plot. default = NULL
PC	used in choice = 4 and 5. Indicates the PC on which choice 4 and 5 will depend. default = $1$

6

var_type	used in choice = 2. available options are: "coord", "cos2" and "contrib". default = "contrib"
	extra arguments passed through the functions imported from various packages. See vignettes for details.

#### Value

Displays various plots and tables as per the combination of input arguments

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

#### Examples

```
PCAlist <- integrate_variables(Assays = c("H2az","H3k4me1",
"H3k9ac"), name = multi_assay,
groups = c("WE","RE"), groupinfo = groupinfo_ext,
scale.unit = FALSE, graph = FALSE)
analyse_variables(name = PCAlist, Assay = "H2az", choice = 1,
title = "variance barplot", addlabels = TRUE)
```

	chart_correlation	Pairwise correlation,	scatter plot and hi	istogram on selected	groups.
--	-------------------	-----------------------	---------------------	----------------------	---------

# Description

This function creates pairwise correlation plots and tables; scatter plots and histograms on selected groups. The type of results should be passed through the argument 'choice'. This is a wrapper on various functions and thus can take additional and non-conflicting arguments specific to them.

#### Usage

```
chart_correlation(name, Assay, groups, choice,
groupinfo = NULL,
...)
```

#### Arguments

name	Name of the "MultiAssayExperiment" object containing the assay data
Assay	Name of an assay
groups	Name of all or subset of groups

choice	"table" = correlation table
	"scatter" = scatterplot of each pair of columns/variables of selected groups
	"hist" = histogram of each column/variables
	"all" = all of 1,2,3 together
groupinfo 	output of create_group or similar object. additional arguments for base functions "chart.Correlation" and "pairs"

# Value

Displays various plots and tables as per the combination of input arguments

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Examples

```
chart_correlation(name = multi_assay,
Assay = "H2az", groupinfo = groupinfo_ext,
groups = "WE", choice = "table")
```

cluster

Cluster data points

# Description

Clustering of data points using various algorithms

# Usage

```
cluster(name, n = NULL, graph = NULL, choice, title = NULL, ...)
```

# Arguments

name	name of the dataframe or matrix object containing the coordinates of data points. The output of "extract()" may be directly put here.
n	Number of clusters
graph	logical. Plots the clusterplot on first 2 dimensions if set TRUE
choice	Clustering algorithm to use. Available choices are: "density", "kmeans", "pam"
title	Title of the plot
	additional non-conflicting arguments to cluster functions

#### cluster

#### Value

returns a list containing the cluster and plot information

#### Author(s)

Subhadeep Das

#### References

Martin Ester, Hans-Peter Kriegel, Joerg Sander, Xiaowei Xu (1996). A Density-Based Algorithm for Discovering Clusters in Large Spatial Databases with Noise. Institute for Computer Science, University of Munich. Proceedings of 2nd International Conference on Knowledge Discovery and Data Mining (KDD-96).

Forgy, E. W. (1965). Cluster analysis of multivariate data: efficiency vs interpretability of classifications. Biometrics, 21, 768–769.

Hartigan, J. A. and Wong, M. A. (1979). Algorithm AS 136: A K-means clustering algorithm. Applied Statistics, 28, 100–108. doi: 10.2307/2346830.

Lloyd, S. P. (1957, 1982). Least squares quantization in PCM. Technical Note, Bell Laboratories. Published in 1982 in IEEE Transactions on Information Theory, 28, 128–137.

MacQueen, J. (1967). Some methods for classification and analysis of multivariate observations. In Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability, eds L. M. Le Cam & J. Neyman, 1, pp. 281–297. Berkeley, CA: University of California Press.

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
"H3k9ac"),
groupinfo = groupinfo,
name = multi_assay, mergetype = 2,
exclude = exclude, graph = FALSE)
name = int_PCA$int_PCA
data <- extract(name = name, PC = c(1:4),
groups = c("WE", "RE"), integrated = TRUE,
rand = 300, groupinfo = groupinfo_ext)
clusters <- cluster(name = data, n = 2,
choice = "kmeans",
title = "kmeans on 2 clusters")
```

cluster\_boxplot Comparis

# Description

boxplot of the assay values in each cluster

#### Usage

```
cluster_boxplot(name, Assay, clusterobject, clustercolumn, choice = NULL)
```

# Arguments

#### name

Assay	name of the assay which is to be compared
clusterobject	a cluster object returned by the cluster() function
clustercolumn	the column in the clusterobject containing the group membership information
choice	type of graph

# Value

a ggplot object

# Author(s)

Subhadeep Das

# Examples

```
bp <- cluster_boxplot(name = multi_assay,
Assay = "H2az", clusterobject = clustered_data,
clustercolumn = 5)
```

# bp

cluster\_parameters Detection of algorithm and number of clusters

# Description

Detection of appropriate clustering algorithm and cluster number for given data using "clvalid" and "NbClust" in background for cluster validation.

# Usage

```
cluster_parameters(name, comparisonAlgorithm = "clValid",
optimal = FALSE, n = 2:6,
clusteringMethods = c("kmeans", "pam"),
validationMethods = c("internal", "stability"),
distance = "euclidean", ...)
```

# Arguments

name	dataframe returned by "extract()".
optimal	logical. If TRUE, returns a dataframe of optimal results
n	a vector of numbers corresponding to the number of clusters to be tested or validated
comparisonAlgo	rithm
	2 choices available: "clValid" or "NbClust"
clusteringMeth	
	a vector of single or multiple names of clustering algorithms. available choices are:
	1) if comparisonAlgorithm = "clValid" : "hierarchical", "kmeans", "diana", "fanny", "som", "model", "sota", "pam", "clara" and "agnes"
	2) if comparisonAlgorithm = "NbClust" : "ward.D", "ward.D2", "single", "com- plete", "average", "mcquitty", "median", "centroid", "kmeans".
validationMeth	ods
	name of the method to validate clusters. Available options (one or more):
	1) if comparisonAlgorithm = "clValid" then one of:
	"kl", "ch", "hartigan", "ccc", "scott", "marriot", "trcovw", "tracew", "friedman", "rubin", "cindex", "db", "silhouette", "duda", "pseudot2", "beale", "ratkowsky", "ball", "ptbiserial", "gap", "frey", "mcclain", "gamma", "gplus", "tau", "dunn", "hubert", "sdindex", "dindex", "sdbw", "all" (all indices except GAP, Gamma, Gplus and Tau), "alllong" (all indices with Gap, Gamma, Gplus and Tau in- cluded).
	2) if comparisonAlgorithm = "NbClust" then one or more of: "internal", "stability", and "biological"
distance	metric used to calculate distance matrix. options: 1) for "clValid" :

"euclidean", "correlation", and "manhattan".
2) for "NbClust" :
This must be one of: "euclidean", "maximum", "manhattan", "canberra", "binary", "minkowski"
additional non-conflicting arguments to "clValid" or "Nbclust"

# Value

. . .

1) for "clValid"

an object of class "clValid" (optimal = FALSE) or a dataframe of optimal values (optimal = TRUE)

2) for "NbClust"

a list of :

All.index, All.CriticalValues, Best.nc and Best.partition.

See the help pages of "clValid" (?clValid) and "NbClust" (?NbClust) for more details.

#### Author(s)

Subhadeep Das

# References

Brock, G., Pihur, V., Datta, S. and Datta, S. (2008) clValid: An R Package for Cluster Validation Journal of Statistical Software 25(4) http://www.jstatsoft.org/v25/i04

Charrad M., Ghazzali N., Boiteau V., Niknafs A. (2014). "NbClust: An R Package for Determining the Relevant Number of Clusters in a Data Set.", "Journal of Statistical Software, 61(6), 1-36.", "URL http://www.jstatsoft.org/v61/i06/".

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
    "H3k9ac"),
groupinfo = groupinfo,
name = multi_assay, mergetype = 2,
exclude = exclude, graph = FALSE)
name = int_PCA$int_PCA
data <- extract(name = name, PC = c(1:4),
groups = c("WE", "RE"), integrated = TRUE, rand = 600,
groupinfo = groupinfo_ext)
##### Using "clValid" ####
clusterstats <- cluster_parameters(name = data,
optimal = FALSE, n = 2:4, comparisonAlgorithm = "clValid",
distance = "euclidean", clusteringMethods = c("kmeans"),</pre>
```

#### create\_group

```
validationMethods = c("internal"))
```

create\_group Subsets an Assay dataframe into smaller groups

#### Description

This function subsests an user defined Assay into smaller groups according to user supplied instructions.

# Usage

```
create_group(name, group_names = NULL,
    grouping_factor = NULL,
    comparison = NULL,
    condition = NULL
    )
```

# Arguments

name	Name of the "MultiAssayExperiment"" object containing the assay data
group_names grouping_facto	A vector containing the user defined names of the groups to be created r
	name of the dataframe on which grouping will be done
comparison	A vector of comparison symbols such as >, <, ==, >=, <=,%in%, etc
condition	A vector of conditions corresponding to 'comparison'. "condition"" should be a vector or range of digits (e.g. $c(1,3,7,9)$ or 1:5) if%in% is chosen as comparison. Otherwise, a single digit should be chosen.

# Value

returns the group membership of individuals as a dataframe

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

## Examples

```
groupinfo <- create_group(name = multi_assay,
group_names = c("WE","RE","NE","IntE"),
grouping_factor = "CAGE",
comparison = c(">=","%in%","==","%in%"),
condition = c("25","1:5","0","6:24"))
```

head(groupinfo)

```
descriptor
```

# Description

descriptor() produces panels of boxplots displaying the distribution of various factors(e.g. ChIP-seq of H3k9ac histone modification) in percentage of cell lines.

choice = 1 output is collection of boxplots explaining the distribution of percentage of cell lines having or overlapping with the input factors (e.g. H3k9ac).

choice = 2 displays the distribution of percentage of cell lines having or overlapping with increasing number of factors for the selected group (e.g. WE or RE etc). If this option is selected, then a group name should be provided through choice2group.

#### Usage

descriptor(name, factors, groups, choice = 1, choice2group = NULL, title = NULL, groupinfo = NULL)

#### Arguments

name	Name of the "MultiAssayExperiment" object containing the assay data
factors	A vector containing the name of the Assays
groups	Name of all or subset of groups
choice	Numerical value between 1 and 2. See Description and vignettes for details of plots generated by this 2 choices
choice2group	Used when choice is 2. This argument allows a name of group.
title	Title of the plot
groupinfo	output of create_group() or similar object

#### Value

Displays various plots as per the combination of various input arguments.

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

```
descriptor(name = multi_assay,
factors = c(
 "H2az","H3k9ac","H3k4me1"),
groups = c("WE","RE","IntE","NE"),
choice = 1,
title = "Distribution of percentage of cell types overlapping
```

# extract

```
with various factors",
groupinfo = groupinfo_ext)
```

extract

extraction of projected cooordinates from "PCA" object.

# Description

funtion to extract projected coordinates of individuals on principal components.

#### Usage

extract(name, groups, integrated = TRUE, Assay = NULL, rand = NULL, PC = c(1,2,3,4), groupinfo = NULL)

# Arguments

name	name of the OMICs object.
groups	one or more groups created by or supplied through "create_group()"
integrated	whether to extract coordinates from integrated or individual assay.
Assay	name of the assay if integrated = FALSE
rand	number of rows to be extracted randomly.
PC	principal component numbers. The projected informaton on this PCs will be extracted.
groupinfo	<pre>same as integrate_variables()</pre>

# Value

a data frame of the extracted value.

# Author(s)

Subhadeep Das

# Examples

exclude <- list(0,c(1,9))

```
int_PCA <- integrate_pca(Assays = c("H2az",
"H3k9ac"),
groupinfo = groupinfo,
name = multi_assay, mergetype = 2,
exclude = exclude, graph = FALSE)
```

name = int\_PCA\$int\_PCA

```
data <- extract(name = name, PC = c(1:4),
groups = c("WE","RE"), integrated = TRUE, rand = 600,
groupinfo = groupinfo_ext)
```

extract\_assay

# Data extraction from "MultiAssayExperiment" object

## Description

Extraction of individuals according to user supplied group and Assay

#### Usage

```
extract_assay(name, Assay, groups, groupinfo = NULL,
addgroupnames = TRUE)
```

# Arguments

name	name of the "OMICs" object from which data is to be retrieved.
Assay	name of the "Assay" from which data is to be retrieved.
groups	name of the "groups" from which data is to be retrieved.
groupinfo	output of create_group() or similar object
addgroupnames	logical argument. If "TRUE", adds the group membership info of returned individuals

# Value

a data frame containing the intended information

# Author(s)

Subhadeep Das

# Examples

```
x <- extract_assay(name = multi_assay, Assay = "H2az",
groups = c("WE","RE"), groupinfo = groupinfo_ext)
```

#### х

16

integrate\_pca

#### Description

This function integrates multiple assays read by InputOMICs() into linear combinations using PCA. Following earlier analyses, user may want to exclude some variables/columns/cell lines which should be supplied through the "exclude" argument (see vignettes for more explanation)

PCA is done by the function PCA() from package "FactoMineR". Additional arguments may be supplied to PCA() through "...".

#### Usage

```
integrate_pca(Assays, name, exclude, mergetype = 1,
groupinfo = NULL, ...)
```

#### Arguments

Assays	A vector of names of an Assays read by "InputOMICs()"
name	Name of the "MultiAssayExperiment" object containing the assay data
exclude	A list that indicates which columns to exclude. See vignettes for details
mergetype	2 options :
	1 = subset of groups should be entered by user and PCA is run on this subset to integrate Assays into PCs
	2 = PCA is run on all groups
groupinfo	same as plot density()
groupino	same as plot_density()
	aditional arguments supplied to function "PCA()" from package "FactoMineR"

# Details

The "mergetype" argument lets user decide whether to combine all groups or a subset of groups created by "create\_group". If mergetype is set to 1, "integrateAssays" asks for the name of groups which should be a character or charagter vector (see vignettes for more details).

# Value

"integrateAssays()" returns a list containing the start and end column of each Assay as ordered in the "Assays" argument.

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

#### Examples

```
int_PCA <- integrate_pca(Assays = c("H2az",
"H3k9ac"), name = multi_assay, mergetype = 2,
exclude = list(0,c(1,9)), graph = FALSE,
groupinfo = groupinfo_ext)
```

integrate\_variables Integration of an experiment/ Assay done on many Cell lines/ time points into linear combinations by PCA.

# Description

If an assay (e.g. ChIP-seq for the histone modification H3k9ac) is done on multiple cells/conditions/treatment/time, it might be, sometimes, necessary to integrate or combine them. Such combination may be done by several techniques like Principal Component Analysis (PCA) or Factor Analysis (FA). This function integrates an assay experimented on multiple cells or conditions into many linear combinations using PCA.

# Usage

```
integrate_variables(Assays, name, groups, groupinfo = NULL, ...)
```

#### Arguments

Assays	A vector of names of an assays
name	Name of the "MultiAssayExperiment" object containing the assay data
groups	Name of all or subset of groups
groupinfo	output of create_group or similar object containing group information
	Additional arguments supplied to "PCA()" imported from "FactoMineR"

#### Value

an object of class "PCA"

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Examples

```
PCAlist <- integrate_variables(Assays = c("H2az","H3k4me1",
"H3k9ac"), name = multi_assay,
groups = c("WE","RE"), groupinfo = groupinfo_ext,
scale.unit = FALSE, graph = FALSE)
```

18

intersect

# Description

intersects a factor file and an annotation file

#### Usage

intersect(fact, anno, ...)

# Arguments

fact	Full path of the directory containing only the bed (.bed) files corresponding to several cell lines of the specified factor.
anno	Full path of the annotation file (e.g. gene, TSS, exon etc.)
	$additional \ arguments \ received \ from \ "prepareDataset()" \ and \ suppliet \ to \ "R_bedtools\_intersect()"$

# Value

A list containing the intersections of the files (paths of which are supplied through the arguments).

# Author(s)

Subhadeep Das

```
anno <- system.file("extdata/annotation2/TSS_groups.bed",
package = "OMICsPCAdata")
fact <- system.file("extdata/factors2/demofactor",
package = "OMICsPCAdata")
Cells <- intersect(fact = fact, anno = anno)
Cells
```

merge\_cells

#### Description

This function takes the output list of the function "intersect" and stores the intersected value of the factor and annotation files into a dataframe. The values corresponding to the entries in annotation file represents the corresponding value of in each cell line. If an entry is not found in a factor file, its corresponding value will be 0.

#### Usage

merge\_cells(list, Cells)

#### Arguments

list	Full path of the file containing name of the Annotations (e.g. gene) in a single column
Cells	output list of function "integrate()"

# Value

A dataframe wich rows corresponding to each entry in annotation file and columns corresponding to the intersected value with the corresponding annotation in each cell line.

#### Author(s)

Subhadeep Das

```
anno <- system.file("extdata/annotation2/TSS_groups.bed",
package = "OMICsPCAdata")
fact <- system.file("extdata/factors2/demofactor",
package = "OMICsPCAdata")
Cells <- intersect(fact = fact, anno = anno)
list <- system.file("extdata/annotation2/TSS_list",
package = "OMICsPCAdata")
merged_Cells <- merge_cells(list = list, Cells = Cells)
head(merged_Cells)
```

plot\_density

# Description

This function displays density of individuals

# Usage

```
plot_density(name, Assay, PC = 1, groups,
groupinfo = NULL, ...)
```

# Arguments

name	Name of the "PCA" object containing the assay data
Assay	Name of an Assay
PC	A number corresponding to the principal component on which density is to be calculated
groups	A vector of names or subset of names of groups
groupinfo	same as integrate_variables()
	additional arguments of base function "geom_density" from package "ggplot2"

# Value

a "gg" "ggplot" object

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

```
PCAlist <- integrate_variables(Assays = c("H2az","H3k4me1",
"H3k9ac"), name = multi_assay,
groups = c("WE","RE"), groupinfo = groupinfo_ext,
scale.unit = FALSE, graph = FALSE)
densityplot <- plot_density(name = PCAlist,
Assay = "H2az", groupinfo = groupinfo_ext,
PC = 1, groups = c("WE","RE"),
adjust = 1)
```

plot\_density\_3D

#### Description

This function displays density of individuals from various groups created by create\_group(). This wraps "persp" from package "graphics" and thus takes additional graphical input shown in example.

# Usage

```
plot_density_3D(name, Assay, group, PC1 = 1,
PC2 = 2, static = FALSE,
gridsize = 100, groupinfo = NULL, ...)
```

# Arguments

name	Name of the "PCA" object containing the assay data
Assay	Name of an Assay
group	Names of a group
PC1, PC2	Numbers corresponding to the principal components on which density is to be calculated
static	Logical if TRUE a static plot is generated. default = FALSE
gridsize	A number used in kernel smoothing. default is 100
groupinfo	Same as integrate_variables()
	additional arguments passed to base function "persp" from package "graphics"

#### Details

2D density is calculated using the "kde2d"" function from package "MASS"" which use kernel density estimation (KDE) to calculate density of 2D data. If the variance on either or both of the PCs are 0, the KDE can't be calculated.

# Value

Displays 3D density plots of PCs

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# plot\_integrated\_density

# Examples

```
PCAlist <- integrate_variables(Assays = c("H2az"),
name = multi_assay,
groups = c("WE","RE"), groupinfo = groupinfo_ext,
scale.unit = FALSE, graph = FALSE)
plot_density_3D(name = PCAlist, Assay = "H2az",
group = "WE", PC1 = 1, PC2 = 2, grid_size = 100,
static = FALSE, groupinfo = groupinfo_ext)
```

```
plot_integrated_density
```

Visualization of Density of various groups on Principal components of integrated assays

# Description

This function works similarly as "plot\_density()". See the man page of "plot\_density()" for details.

#### Usage

```
plot_integrated_density(name, PC = 1, groups,
groupinfo = NULL, ...)
```

#### Arguments

name	Name of the integrated "PCA"" object
PC	A number corresponding to the principal component on which density is to be calculated. default = 1
groups	A vector of names or subset of names of groups
groupinfo	same as plot_density()
	additional arguments allowed to base function "geom_density" of package "gg-plot2"

# Value

a "gg" "ggplot" object

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Examples

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
    "H3k9ac"), name = multi_assay, mergetype = 2,
exclude = exclude, groupinfo = groupinfo_ext,
,graph = FALSE)
name = int_PCA$int_PCA
densityplot <- plot_integrated_density(name = name, PC = 1,
groups = c("WE", "RE", "IntE", "NE"), groupinfo = groupinfo_ext)
# additional graphical functions (e.g. xlim, ylim, theme) may be
#added with densityplot (see section VIII. Density analysis)
densityplot</pre>
```

plot\_integrated\_density\_3D

Visualization of 3D Density of various groups on Principal components of multiple Assays

# Description

works similarly as "plot\_density\_3D". See vignettes and man page of "plot\_density\_3D" for more details.

#### Usage

```
plot_integrated_density_3D(name, PC1 = 1, PC2 = 2, group,
gridsize = 100, static = FALSE, groupinfo = NULL, ...)
```

# Arguments

name	Name of the integrated "PCA" object
PC1, PC2	Numbers corresponding to the principal components on which density is to be calculated
group	Names of a group
gridsize	A number used in kernel smoothing. default is 100
static	Logical if TRUE a static plot is generated. default = FALSE
groupinfo	same as integrate_variables()
	additional arguments allowed to base function "persp" of package "graphics"

24

#### prepare\_dataset

#### Details

2D density is calculated using the "kde2d"" function from package "MASS"" which use kernel density estimation (KDE) to calculate density of 2D data. If the variance on either or both of the PCs are 0, the KDE can't be calculated.

#### Value

Displays 3D density plots.

#### Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

#### Examples

```
exclude <- list(0,c(1,9))
int_PCA <- integrate_pca(Assays = c("H2az",
"H3k9ac"), name = multi_assay, mergetype = 2,
exclude = exclude, groupinfo = groupinfo_ext,
,graph = FALSE)</pre>
```

```
name = int_PCA$int_PCA
```

```
plot_integrated_density_3D(name = name, PC1 = 1, PC2 = 2,
group = c("WE","RE"), gridsize = 100, static = FALSE,
groupinfo = groupinfo_ext)
```

prepare\_dataset Intersects and merge data from multiple experiments into a dataframe

#### Description

This function intersects feature files (e.g. ChIP-seq, RNA seq) with annotation files(e.g.exons, genes) and represents the intersected values of each annotation from all cell lines in a data frame.

#### Usage

```
prepare_dataset(factdir, annofile, annolist, ...)
```

# Arguments

factdir	Full path of directory containing (only) feature (e.g ChIP-seq, CAGE) files in bed format
annofile	Full path of the annotation file (e.g. TSS, exon, gene) in bed format
annolist	Full path of the file containing name of the annotations (e.g. name of exons)
• • •	additional arguments to pass to "R_bedtools_intersect()" through "intersect()"

# Details

If an annotation (e.g. gene) does not present in an assay file (e.g ChIP-seq of H3k9ac in the Cell Gm12878), this function puts a 0 as its value.

# Value

a dataframe of intersected values.

# Author(s)

Subhadeep Das <subhadeep1024@gmail.com>

# Index

```
* density-based clustering
    cluster, 8
* kmeans clustering
    cluster,8
analyse_individuals, 2
analyse_integrated_individuals, 3
analyse_integrated_variables, 5
analyse_variables, 6
chart_correlation, 7
cluster, 8
\texttt{cluster\_boxplot}, 10
cluster_parameters, 11
create\_group, 13
descriptor, 14
extract, 15
extract_assay, 16
integrate_pca, 17
integrate_variables, 18
intersect, 19
\texttt{merge\_cells}, \underline{20}
plot_density, 21
plot_density_3D, 22
plot_integrated_density, 23
plot_integrated_density_3D, 24
prepare_dataset, 25
```